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Outline

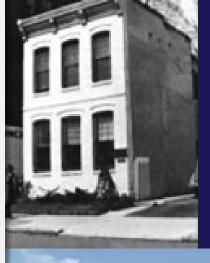


- CRISPR technology in drug discovery
- The use of CRISPR/Cas9 to create new diseasemodel cell lines
 - EML4-ALK fusion in lung cancer
 - IDH1R132H mutant in glioma
 - IDH2R140Q mutant in leukemia
- The use of CRISPR/Cas9 to create new types of drug resistant cancer cell models
 - NRASQ61K mutant in drug-resistant melanoma
 - KRASG13D mutant in drug-resistant melanoma



ATCC Snapshot

- Founded in 1925, ATCC is a non-profit organization with HQ in Manassas, VA (175,000 sf) and an R&D and Services center in Gaithersburg, MD (36,000 sf)
 - Worldwide brand name and quality recognition
- World's premiere biological materials resource and standards development organization
 - 4,000 cell lines
 - 70,000 microbes
- ATCC collaborates with and supports the scientific community with industry-standard and innovative biological solutions
 - Growing portfolio of products and services
 - Sales and distribution in 140 countries, 12 International distributors
- Talented team of 425+ employees; over one third with advanced degrees
- Multiple recognized accreditations including ISO 9001 and ISO 13485



Established partner to global researchers and scientists



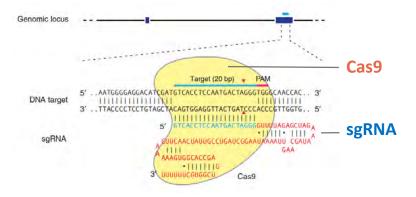


CRISPR/Cas9 gene editing technology

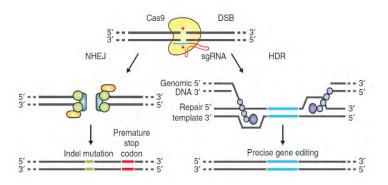
- Important novel gene editing technology
- A PubMed search will pull 5,477 articles about CRISPR
- The 'single name' journals have published issues with CRISPR on the cover
- There are many basic science and drug discovery applications for CRISPR/Cas9 gene editing

Use CRISPR/cas9 for gene editing

RNA-guided Cas9 nuclease



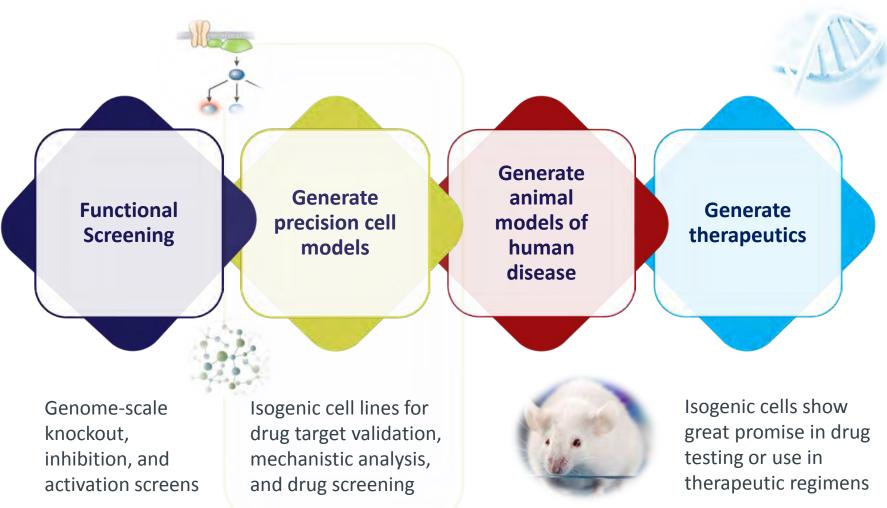
DNA repair promotes gene editing



Ran F, et al. Nat Protoc 8(11): 2281-308, 2013.



