

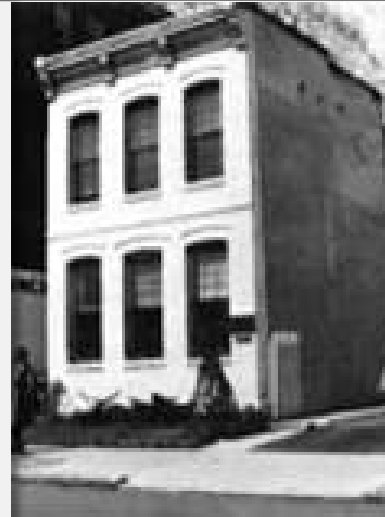
Phenotypic Characterization and Quality Control of Cells from Bacteria to Human Cells

Barry R. Bochner, Ph.D.
CEO & CSO, Biolog, Inc.
October 29, 2015



About ATCC

- Founded in 1925, ATCC is a non-profit organization with headquarters in Manassas, VA
- World's premiere biological materials resource and standards development organization
- ATCC collaborates with and supports the scientific community with industry-standard biological products and innovative solutions
- Strong team of 400+ employees; over one-third with advanced degrees



Established partner to global researchers and scientists



Certification and Accreditation

ISO 9001:2008 Certification for quality management system

- Demonstrates commitment to quality products, customer service, and continued improvement



ISO 13485:2003 Certification for the design, development, production, testing, and distribution of medical devices

- Applies to synthetic molecular standards, the HIV surveillance kit, and other diagnostic and research kits



ISO Guide 34:2009 accreditation for production

- Applies to Certified Reference Materials (CRMs)



ISO/IEC 17025:2005 accreditation for testing

- Applies to all ATCC cultures, derivatives, and bioproducts tested in our laboratories



Highly Characterized Microbial Reference Strains

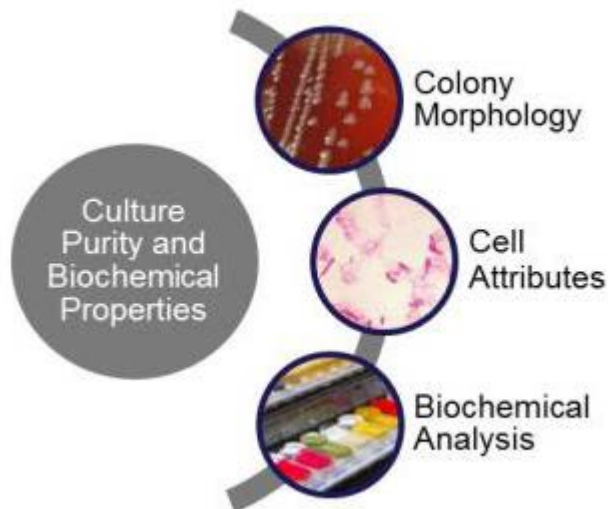


ATCC utilizes both classical and modern techniques

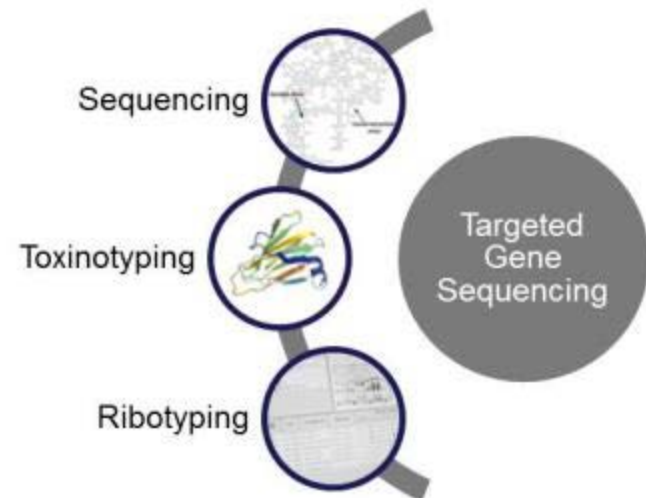
- Phenotypic analysis
- Genotypic analysis
- Proteotypic analysis
- Functional analysis

No single method of identification is sufficient

Phenotypic testing



Genotypic testing



Phenotypic Characterization and QC of Cells

From Bacteria to Human Cells



Barry Bochner, Biolog, Inc., bbochner@biolog.com





Part One:

Phenotypic Characterization of Cells

Tremendous Advances in Phenotyping Technology

Corynebacterium jeikeium:



Arcanobacterium haemolyticum:



Actinomyces pyogenes:



**1970s
Technology**

20 tests



**21st Century
Technology**

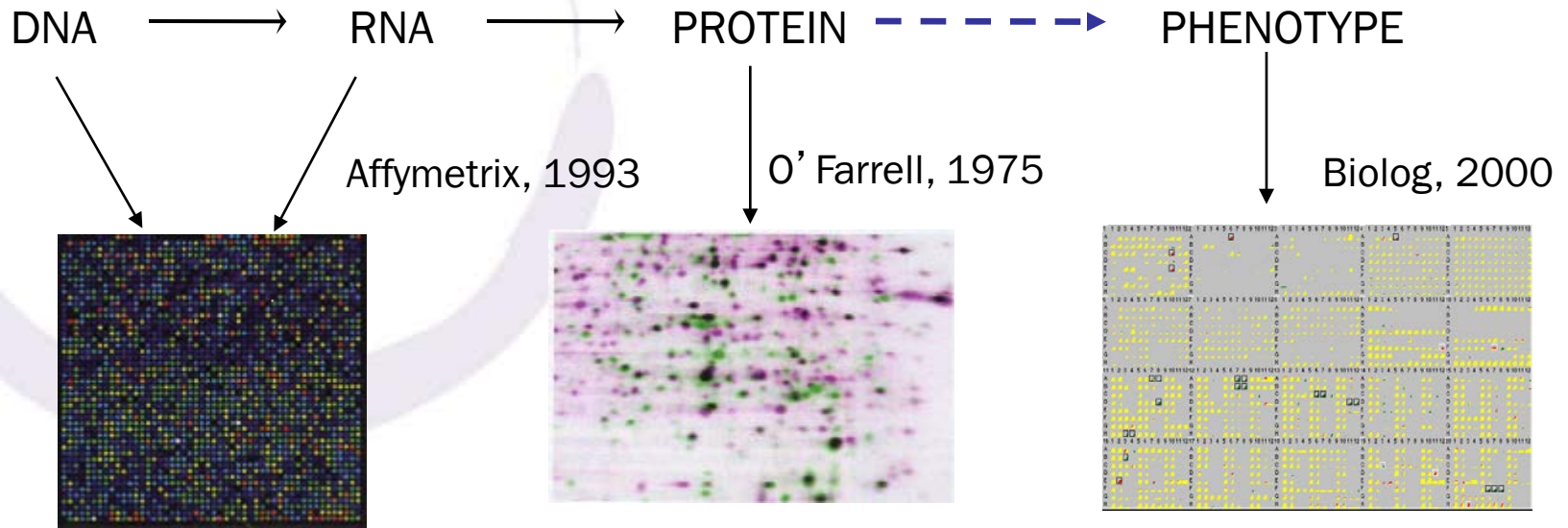
2,000 tests



Tremendous Advances in Phenotyping Technology

- Hundred fold expansion in tests
- Much broader range of tests
- Single dye, single color chemistry
- Bacteria
- Yeast
- Filamentous fungi
- Algae
- Human and other Animal Cells
- Kinetic phenotypes

Phenomics = High Content Cell Phenotyping



Molecular Analyses

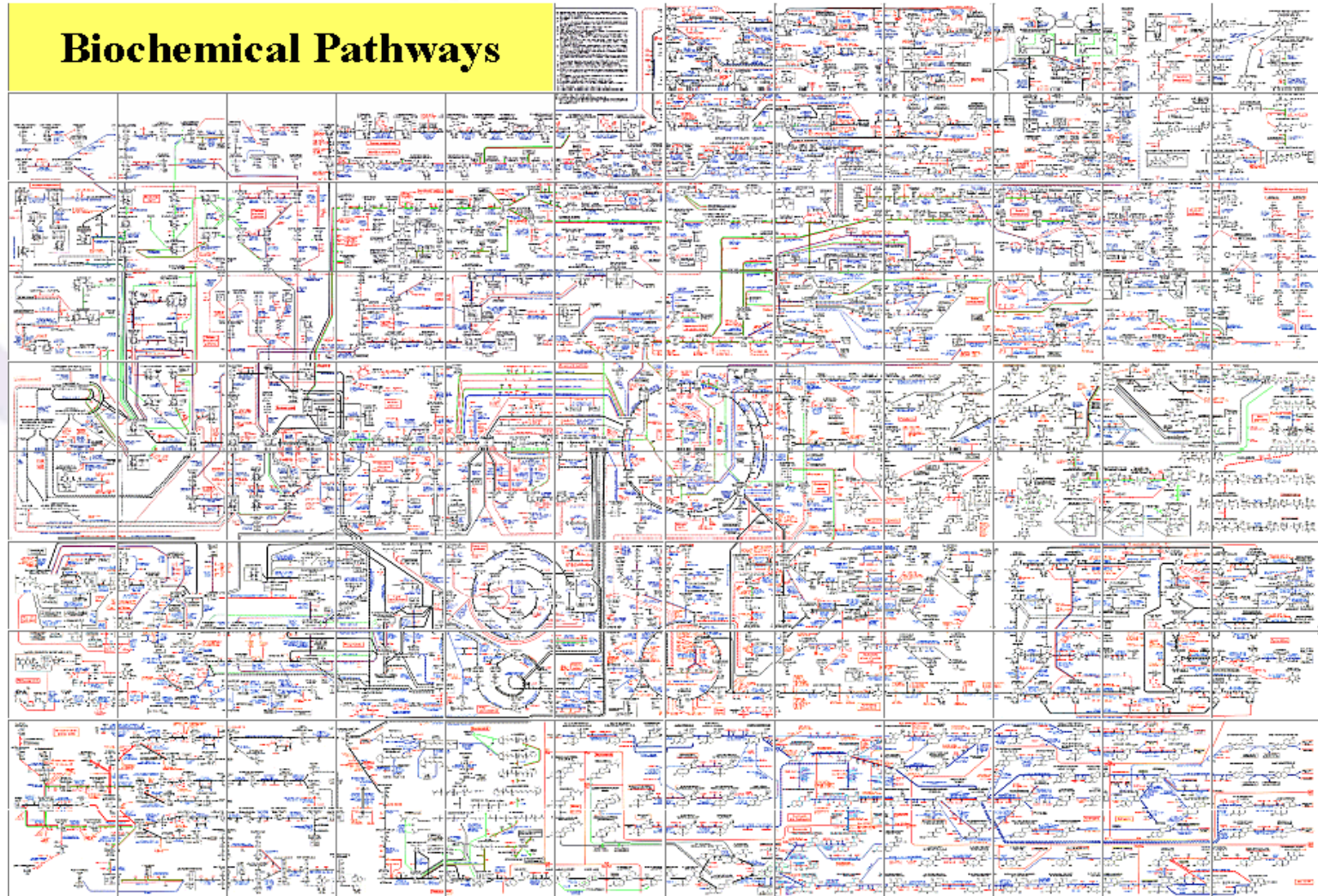
Cellular Analysis

Transcriptomics

Proteomics

Phenomics

Complex Metabolic Circuitry of Cells





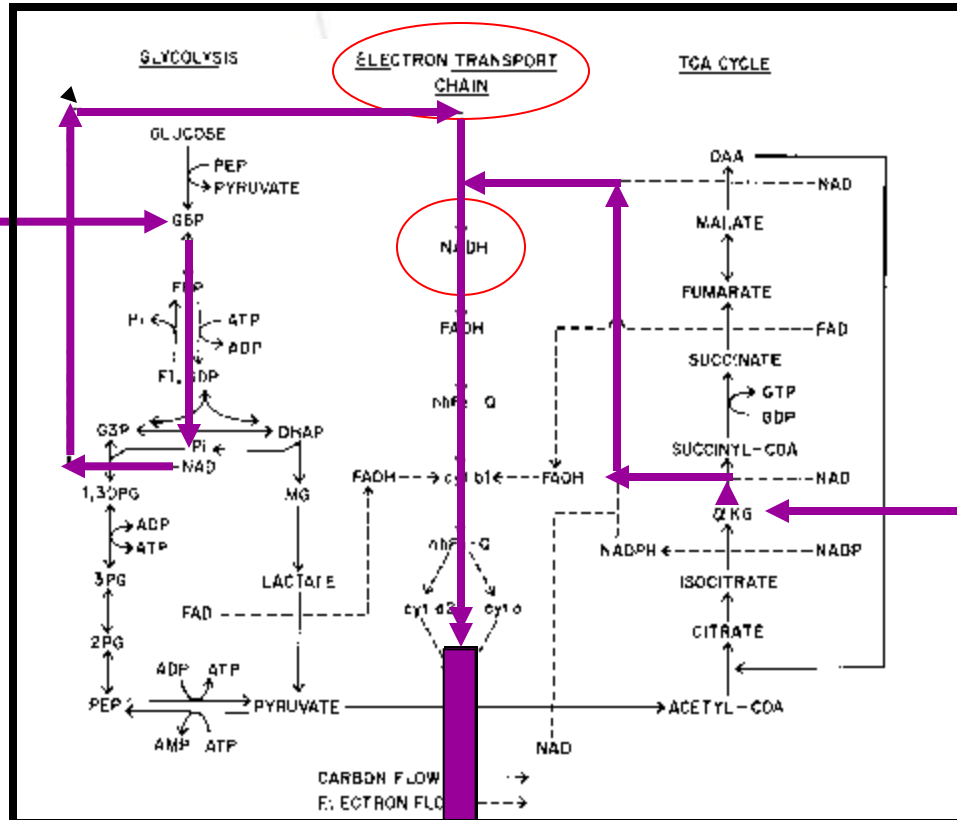
Assay Principal:

Colorimetric Analysis of Energy Production

Metabolism of C-sources Produces an Electron Flow

fructose

Testing sub-circuits within the cell's master circuitry



glutamine

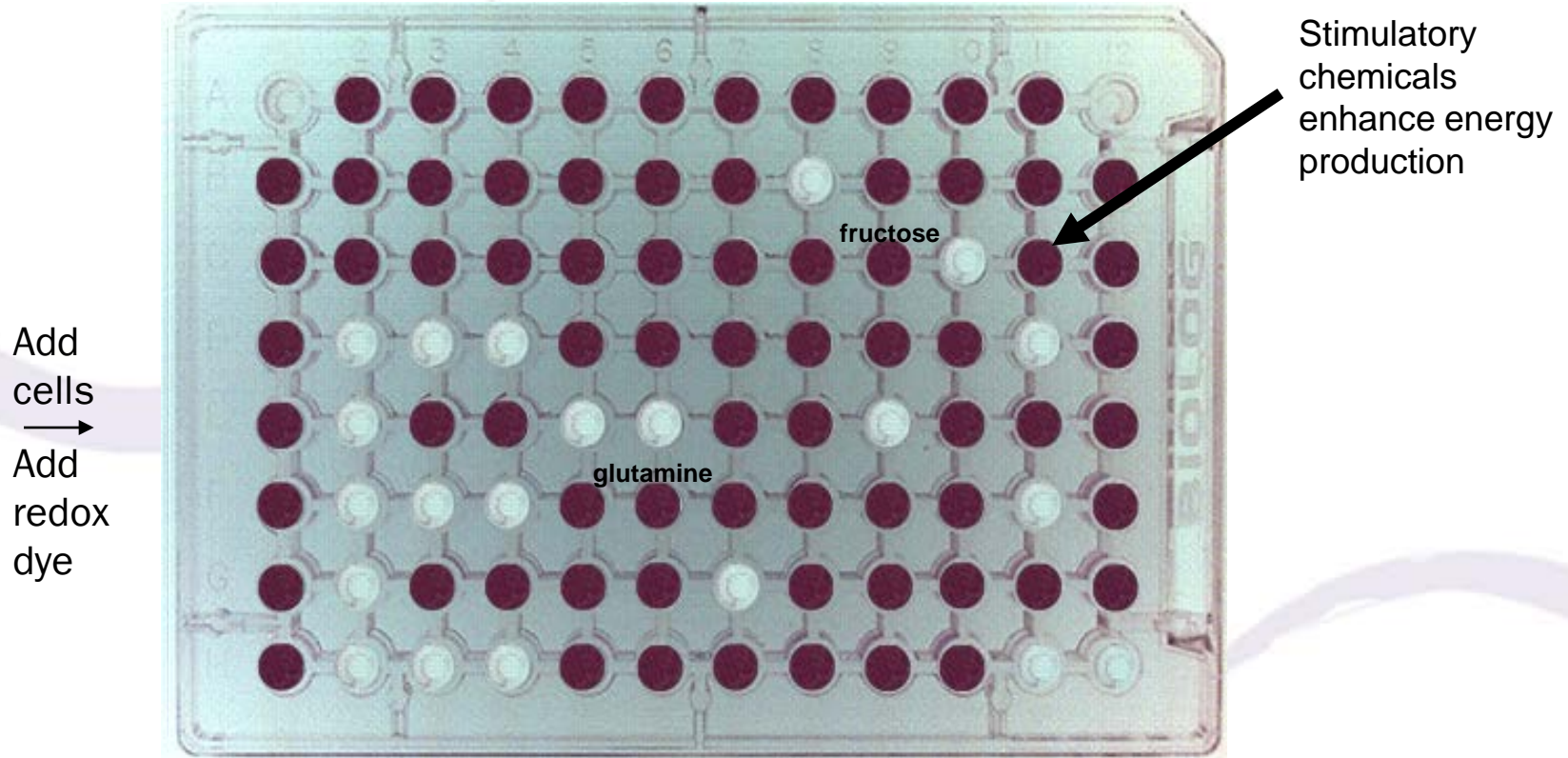
FIG. 3. The pathways of central metabolism in *E. coli* and *S. typhimurium*. The following abbreviations are used: glucose-6-P (G6P), fructose-6-P (F6P), fructose-1,6-diP (F1,6DP), glyceraldehyde-3-P (G3P), 1,3-diP-glycerate (1,3DPG), 3-P-glycerate (3PG), 2-P-glycerate (2PG), P-enolpyruvate (PEP), dihydroxyacetone-P (DHAP), methyl glyoxal (MG), non-heme iron-sulfur center (ahFe-S), cytochrome *Q* complex (ahFe-Q), cytochrome (*cyt*), oxaloacetate (OAA), and α -ketoglutarate (α KG).

Redox Dye

BiOLOG

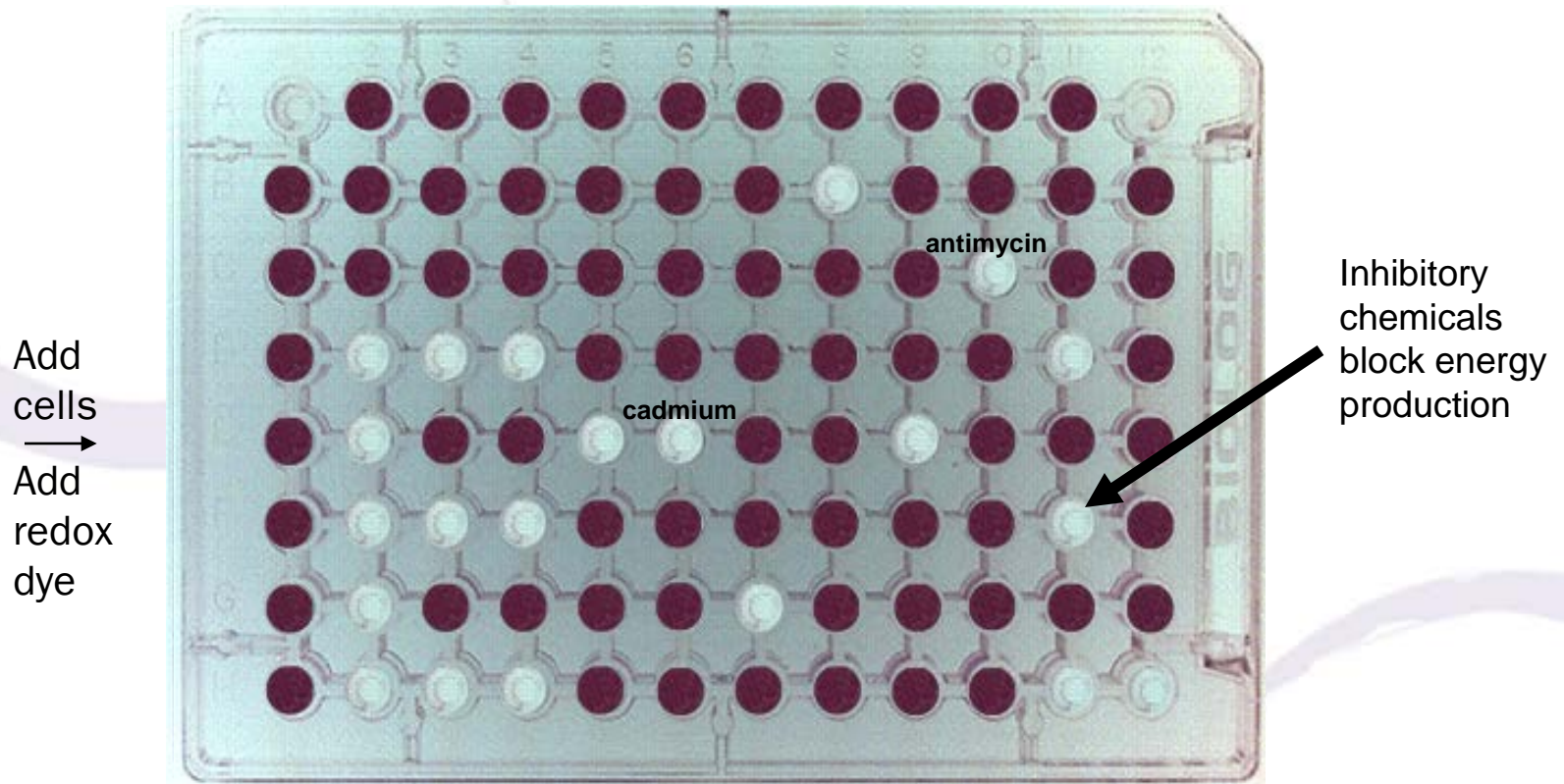
Redox Chemistry Measures Cell Energetics

Microplate containing a negative control well and 95 different carbon substrates



Wells contain different tests and measure different pathway activities and phenotypes of cells

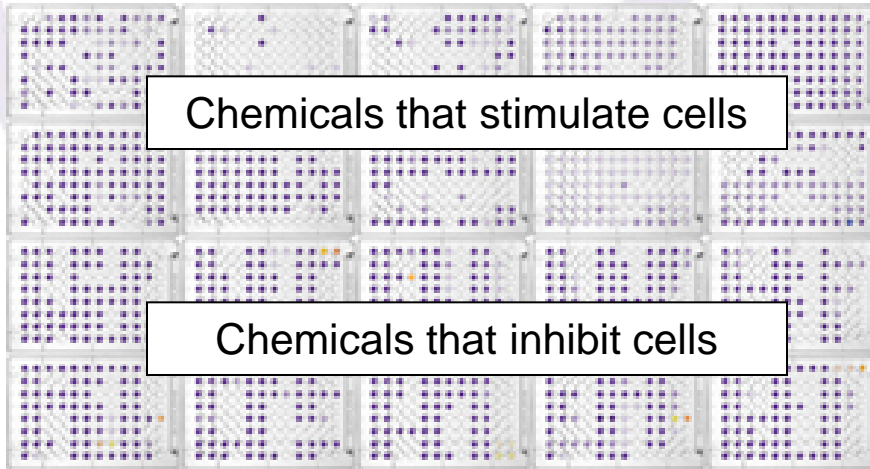
Redox Chemistry Measures Cell Energetics



Wells contain different tests and measure different pathway activities and phenotypes of cells

2 Components of the Phenotyping Assay Platform

Phenotype MicroArrays™



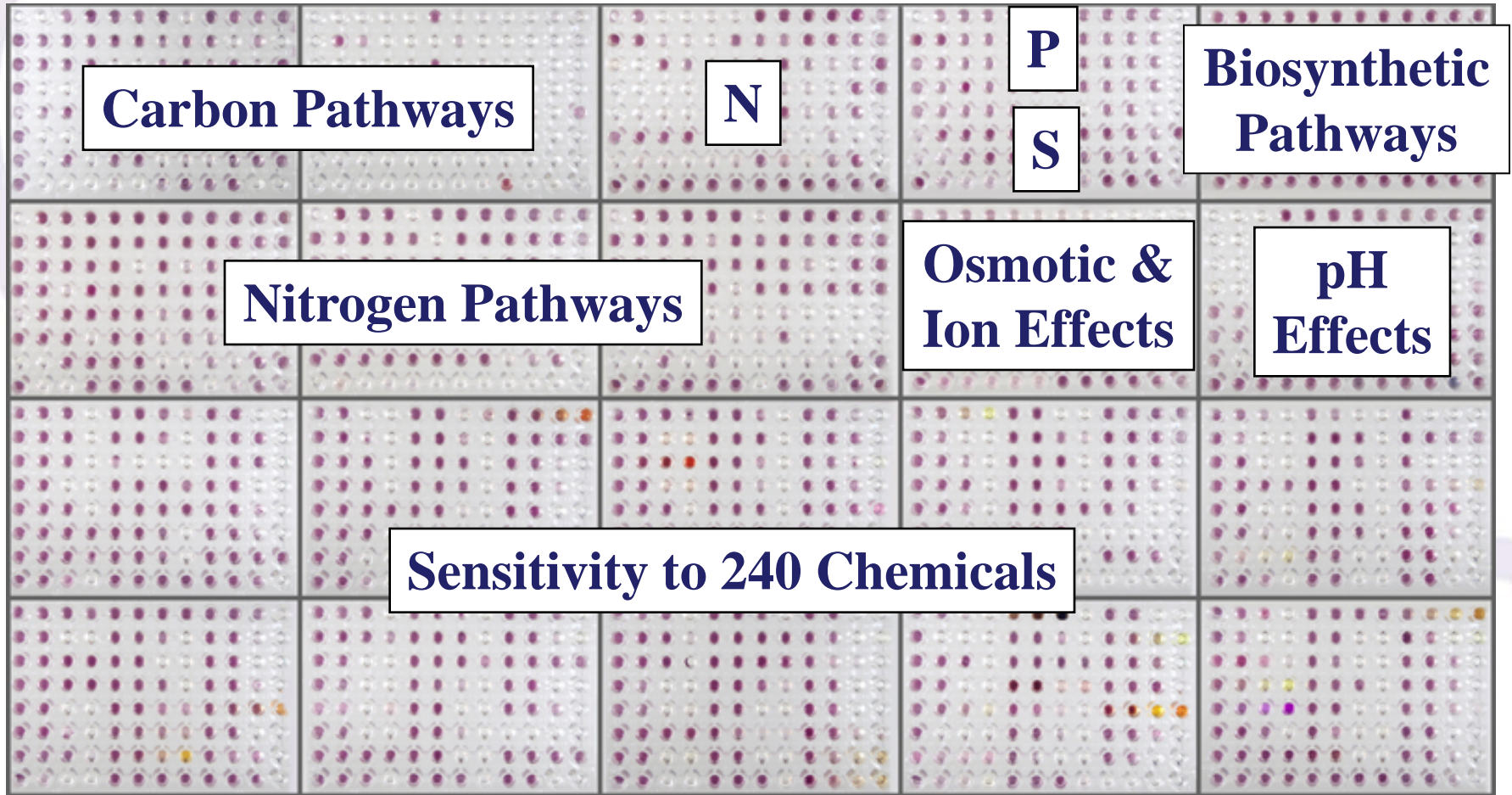
colorimetric cell assays in 96-well microplates

OmniLog™ Incubator/Reader



incubation and recording of data in the OmniLog

PM Platform - ~2,000 Phenotypic Assays



PM Assays are Easy to Run



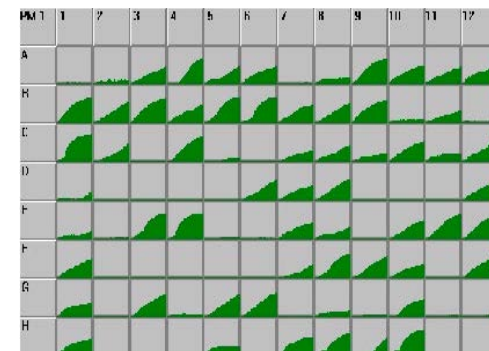
Assays Initiated by
adding cells to wells

100 μ l per well



OmniLog PM System

Holds 50 microplates at a
set temperature
and measures color formation
at 15-minute intervals

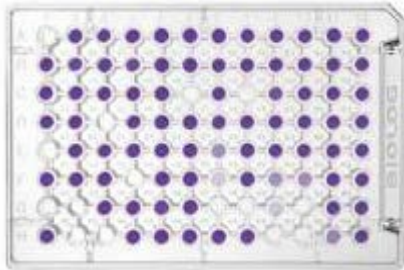


Kinetic assay readout
for up to 5,000 wells

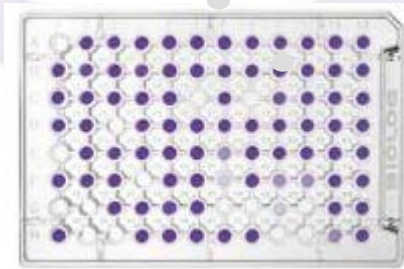
CVs typically < 10%

PM Platform - Comparing Two Cell Lines

Add cell **A**



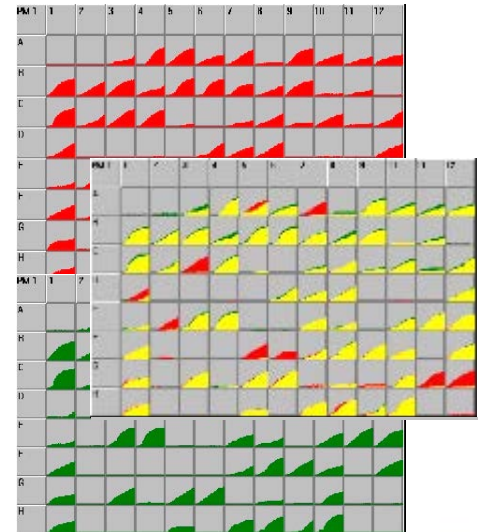
Add cell **B**



PM Pattern

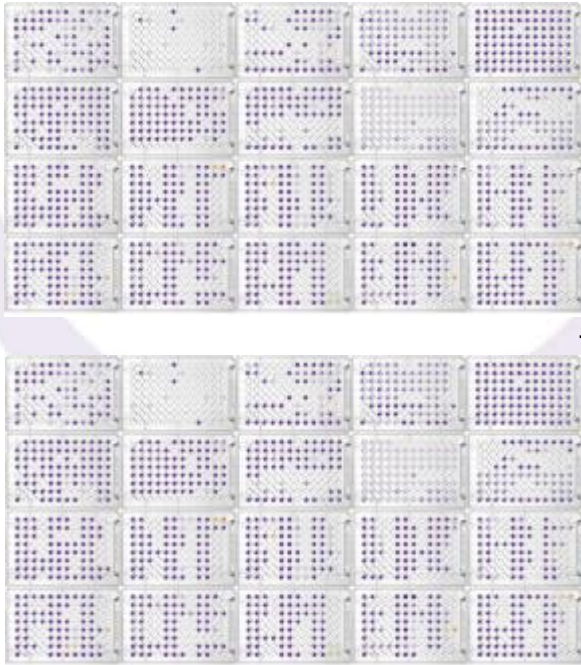


OmniLog PM System



PM Kinetic Result

PM Platform – Comparing Two Cell Lines



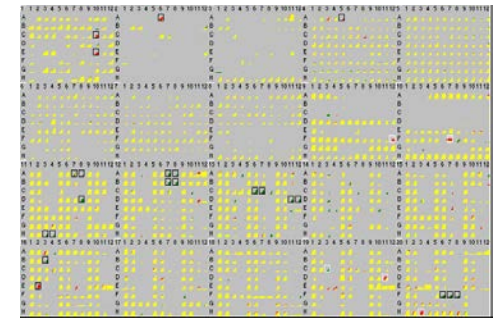
PM Pattern

1 hr



OmniLog PM System

Automatic



PM Kinetic Result

24-48 hr

Examples of Phenomic Applications

- Profiling metabolism and chemical sensitivities of a cell
- Comparing properties of pathogenic vs non-pathogenic strains
- Determining the function(s) of a gene
- Analyzing environmental effects on cell phenotypes
- Optimizing production of a cell product – e.g. a toxin