Application of CRISPR in drug discovery



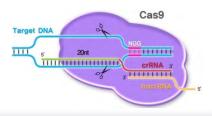


ATCC CRISPR/Cas9 gene editing platform

Molecular biology capability

- NGS
- Sanger sequencing
- ddPCR™ and qPCR
- CRISPR reagent design
- Primer design
- Expression vector toolbox
- Plasmids or oligos
 - Cell biology expertise
- Cell banking
- Cell line authentication
- Modification of extant cell lines
- Single cell cloning
- Cell line characterization

Use genome editing to create new cell models that contain rare mutations and biomarkers



CRISPR platform

- Gene knock out
- Gene insertion
- Gene modification





ATCC gene editing workflow

- Delivery of Cas9 system
- In silico design of sgRNAs
- In vitro sgRNA validation
- Donor template design (ss/dsOligo/plasmid)
- Genome wide off-target prediction analysis (computational tools)

- Single-cell cloning by cell sorting or limited dilution
- Clonal expansion and high-throughput genotyping for positive mutants
- Junction PCR and on-target sequencing of target gene(s)
- Deep sequencing of genomic/cDNA target sequence

- Gene expression via mRNA sequence analysis
- Protein expression via western blot
- Cell-based assays to validate functional and phenotypic characteristics

Cas9 reagent design and validation

CRISPR gene editing

Analysis of pooled cells for selection

Single cell isolation and validation

Off-target QC

Functional validation

Positive mutant clone

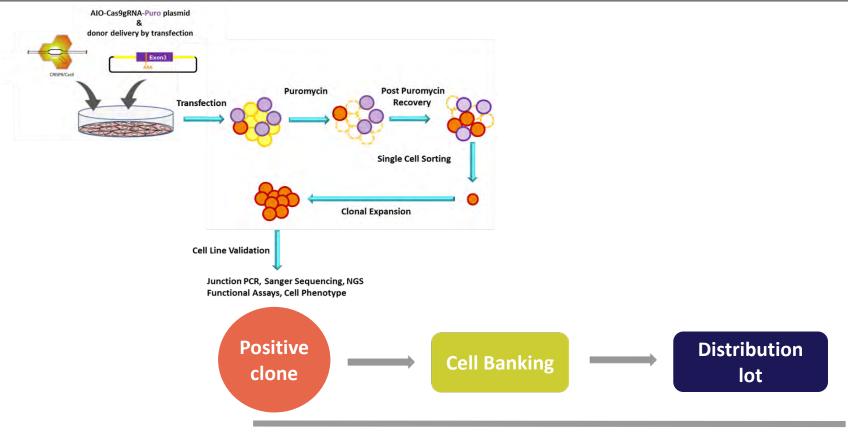
Cell line evaluation

- Selection of biologically relevant host cell line
- Sequence verification and copy number analysis of target gene(s)
- Optimize conditions for: Transfection efficiency, antibiotic selection, FACS0
- Antibiotic selection/FACS of edited cell population
- T7E1 assay/sequencing to evaluate cutting efficiency
- NGS/ddPCR™ to assess point mutation rates
- Or junctional PCR to evaluate insertion, translocation

- Screen the genome for potential off-targets by PCR amplifying OT-sites, then verify via sequencing
- Screen genome for plasmid integration including Cas9



Authenticated isogenic cell line in a vial



ATCC Quality and Standards

- Genetic verification
- Cell line purity and sterility confirmation
- Species and identify verification
- Post-freeze viability
- Functional/characterization test



Use of CRISPR system to create cancer disease models

ATCC precision cell models for the development of new anti-cancer therapeutics

Project example

 EML4-ALK fusion in non-small cell lung cancer model (ATCC® CCL-185IG™) **Gene** translocation



Driver gene mutations

Project example

- IDH1^{R132H} mutant glioma isogenic cell model (ATCC[®] HTB-14IG[™])
- IDH2^{R140Q} mutant leukemia isogenic cell model (ATCC[®] CRL-2003IG[™])