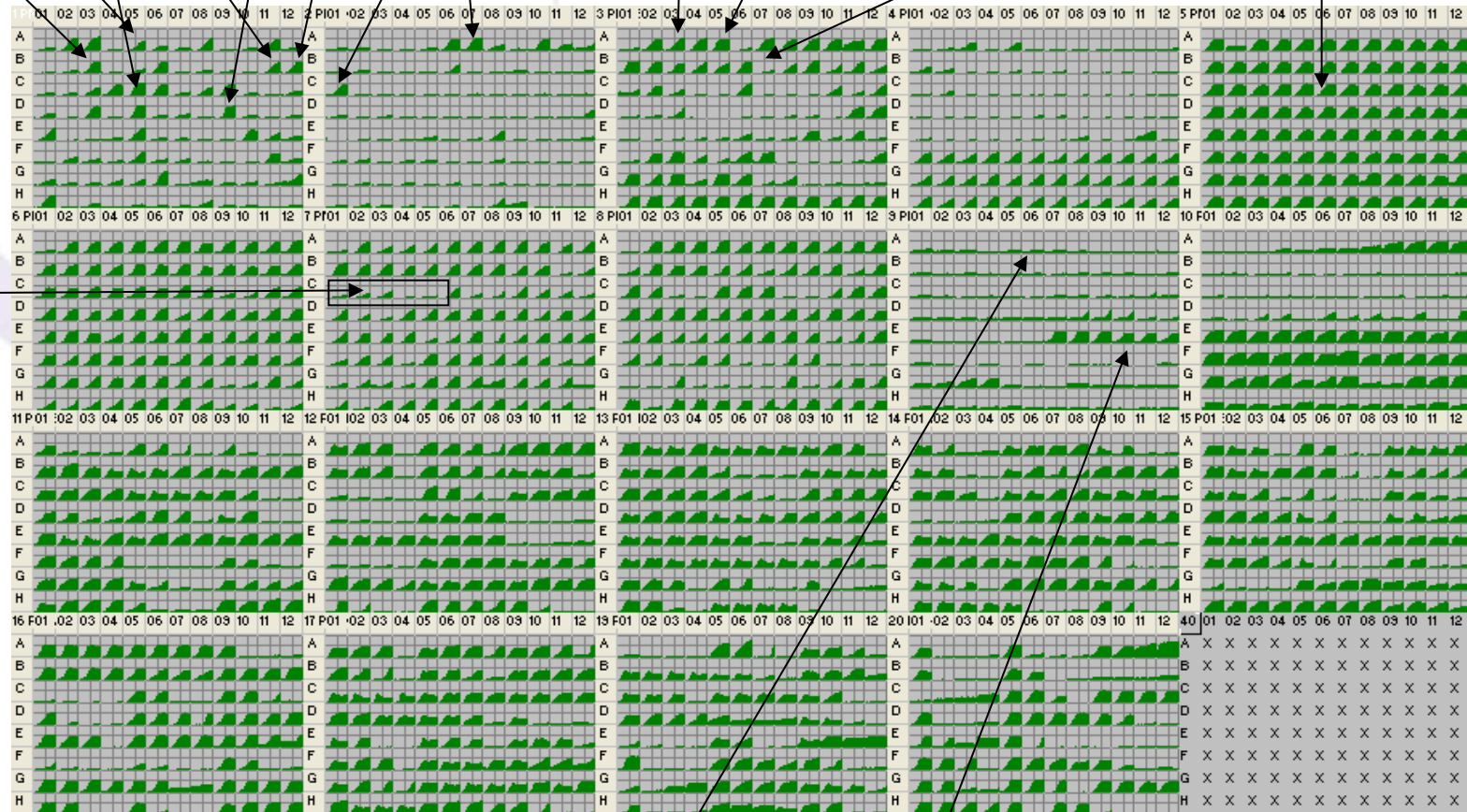
PM Analysis of *Streptomyces coelicolor*

succinate mannitol glutamate gelatin
glycerol tweens lactose gentiobiose

nitrite, urea
most amino acids (not met)

prototrophic

no met peptides



osmotically sensitive except to urea

PM Analysis of *Mycobacterium smegmatis* MC²-155

sorbitol, adonitol, xylitol, erythritol
glycerol, inositol, mannitol, arabitol

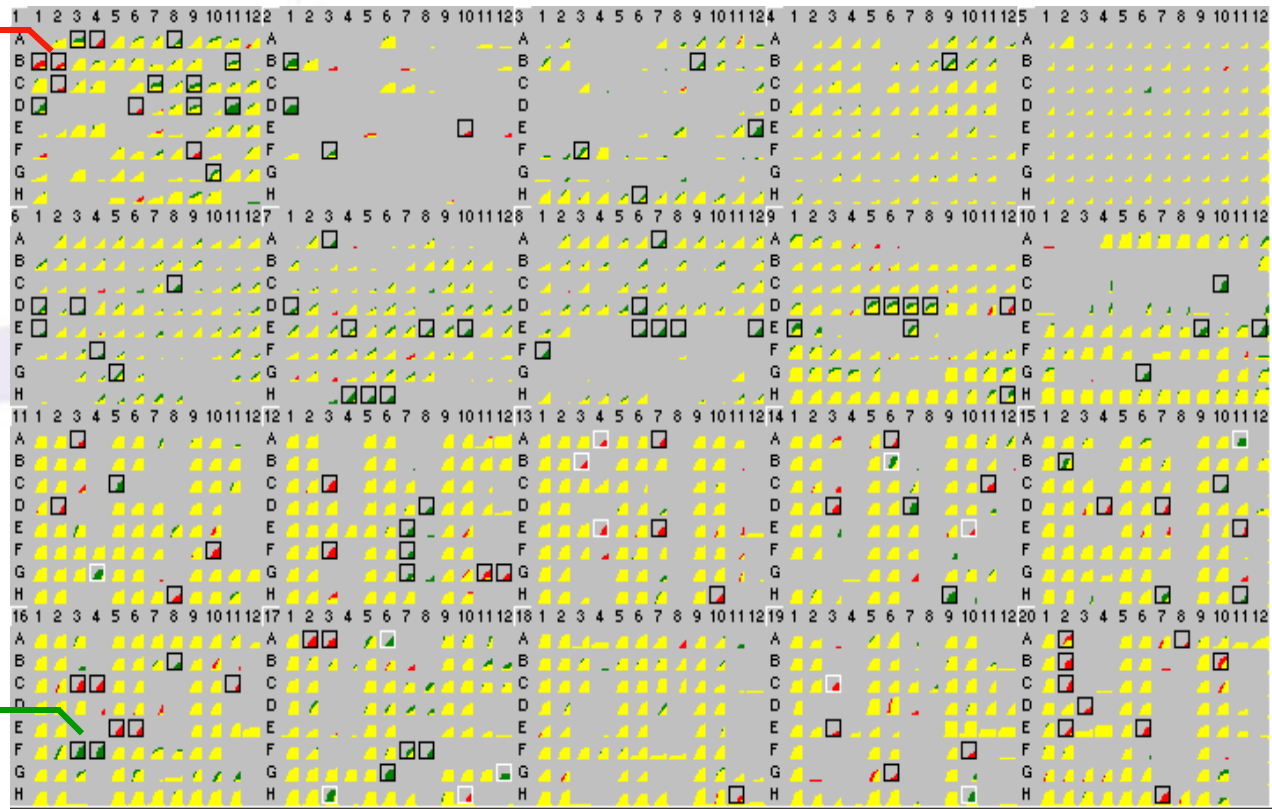
Also – glucose, trehalose, glutamate, serine, alanine
gluconate, acetate, acetoacetate, tween40 and others



Comparing Two E. coli Strains:

Pathogenic (**0157**) vs non-Pathogenic (**MG1655**) E. coli

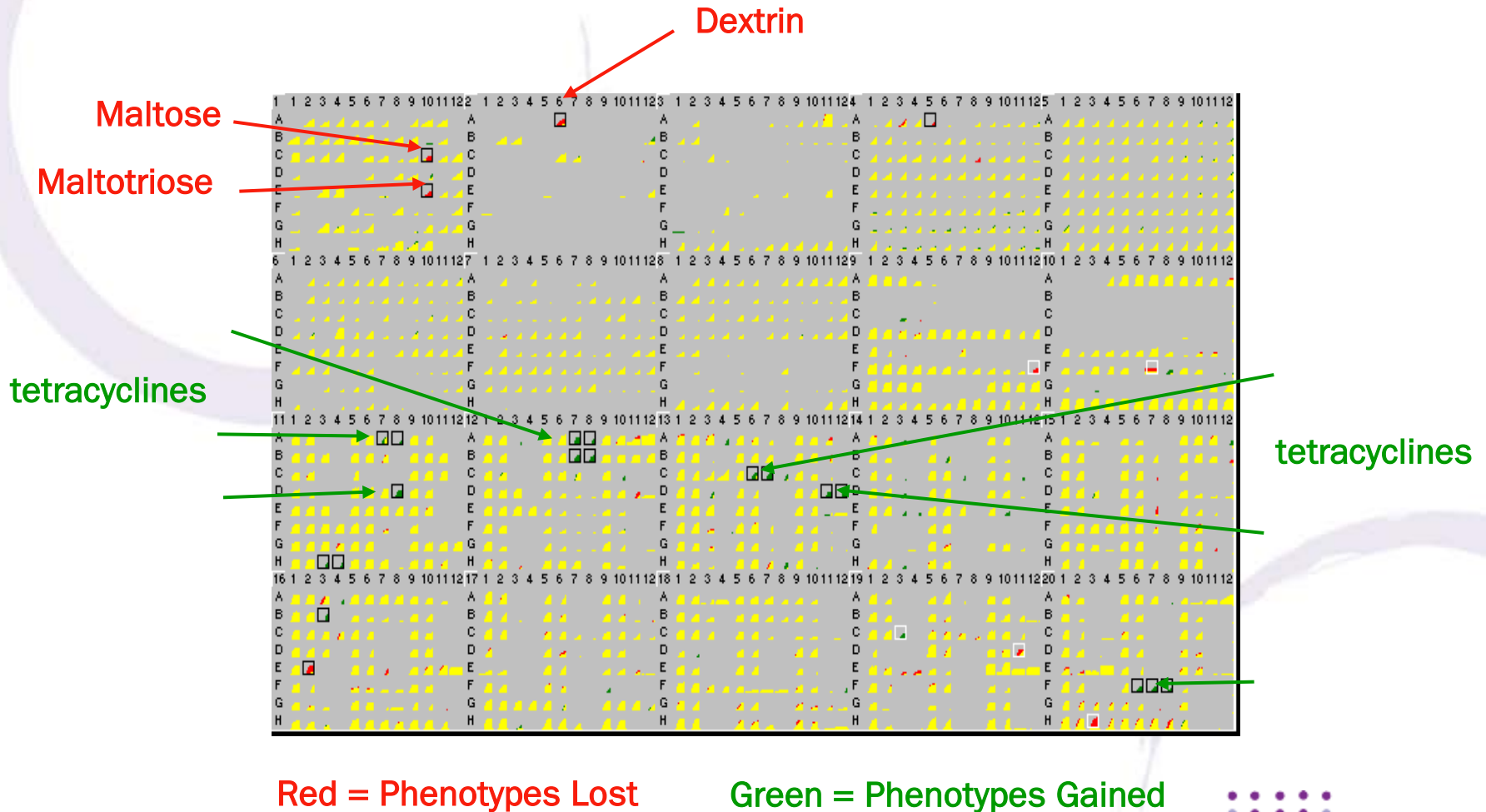
sorbitol
negative



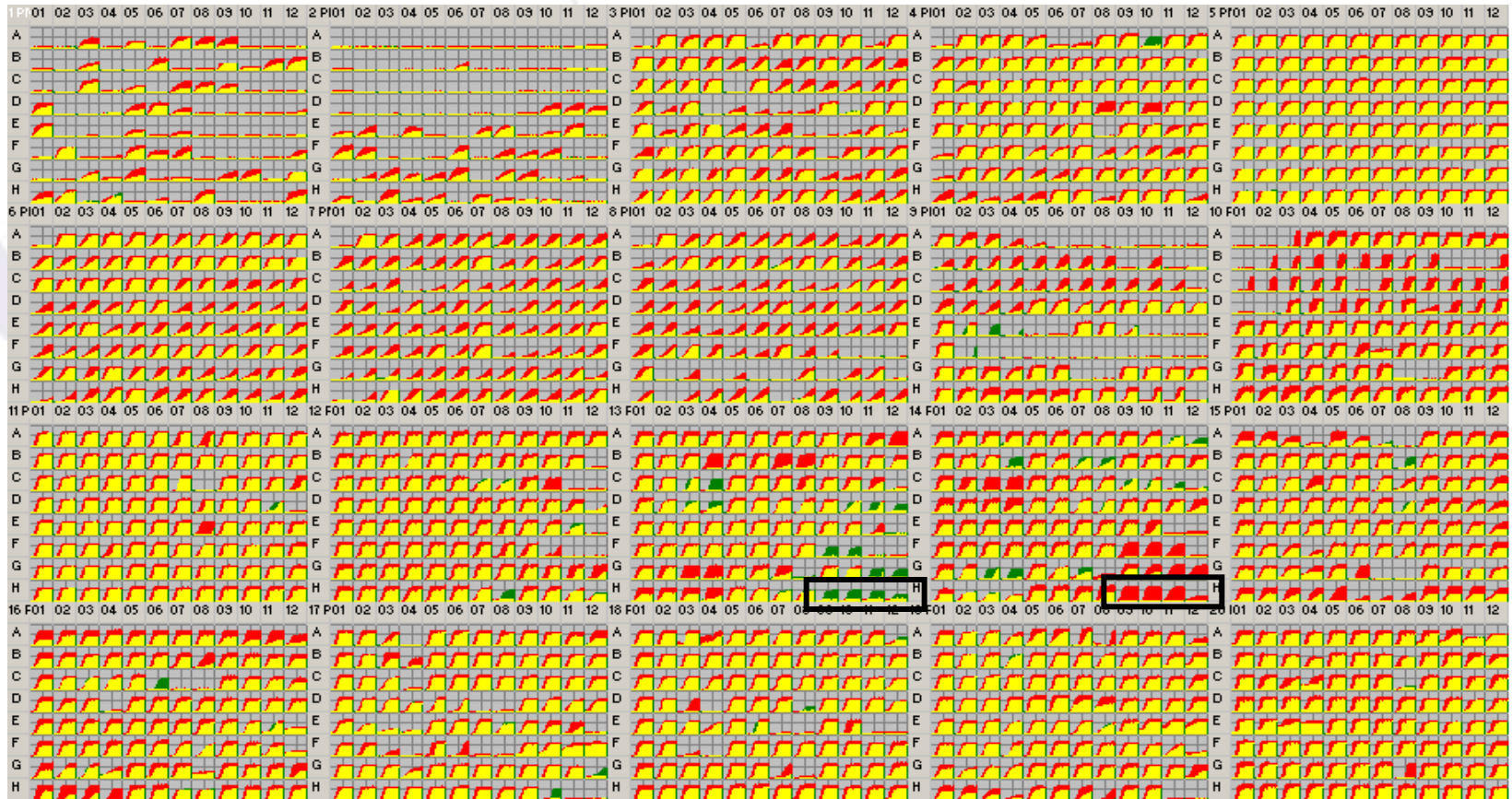
tellurite
resistant

Comparing Wild Type vs Mutant:

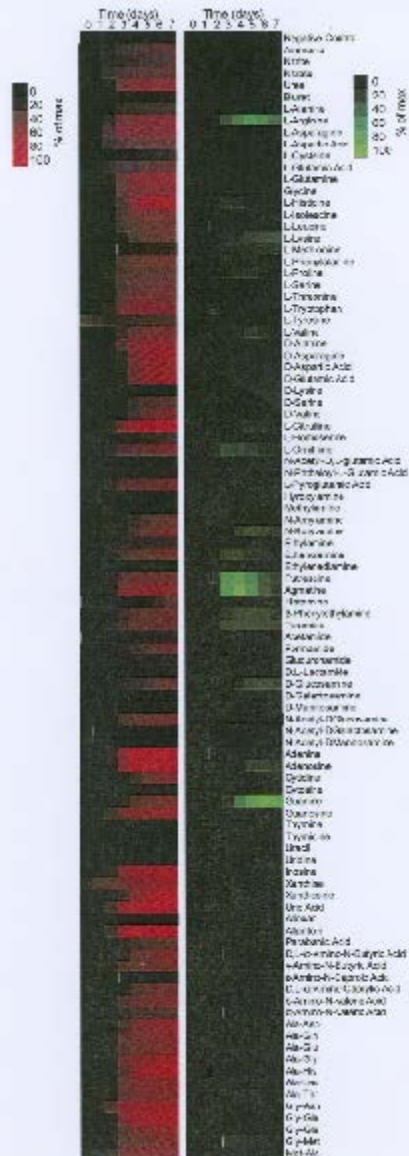
E. coli **malF::Tn10** vs **MG1655**



Comparing *Psd.aeruginosa* 8512 at 26 vs 36 C.



Culture Conditions Inducing Toxin Synthesis



Culture conditions inducing synthesis of a trichothecene mycotoxin in the wheat pathogen, *Fusarium graminearum*. A special strain was constructed with a toxin gene promoter fused to GFP

Induction was highest with arginine, putrescine, agmatine, and guanine as nitrogen sources.

D. Gardiner et al – Fungal Gen & Biol (2009)

Phenotyping Human Cells:

**Analyzing Human Cell Metabolism,
Human Genes, Human Disorders, and
Effect of Drugs**

~1500 Assays and Culture Media for Mammalian Cells



Sugars, amino acids, lipids, peptides as C-source

Ions & Trace
Elements

Amino acids, peptides as N-source

Cytotoxic cancer drugs

Plus 3 panels with hormones,
cytokines and other bioactives

Harry Eagle, JBC 1958

The Utilization of Carbohydrates by Human Cell Cultures

HARRY EAGLE, STANLEY BARBAN, MINA LEVY, AND HENRY O. SCHELES

From the Section on Experimental Therapeutics, Laboratory of Infectious Diseases, National Institute of Allergy and Infectious Diseases, National Institutes of Health, Public Health Service, United States Department of Health, Education, and Welfare, Bethesda, Maryland

(Received for publication, April 21, 1958)

TABLE I
Ability of a variety of carbohydrates* and related compounds to substitute for D-glucose in cell cultures

The compounds were tested at varying levels (0.5 to 50 mM) in a glucose-free but otherwise "complete" growth medium supplemented with dialyzed serum. The letters in parentheses after each compound indicate the cell lines against which it was tested.

Compounds which did not support growth	Compounds which grew slightly poorly, but less than D-glucose	Compounds permitting growth at rate comparable with D-glucose
Lactose (H, E, J, T)	Cellulose (L, G, H, K, M, T)	Trehalose (C, G, H, K, M, T)
Sucrose (H, L, T)	Mellician (G, J, H, K, M, T)	Tetraose (L, G, H, K, M, T)
D-Allose (G, T)	Sorbitol (all)	D-Fructose (all)
D-Alerose (G, T)	D-Triose (G, T)	D-Galactose (G, G, H, K, M, T)†
D-Galacturonic acid (H, T)	D-Xylose (all)	D-Glucose (all)
D-Gluconic acid (H, T)	D-Ribose (G, H, K)‡	D-Glucose 1-P, (H, K, M)
Methyl-α-D-glucoside		D-Glucose-6-P, (H, K, M)
Methyl-β-D-glucoside		D-Mannose (all)
D-Gluconic acid (H, T)		
D-Glucose (K, T)		
D-Mannitol (all)		
D-Rhamnose (H, T)		
D-Sorbitol (all)		
L-Ascorbic acid (G)		
D-Arabinose (G)		
D-Glycerol (G)		
D-Ribose (C, L, M, T)‡		
Xylitol (G)		
D-Erythrose (H, L)		
Arabinol (H, L, T)		
Citrate (H, L, T)		
Phosphate (H, L, T)		
Glycerol (H, L, T)		
Lactate (H, L)		
Malate (H, L, T)		
Ethanolglycerate (H, K)		
Pyruvate (all)		
Succinate (H, L, T)		

6

8

PM-M1 to M4: 367 Carbon-Energy Substrates for Cells

A1 Negative Control	A2 Negative Control	A3 Negative Control	A4 α -Cyclodextrin	A5 Dextrin	A6 Glycogen	A7 Maltitol	A8 Maltotriose	A9 D-Maltose	A10 D-Trehalose	A11 D-Cellobiose	A12 β -Gentiobiose
No Substrate											
B1 D-Glucose-6-Phosphate	B2 α -D-Glucose-1-Phosphate	B3 L-Glucose	B4 α -D-Glucose	B5 α -D-Glucose	B6 α -D-Glucose	B7 3-O-Methyl-D-Glucose	B8 α -Methyl-D-Glucoside	B9 β -Methyl-D-Glucoside	B10 D-Salicin	B11 D-Sorbitol	B12 N-Acetyl-D-Glucosamine
C1 D-Glucosaminic Acid	C2 D-Glucuronic Acid	C3 Chondroitin Sulfate	C4 Mannitol	C5 Mannose	C6 Mannose-6-Phosphate	C7 D-Mannitol	C8 D-Mannitol	C9 D-Mannitol	C10 D-Mannitol	C11 Palatinose	C12 D-Turanose
Monosaccharides, Oligosaccharides, and Polysaccharides											
D1 D-Tagatose	D2 L-Sorbose	D3 L-Rhamnose	D4 L-Fucose	D5 D-Fucose	D6 D-Fructose-6-Phosphate	D7 D-Fructose	D8 Stachyose	D9 D-Raffinose	D10 D-Lactitol	D11 Lactulose	D12 α -D-Lactose
E1 Melibionic Acid	E2 D-Melibiose	E3 D-Galactose	E4 α -Methyl-D-Galactoside	E5 β -Methyl-D-Galactoside	E6 N-Acetyl-Neuraminic Acid	E7 Pectin	E8 Sedoheptulosan	E9 Thymidine	E10 Uridine	E11 Adenosine	E12 Inosine
Nucleosides											
F1 Adonitol	F2 L-Arabinose	F3 D-Arabinose	F4 β -Methyl-D-Xylopyranoside	F5 Xylitol	F6 Myo-Inositol	F7 Meso-Erythritol	F8 Propylene glycol	F9 Ethanolamine	F10 D,L- α -Glycerol-Phosphate	F11 Glycerol	F12 Citric Acid
G1 Tricarballic Acid	G2 D,L-Lactic Acid	G3 Methyl D-lactate	G4 Methyl pyruvate	G5 Pyruvic Acid	G6 α -Keto-Glutaric Acid	G7 Succinamic Acid	G8 Succinic Acid	G9 Mono-Methyl Succinic Acid	G10 L-Malic Acid	G11 D-Malic Acid	G12 Meso-Tartaric Acid
Alcohols and Organic Acids											
H1 Acetoacetic Acid (a)	H2 γ -Amino-N-Butyric Acid	H3 α -Keto-Butyric Acid	H4 α -Hydroxy-Butyric Acid	H5 D,L- β -Hydroxy-Butyric Acid	H6 γ -Hydroxy-Butyric Acid	H7 Butyric Acid	H8 2,3-Butanediol	H9 3-Hydroxy-2-Butanone	H10 Propionic Acid	H11 Acetic Acid	H12 Hexanoic Acid
Ketone Bodies and Short Chain Fatty Acids											

Each well has a different substrate

PMs Assay Carbon/Energy Pathways in Cells

Negative Controls

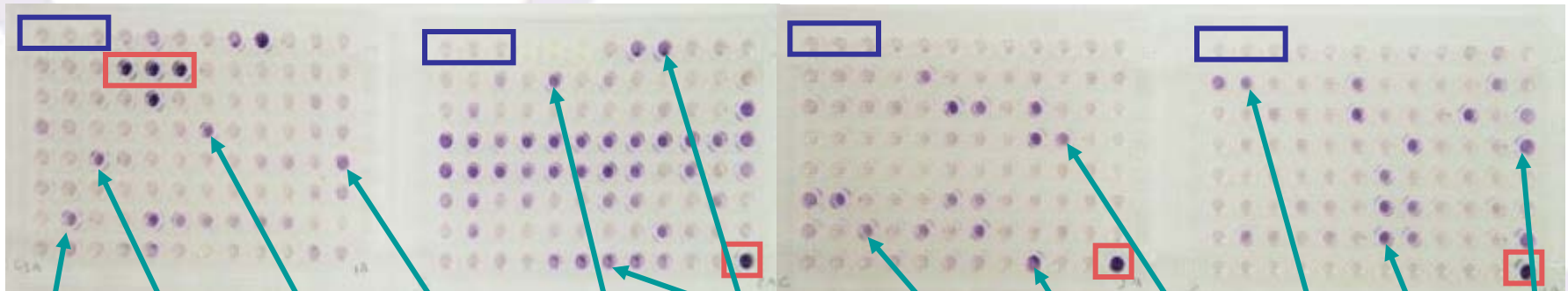
Positive Controls - Glucose

PM-M1

PM-M2

PM-M3

PM-M4



lactate galactose fructose inosine glutamine alanine gln-gly lys-tyr phe-ala leu-arg pro-arg val-ala thr-gln

sugars, alcohols, acids

fatty acids, amino acids, and dipeptides

Each Cell has a Different Set of Energy Pathways

CCRF-CEM (lymphoid)
maltotriose, maltose (A8-9)
glucose (B4-6)
mannose (C5)

HL-60 (lymphoid)

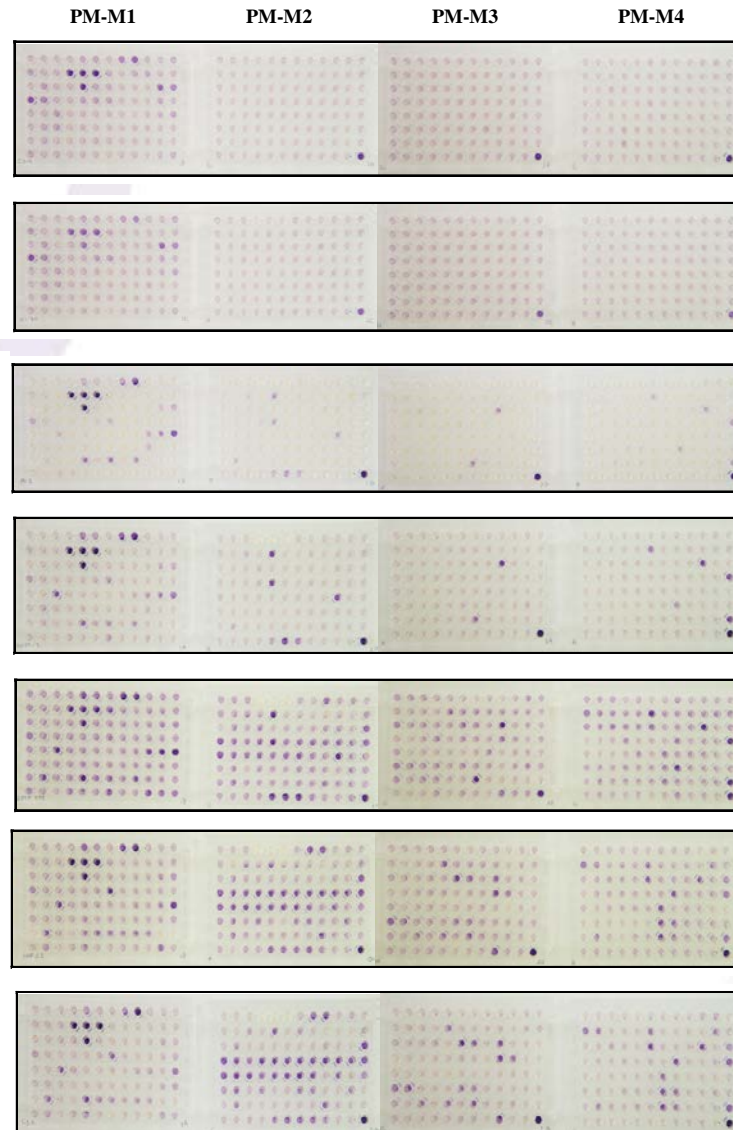
PC-3 (prostate)
fructose (D7)
uridine, adenosine, inosine
(E10-12) pyruvate, succinamate,
mono-methyl succinate (G5,7,9)

A549 (lung)
dextrin, glycogen (A5-6)
darker wells in PM-M2, M3,
and M4 correspond to glutamine
and gln-peptides

COLO 205 (colon)
galactose (E3)
lactate (G2)
butyrate, propionate (H7,10)

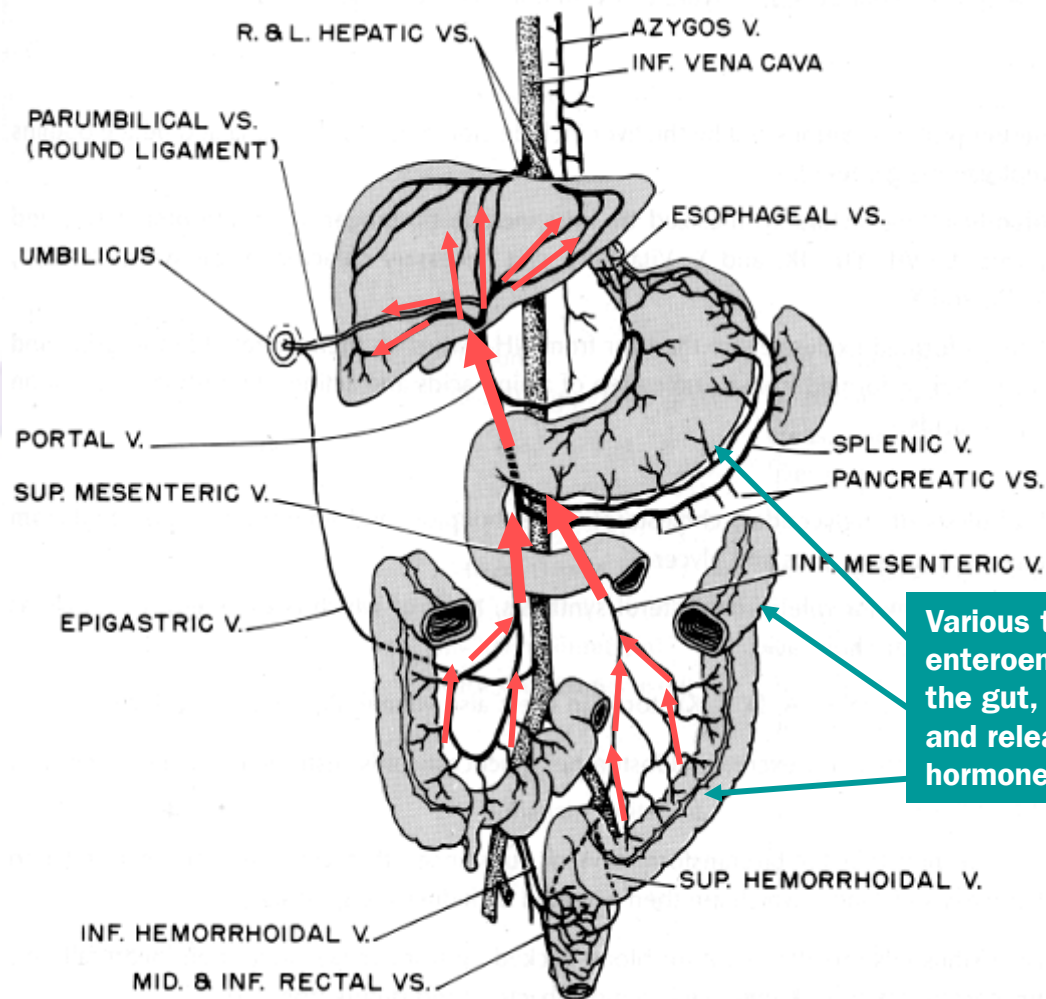
HepG2 (liver)
darker wells in PM-M2, M3,
and M4 correspond primarily to
alanine and glutamine and ala,
gln, and arg-peptides