Project example

Drug-resistant melanoma models (ATCC® CRL-1619IG-1™, CRL-1619IG-2™)

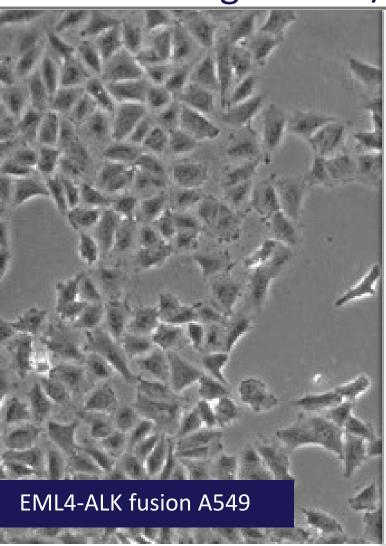
- NRAS^{Q61K} mutation KI with BRAF^{V600E}
- KRAS^{G13D} mutation KI with BRAF^{V600E}

Drugresistant mutants





Case study #1: Creation of a gene translocation and fusion using CRISPR/Cas9



EML4-ALK fusion, isogenic non-small cell lung cancer (NSCLC) model in A549 cells

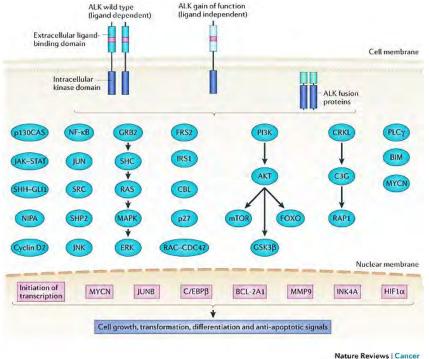
■ ATCC® CCL-185IG™

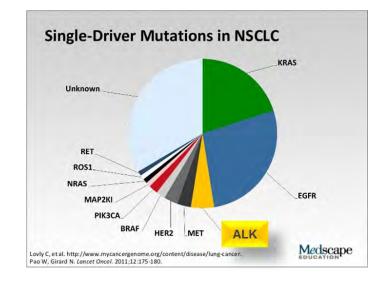


ALK is a drug target and diagnostic marker

Anaplastic lymphoma kinase (ALK) regulates the cell signaling pathway linked to cell growth, transformation, and metastasis. ALK fusion events are gain-of-function tumorigenic mutations found in NSCLC.

ALK gain of function.



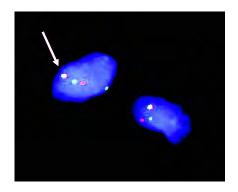


- EML4- ALK fusions have been identified as important drug targets and diagnostic biomarkers
- Lung cancer model cell lines containing EML4-ALK fusion are required for both basic oncology research and clinical drug discovery

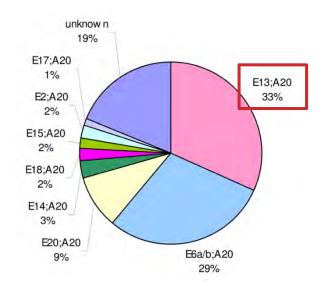


Generation of EML4-ALK fusion in NSCLC A549

EML4-ALK fusion in clinical samples

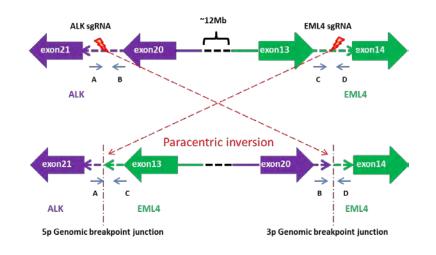


Clin Cancer Res 14(13): 4275-4283, 2008.