HepG2/C3A (liver)
pyruvate (G5)



Adapted from
Bochner et. al.
PLoS ONE (2011)
6:e18147

Physiology of Nutrient Absorption

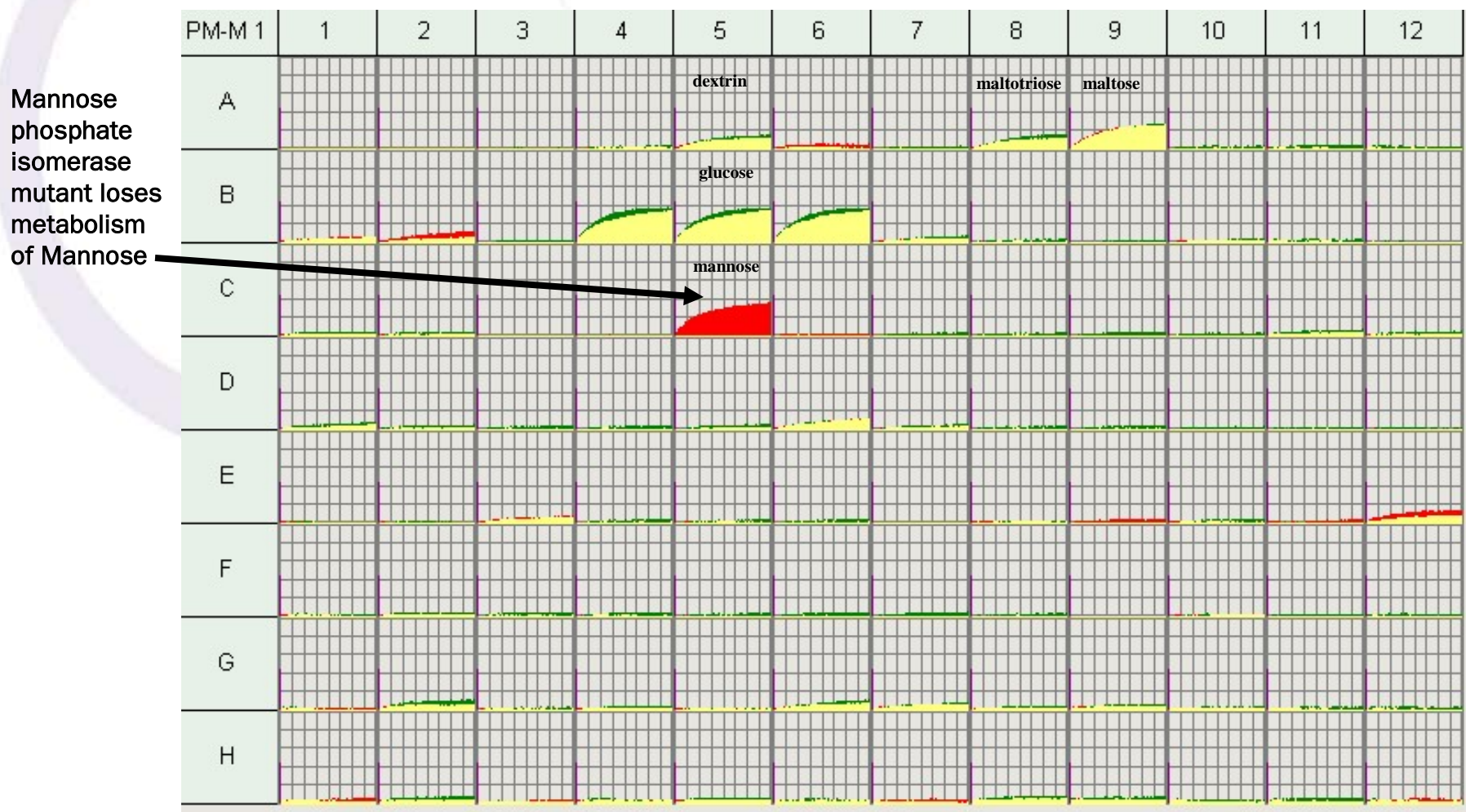


Various types of enteroendocrine cells line the gut, sense nutrients, and release signaling hormones called incretins




Analyzing Mutations in Human Cells

Comparison of HAP1 vs HAP1_MPI_124-16 on Biolog Redox Dye MB reduction (Average of 4) after dispensed into PM-M1 plates at 20,000/well in IF-M1 + 0.3 mM Gln + 5% FCS + PS medium and incubated for 20 h.

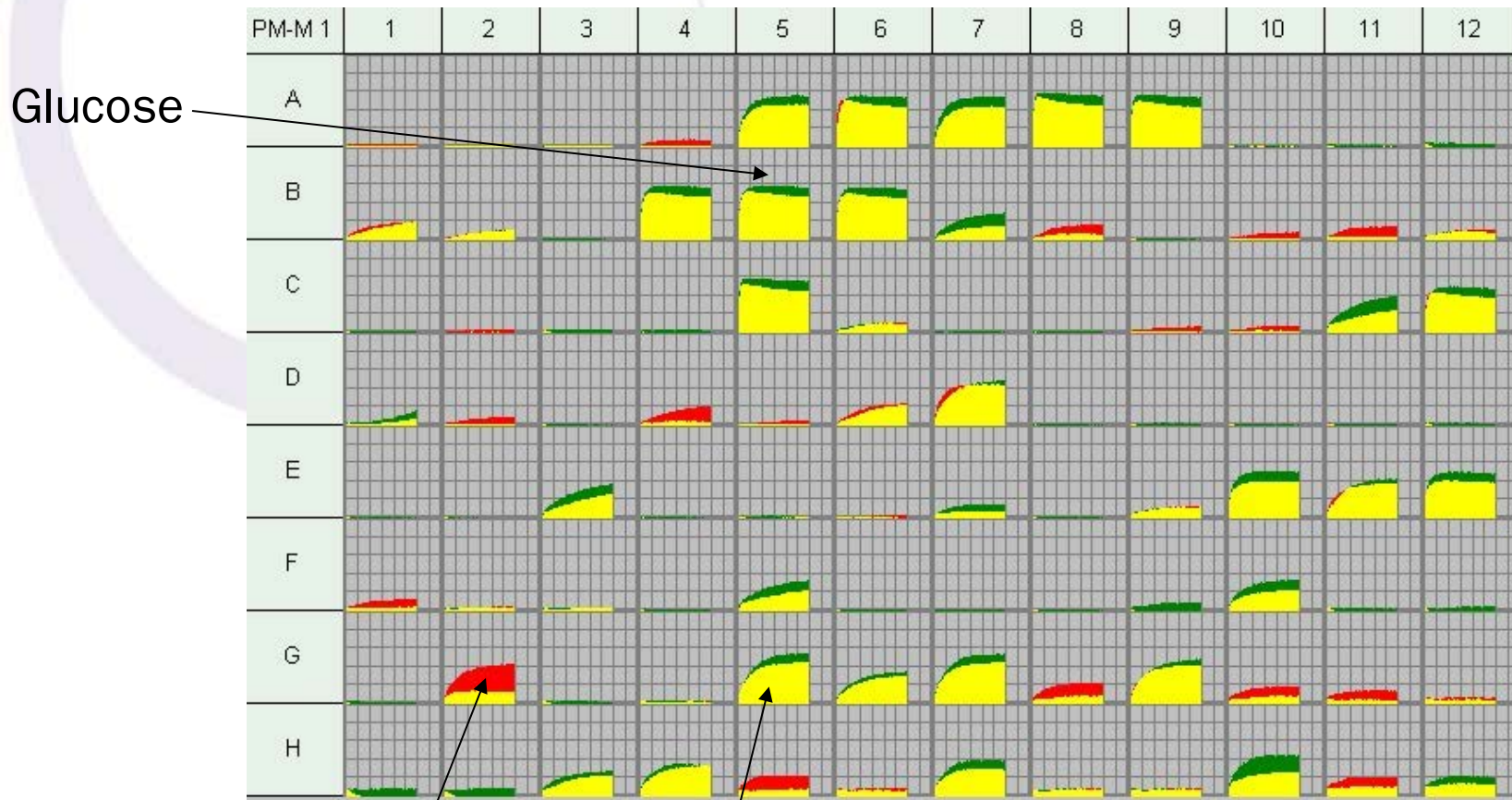


HAP1 and HAP1_MPI_MPI_124-16 (Mannose Phosphate Isomerase) , both grown in IMDM + 10% FCS + PS, were dispensed into a PM-M1 plate at 20,000 cells/well in 50 uL IF-M1 + 0.3 mM Gln + 5% FCS + PS, incubated 20 h at 37°C under 5%CO₂-95% air before adding 10 uL/well Dye MB, plates sealed with tape and placed in a 37°C OmniLog. Dye reduction over ten hour minus background



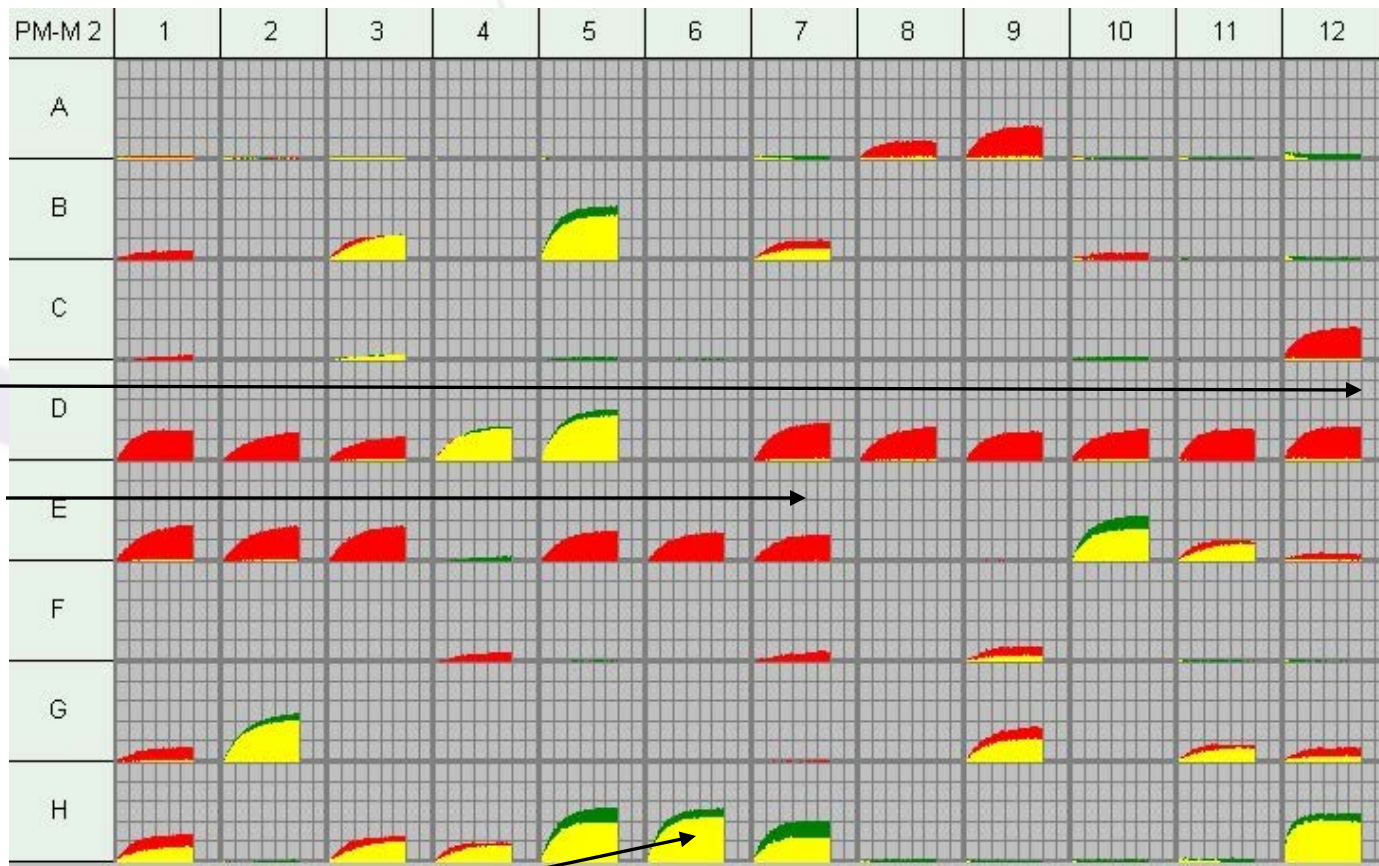
Metabolic Changes in **MCF10a** vs PI3K Clone **CL1**

Red = Normal Cell, Green = Cancer Cell



Metabolic Changes in **MCF10a** vs PI3K Clone **CL1**

Red = Normal Cell, Green = Cancer Cell



Alanine peptides

Glutamine peptides

Assay medium: IF-M1
 +5% horse serum
 +0.3mM Gln +1xP/S+ 0.1
 ug/mL cholera toxin +10
 ug/mL insulin+0.5 ug/mL
 hydrocortisone +0.2 ng/m
 EGF

20141208-12_PM-M Tox2_PI3K Inhibitor Titration_68h in CO₂_24h in OL with Dye MA

Cells: 20k/well

MCF10a / PIK3CA (H1047R/+)

MCF10a (WT)

BYL 719 (nM)

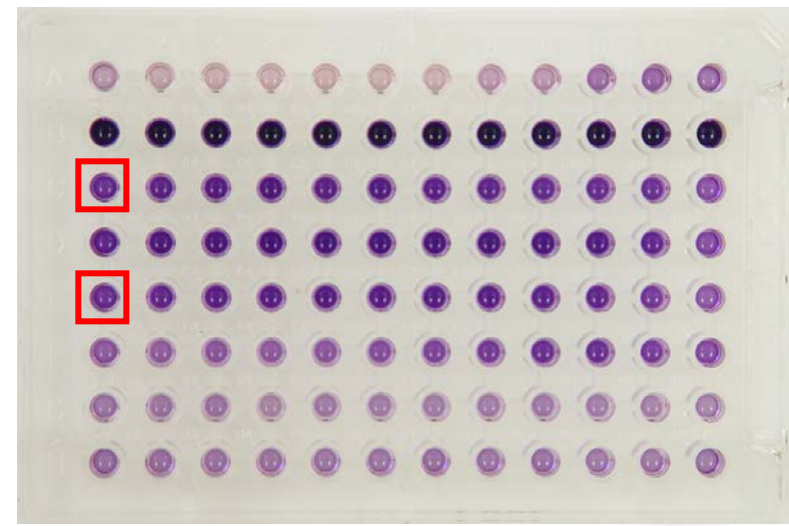
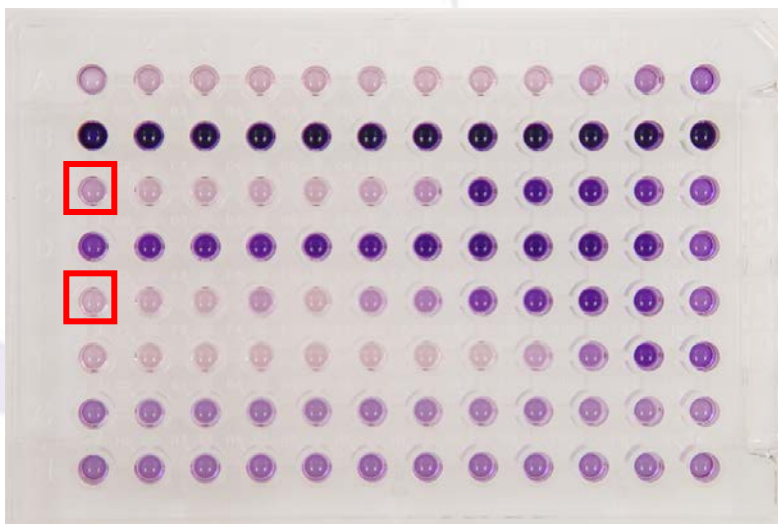
BYL 719 (nM)

0 0.5 1 2 3.9 7.8 16 31 63 125 250 500

0 0.5 1 2 3.9 7.8 16 31 63 125 250 500

BYL 719 (nM)

None
 Glycogen
 Lactate
 Glutamine
 Alanine
 Hexanoate
 Palmitate
 Oleate





Part Two:
Quality Control of Cells

Who Needs to Perform Cell Line QC ?

- Culture collections and cell banks
- Bioprocess scientists banking cells (seed cultures) to inoculate fermentations
- Cell-based assay labs
- Anyone using cells in their research

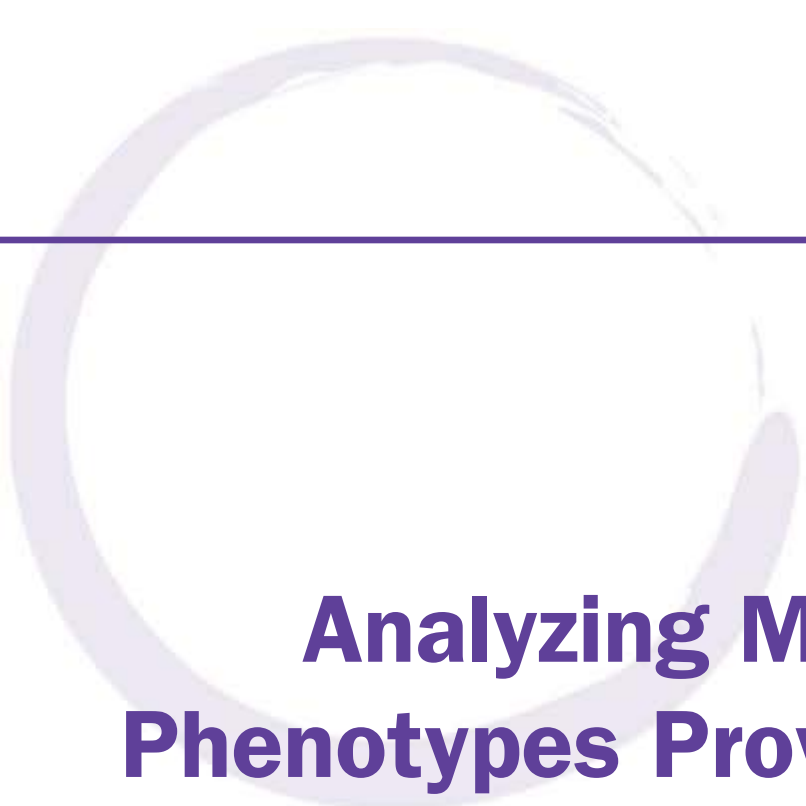
How is Cell Line QC Currently Performed ?

For Microbial Cells

- A mixture of genetic and phenotypic tests

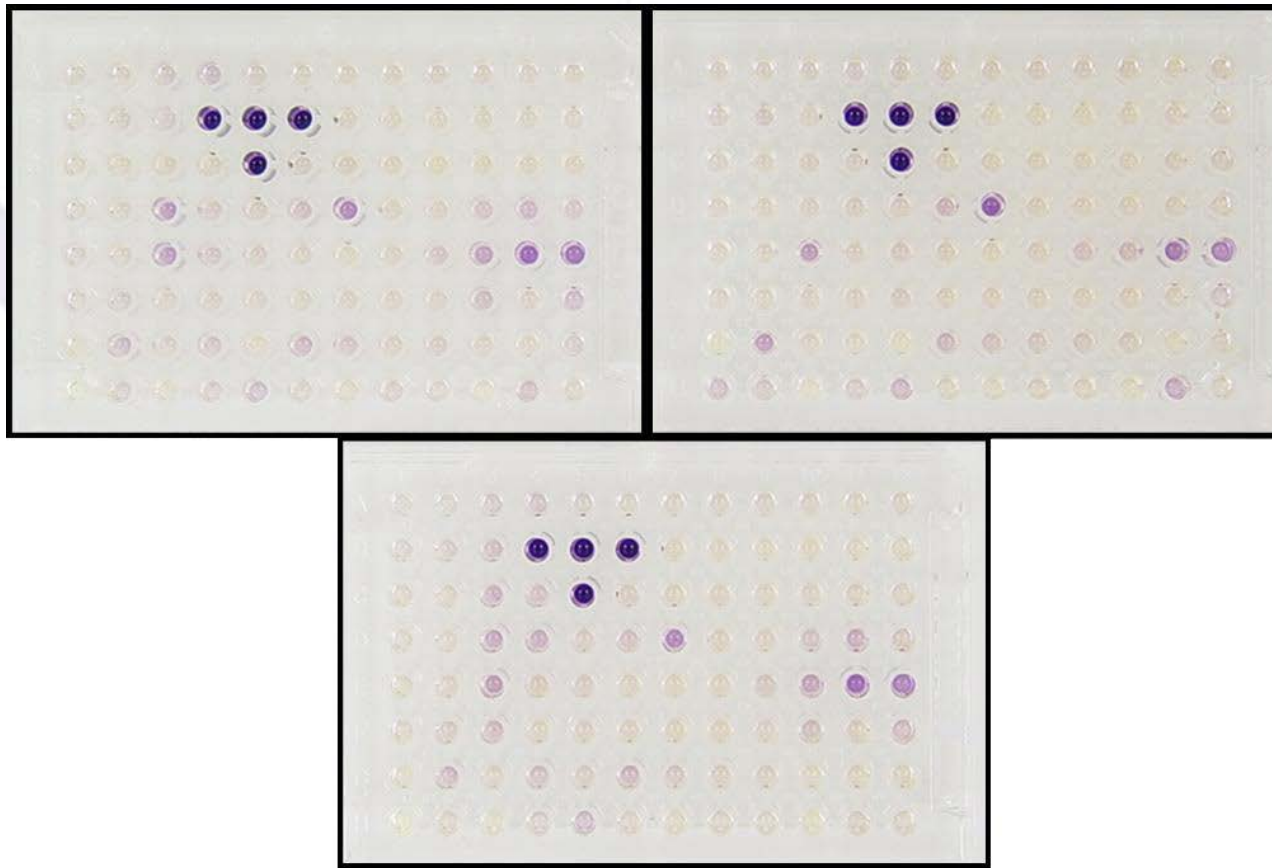
For Human and Animal Cells

- STR profiling according to cell line authentication method ASN-0002 (genetic method)
- Many labs rely on judging cell morphology

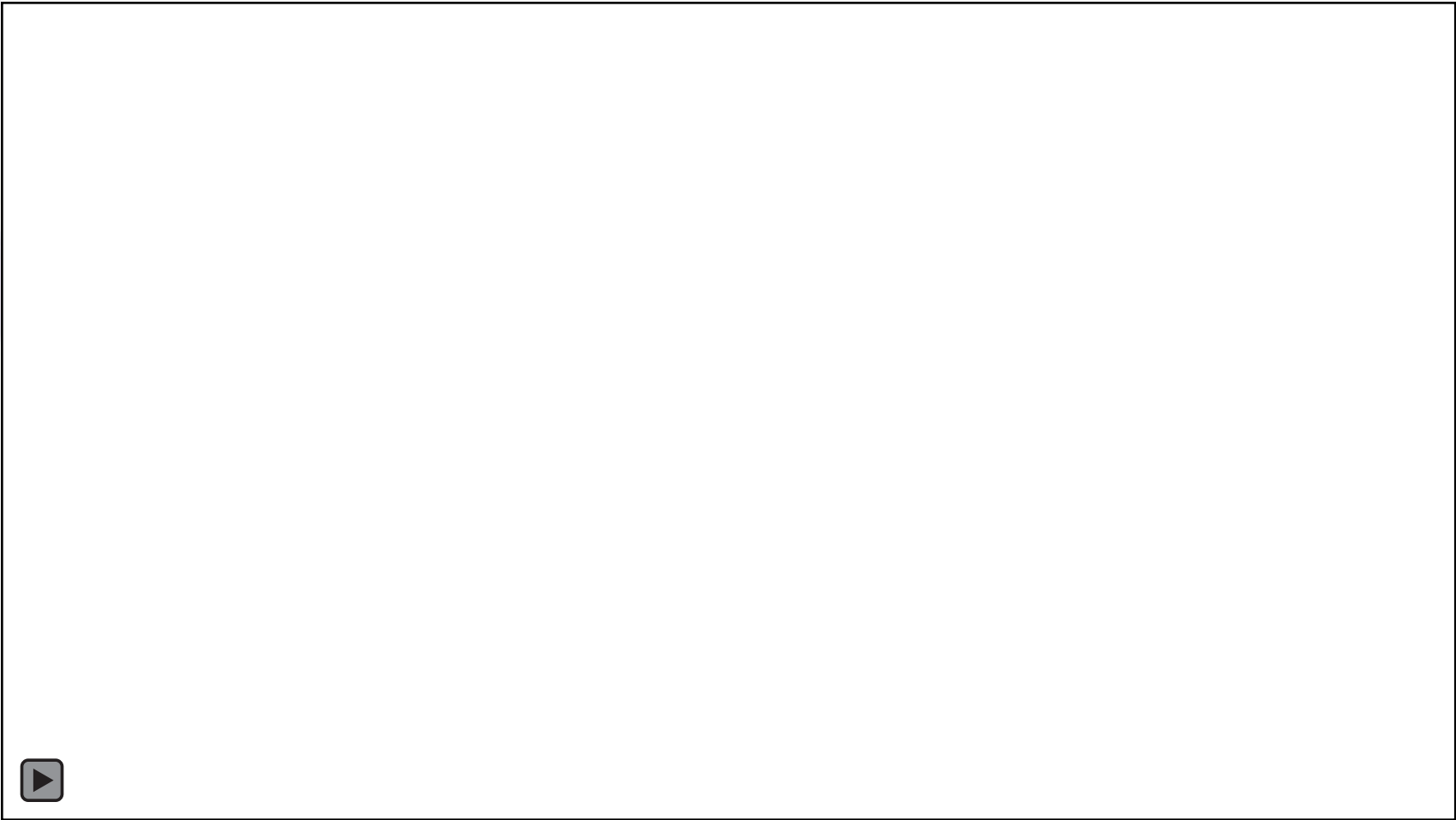


**Analyzing Metabolic and Other
Phenotypes Provides a Complementary
QC Method**

PM Fingerprints are Highly Reproducible



CVs typically
are less
than 10%



Cells Have Different Metabolic Phenotypes

CCRF-CEM (lymphoid)
maltotriose, maltose (A8-9)
glucose (B4-6)
mannose (C5)

HL-60 (lymphoid)

PC-3 (prostate)
fructose (D7)
uridine, adenosine, inosine
(E10-12) pyruvate, succinamate,
mono-methyl succinate (G5,7,9)

A549 (lung)
dextrin, glycogen (A5-6)
darker wells in PM-M2, M3,
and M4 correspond to glutamine
and gln-peptides

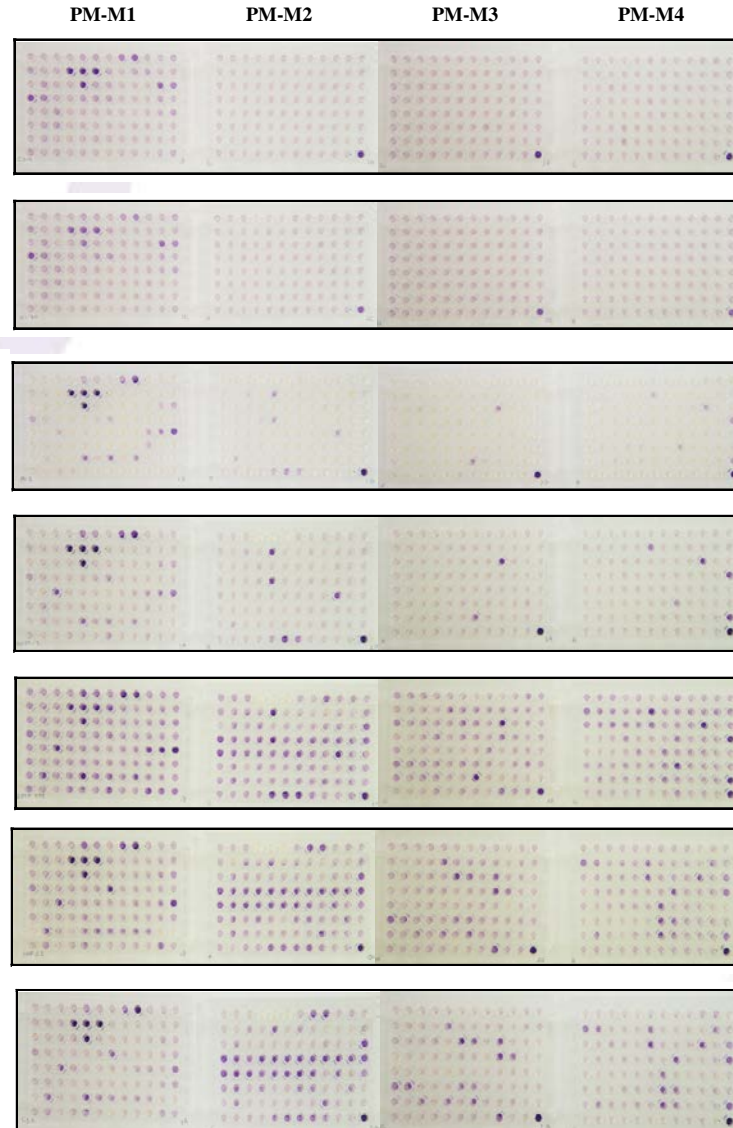
COLO 205 (colon)

galactose (E3)
lactate (G2)
butyrate, propionate (H7,10)

HepG2 (liver)
darker wells in PM-M2, M3,
and M4 correspond primarily to
alanine and glutamine and ala,
gln, and arg-peptides

HepG2/C3A (liver)

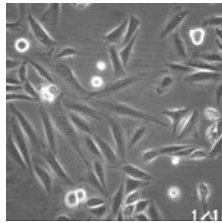
pyruvate (G5)



Adapted from
Bochner et. al.
PLoS ONE (2011)
6:e18147

PMs Distinguish Closely Related Sublines

HepG2



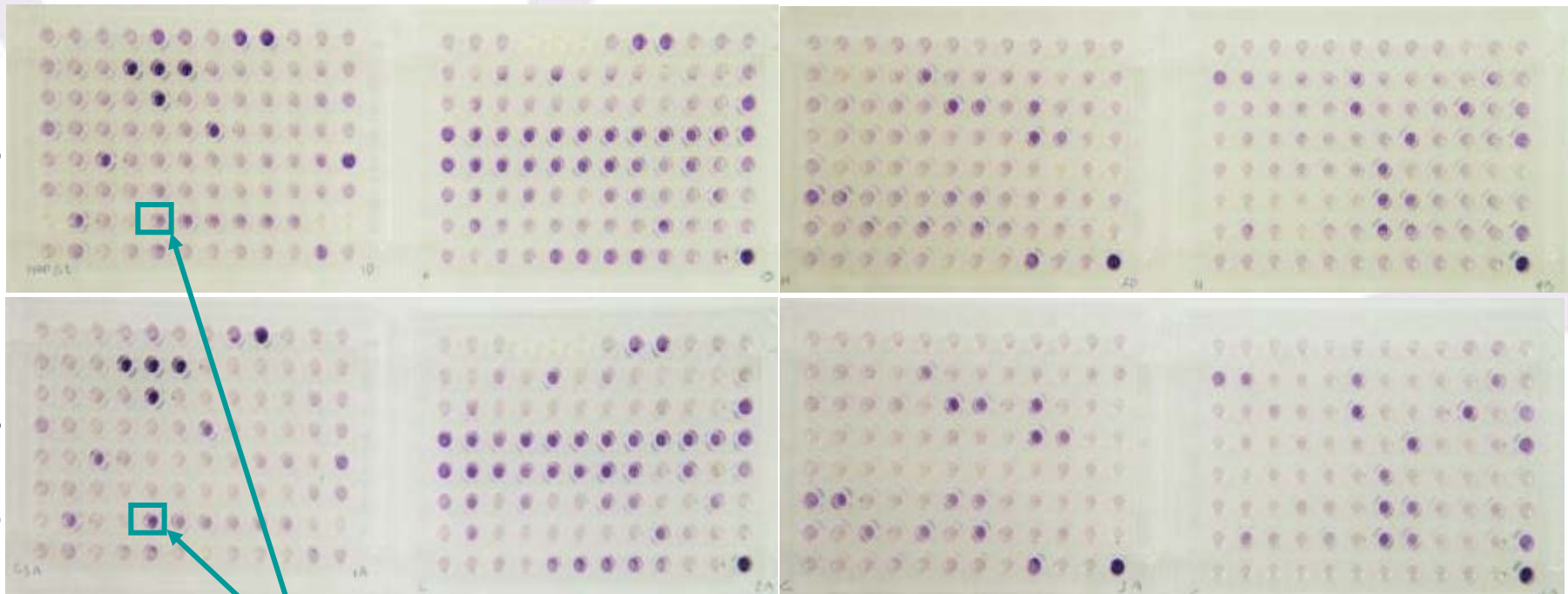
vs HepG2/C3A, a subline created for an artificial liver project

PM-M1

PM-M2

PM-M3

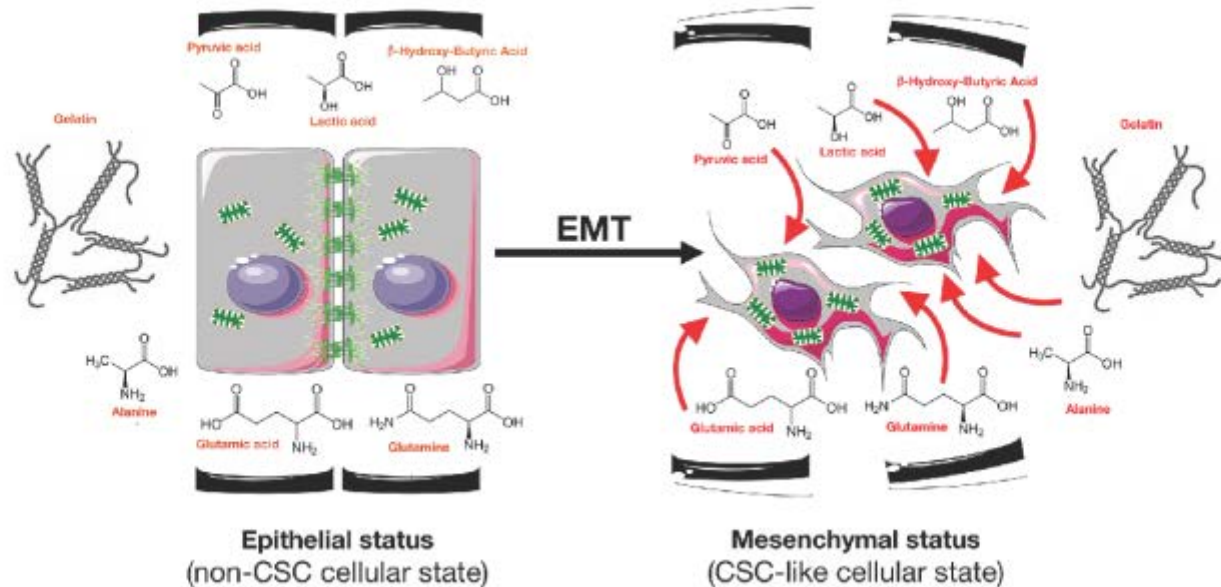
PM-M4



pyruvate

PM Comparison of Breast Cancer vs Cancer Stem Cell

Using the method of Robert Weinberg, cells can be induced to undergo an epithelial to mesenchymal transition (EMT) with stem cell-like properties

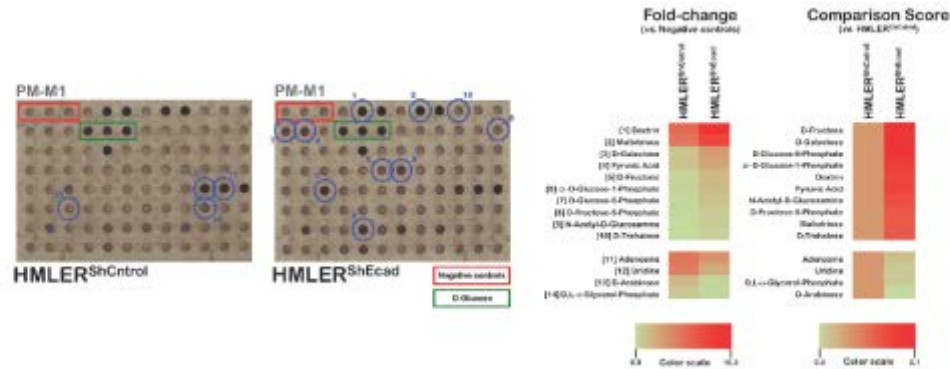


Isogenic stem cells were induced by inhibiting expression of the CDH1 E-cadherin gene via short hairpin RNA as described in

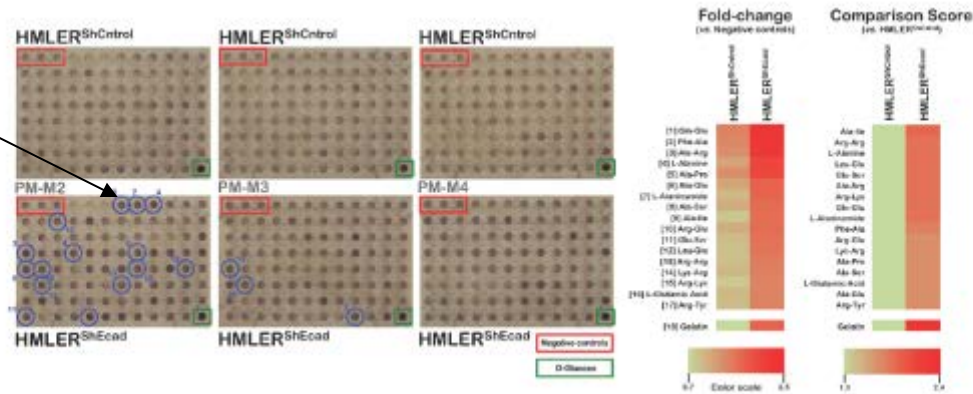
E Cuyas, B Corominas-Faja, JA Menendez, Oncotarget (2014) 5:3970

PM Comparison of Breast Cancer vs Cancer Stem Cell

“Most changes that occurred following the acquisition of a CS-like cellular state (28 out of 31, 90%) were increases in the ability to generate energy”



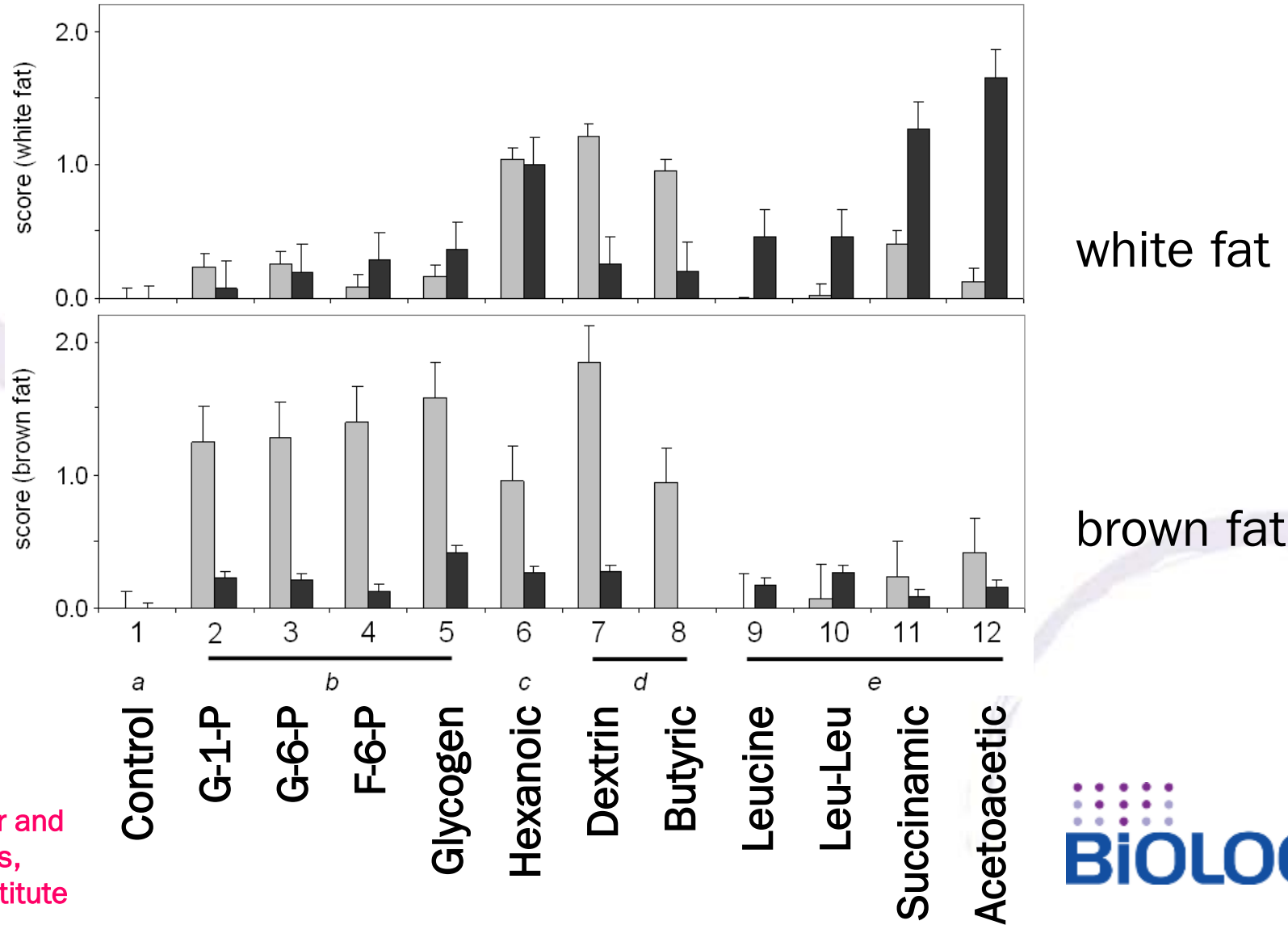
Induced metabolism of gelatin, amino acids, and peptides



Isogenic stem cells were induced by inhibiting expression of the CDH1 E-cadherin gene via short hairpin RNA as described in

E Cuyas, B Corominas-Faja, JA Menendez, Oncotarget (2014) 5:3970

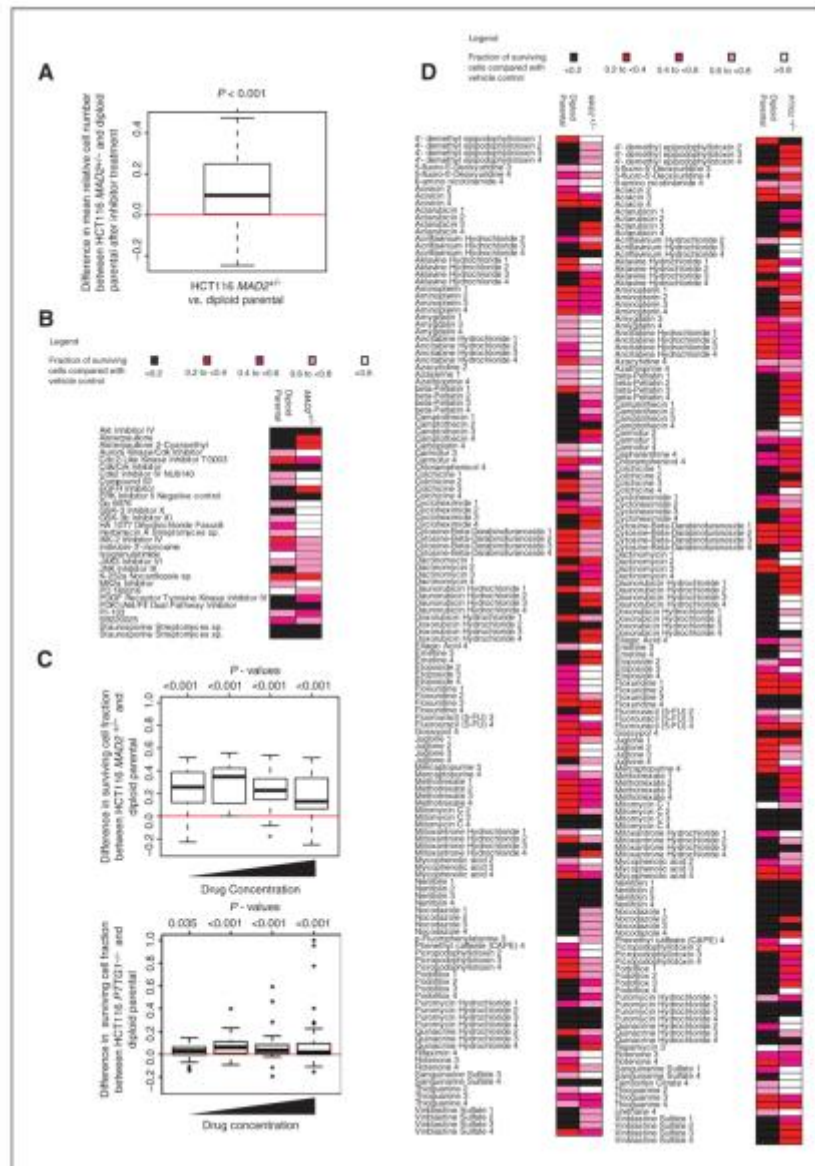
Metabolic Differences in Preadipocytes and Adipocytes




Data of
 B. Wagner and
 P. Clemons,
 Broad Institute



Resistance to Anti-Cancer Drugs in CIN⁺ Cancers



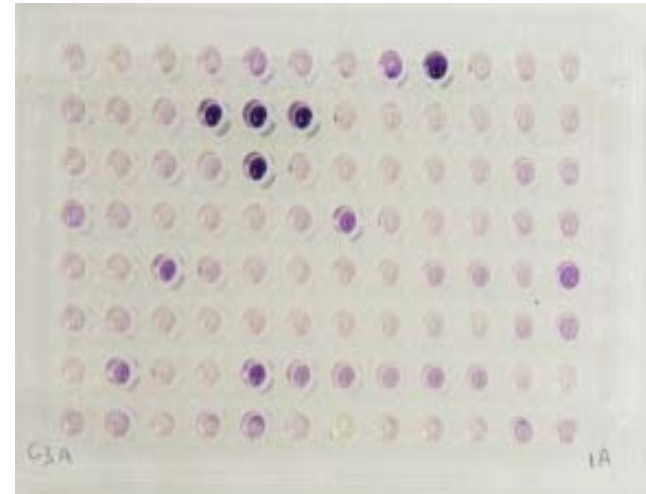
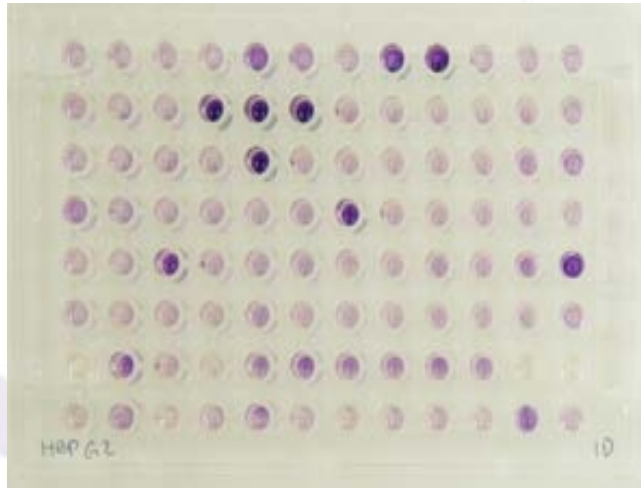
From Lee et. al.
Cancer Res.(2011)
71:1858



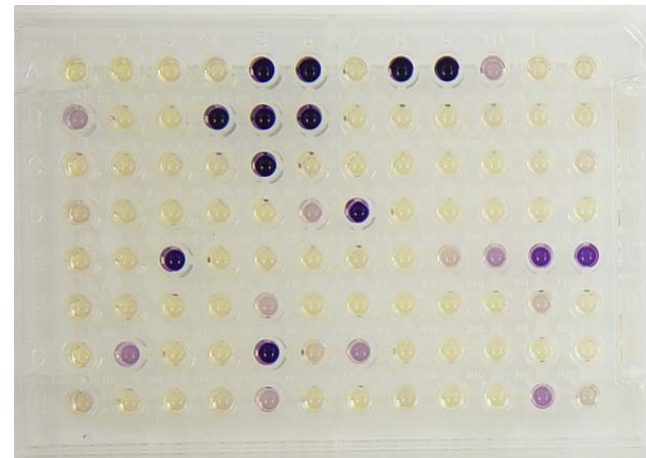
Examples of Cell Line Instability Detected by Metabolic Phenotype Analysis

Modification of a Hepatocyte Cell Line

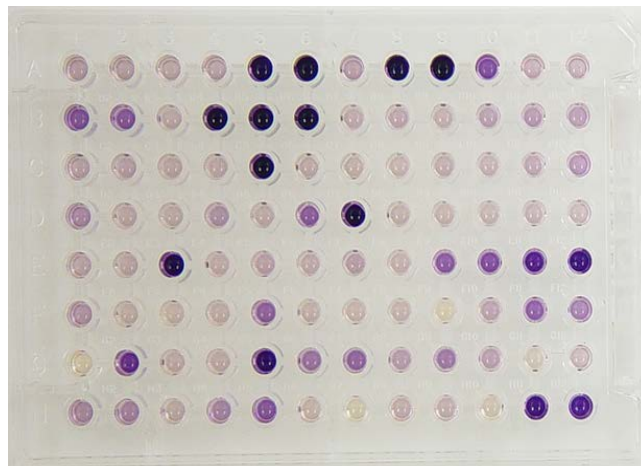
HepG2



1



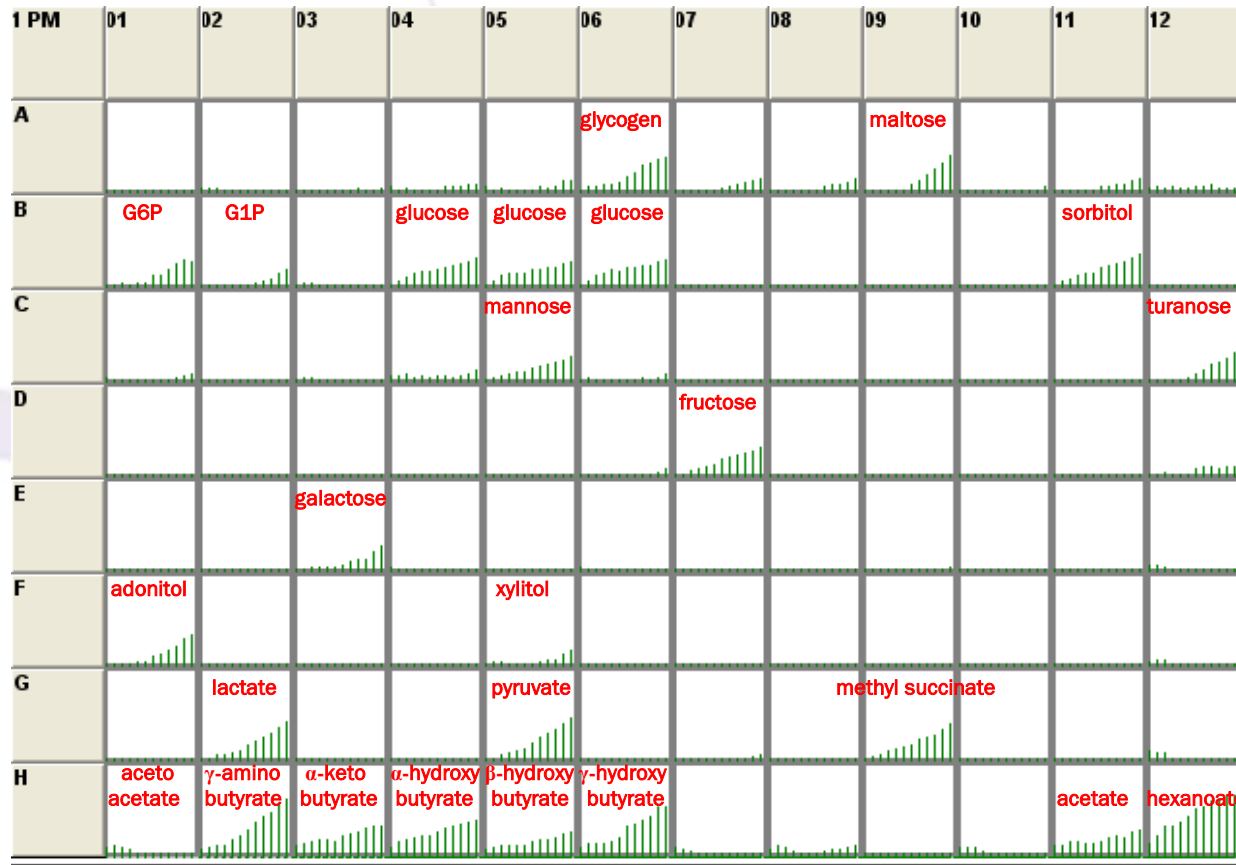
2



3

LOG

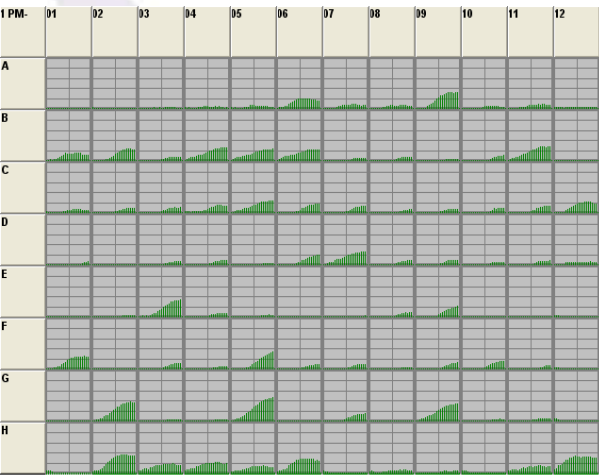
Detailed Metabolic Analysis of Primary Hepatocytes



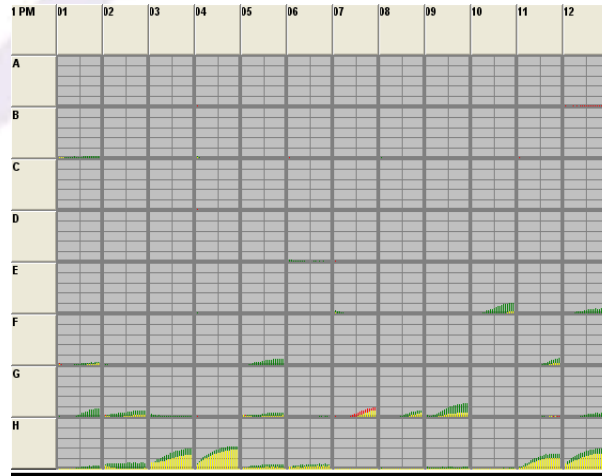
Primary rat hepatocytes, 20,000 per well,
incubated for 3 hours

Hepatocyte Preps Show Lot-to-Lot Metabolic Differences

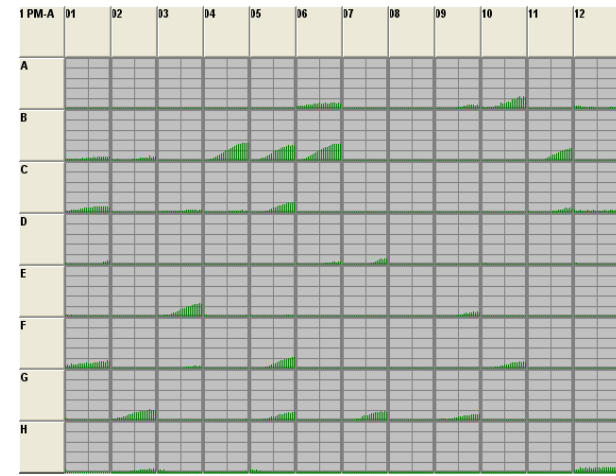
Lot 208



Lot 212

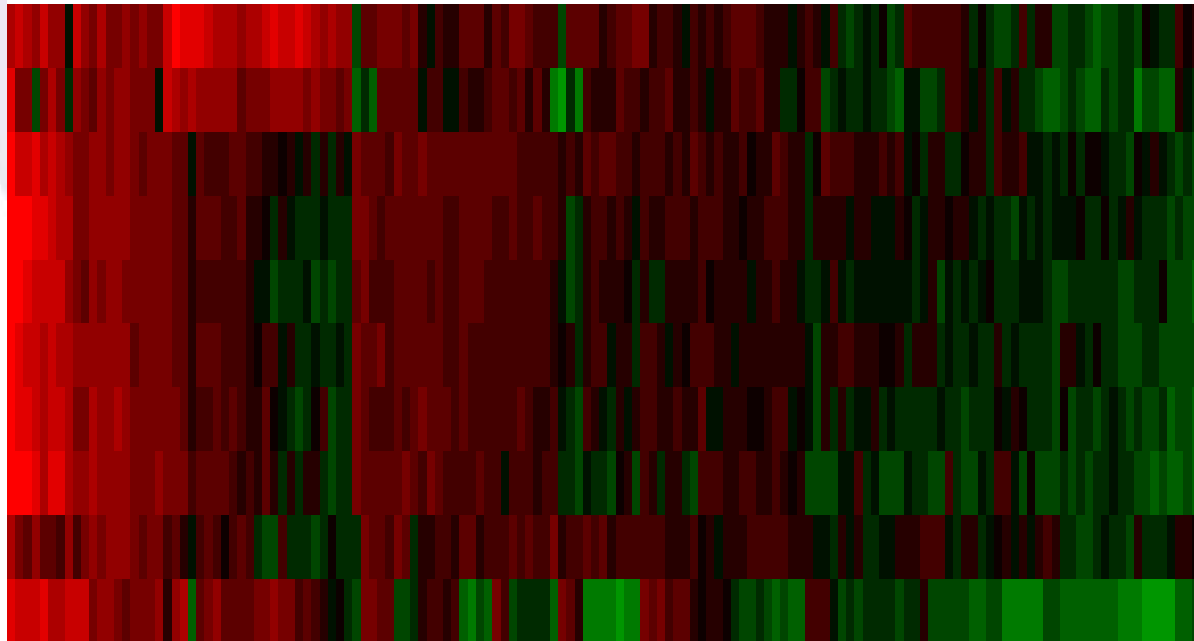


Lot 256



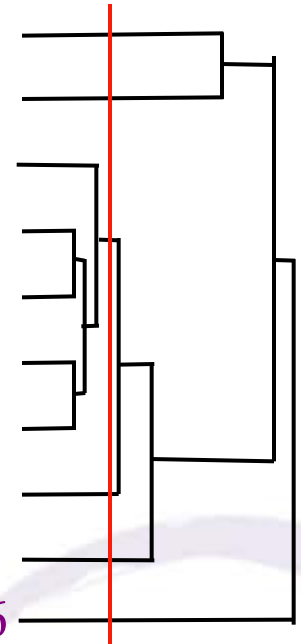
PMs Detect Changes with Cell Passaging

Various Substrates in PM-M1 through PM-M4



C3A_P13
HepG2_P17
A549_P21
A549_P17
A549_P19
A549_P50
A549_P52
A549_P54
PC3_P14
Colo205_P16

Cut Off



- QA criteria for cell line stability can be experimentally determined
- Cross-contamination can be easily detected

Issues and Limitations of DNA-based QC

Wide ranging cell stability from unstable stem cells to relatively stable cell lines that have been maintained in artificial culture for decades.

DNA-based QC would not be a preferred method for characterizing and distinguishing

- cells of different tissue/organ type
- cells with point mutations
- ips reprogrammed cells, stem cells, differentiated cells
- cells with chromosome instability
- cells with epigenetic changes
- cells with metabolic changes

All of these can potentially be detected by analysis of metabolic and other phenotypes

Other Reasons for Performing Metabolic QC

Cell Metabolism based Bioprocess Production

- CHO cells producing recombinant proteins
- Hybridoma cells producing monoclonal antibodies
- Yeast producing special wines
- Bacteria producing antibiotics
- Bacterial cocktails producing special yogurts or probiotics for faecal transplants.

Cell-based Assays

- Primary liver cells used in tox assays

Summary

- There is no perfect solution or right answer to how much and what type of cell line QC should be performed. It is a judgement call based on assessment of how stable the cell is and how best to detect changes that it is prone to make.
- In general, it is prudent to perform a metabolic/phenotypic QC in addition to a genetic QC as these are complementary analyses.
- Biolog phenotypic assays provide an ideal metabolic/phenotypic platform that spans the spectrum of cells from bacteria to human cells.
- Biolog assays are very easy and inexpensive to perform and they can be read with any microplate reader or even by eye. However the best data is obtained by measuring metabolic rates using the OmniLog instrument.

The Importance of Authentication



Methicillin-resistant
Staphylococcus aureus

-Photo courtesy of NIAID

- The use of minimally cultured strains that have been propagated and preserved under the proper conditions can help prevent:
 - Genetic drift
 - Phenotypic variation
 - Changes in functional characteristics
 - Contamination
 - Misidentification

Thank you for joining today!

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- **November 12, 2015**
10:00 AM, 3:00 PM EST
Bill Hirt, Ph.D., *Director of Accreditation*, ANAB
How Does ISO 17025 Accreditation Build International Confidence?



Please email additional questions to:
tech@atcc.org