EML4-ALK variant 1 genomic inversion



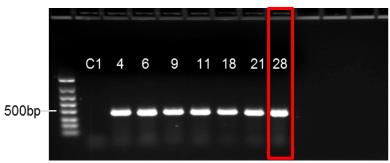




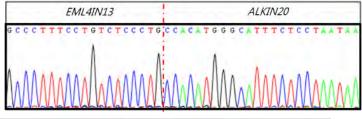
Eur J Cancer 46(10): 1773-1780, 2010.

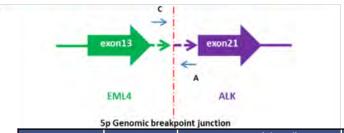
Confirm the designed gene fusion event

5' Genomic breakpoint junction



A. Sequence analysis verifying junction of single clone



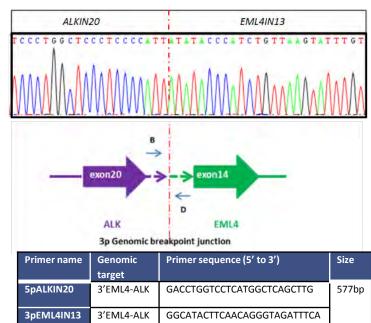


Primer name	Genomic target	Primer sequence (5' to 3')	Size
5pEML4IN13	5'EML4-ALK	TGAAACTCCCACACCTTTGCTTTTTG	545bp
		TGTTTTCTTAC	
3pALKIN20	5'EML4-ALK	TGCAGCCATTTGGAATGTCCCCTTT	
		AAATTTAGAAACAG	

3' Genomic breakpoint junction



B. Sequence analysis verifying junction of single clone



TGGTATGTGAATTATATCG



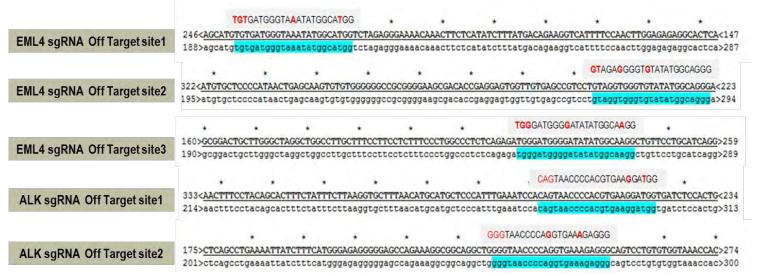
Off-target site evaluation by sequencing

Top potential off target sites:

EML4 sg Target sites/Off Target sites	Target sites/Off Target sites Sequence	On/Off-target score	Gene	Gene Segment	Chromo some
EML4 sgRNA Target site	ACAGATGGGTATATATGGCAGGG	1	EML4		2
EML4 sgRNA Off Target site1	TGTGATGGGTAAATATGGCATGG	0.568575	AGA	IG	4
EML4 sgRNA Off Target site2	GTAGAGGGGTGTATATGGCAGGG	0.5054	ASCL1	1	11
EML4 sgRNA Off Target site3	TGGGATGGGGATATATGGCAAGG	0.568575	ETS1	ı	11
ALK sg Target sites/Off Target sites	Target sites/Off Target sites Sequence	On/Off-target score	Gene	Gene Segment	Chromo some
ALK sgRNA Target site	GTATAACCCCACGTGAACGAGGG	1	ALK	1	2
ALK sgRNA Off Target site1	GTATAACCCCACGTGAA <mark>G</mark> GA T GG	0.16245	RP11-749H17.1	ı	18
ALK sgRNA Off Target site2	GTATAACCCCA <mark>G</mark> GTGAAAGAGGG	0.113715	CPE	ı	4

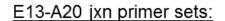
Red: potential mismatched bases

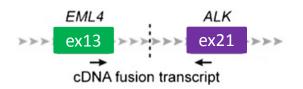
Genome screening for off-target cut sites within the selected isogenic clone:





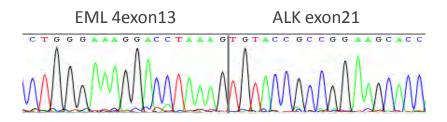
Molecular and functional validation of EML4-ALK





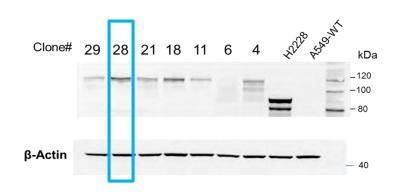


Sequence of EML4-ALK v1 fusion transcript across cDNA breakpoint



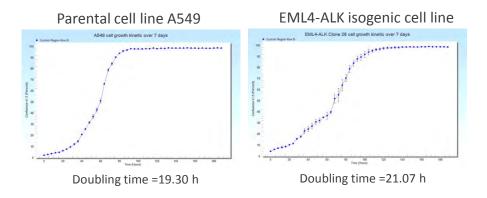
Verify that the EML4-ALK v1 fusion gene express EML4-ALK v1 fusion protein



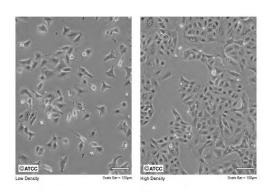


Additional functional validation

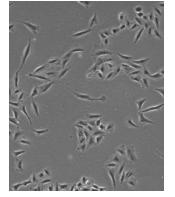
- Cell morphology
- Cell growth kinetics
- Cell STR profile
- Cell response to therapeutics

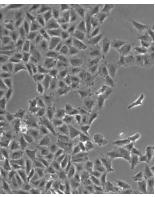


Parental cell line A549



Isogenic cell line CCL-185IG



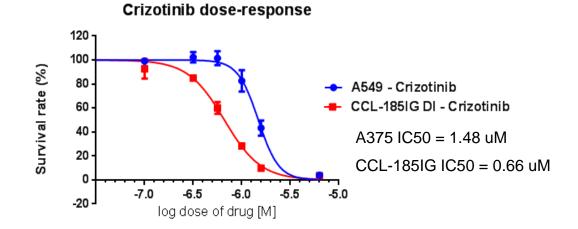




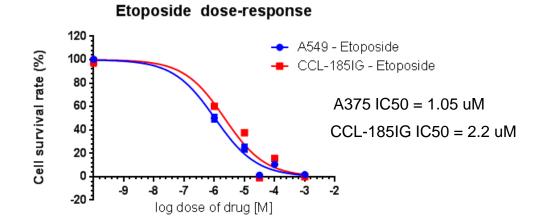
EML4-ALK isogenic cell line drug response

EML4-ALK isogenic line (ATCC® CCL-185IG™) is more sensitive to ALK inhibitor than its parental line

ALK specific inhibitor



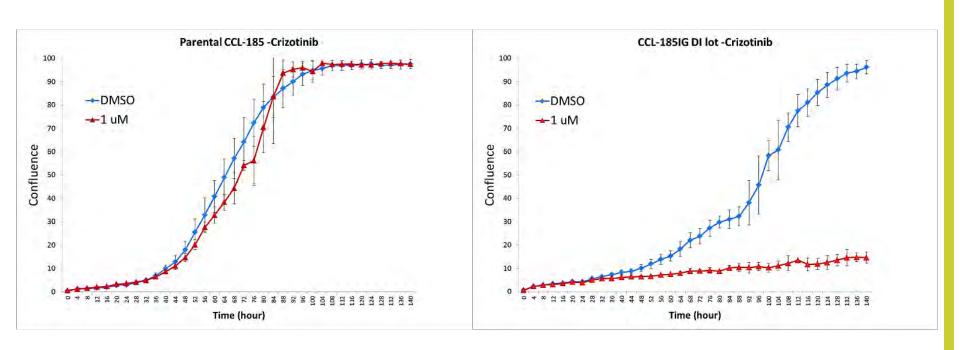
Non-specific chemotherapy drug





CCL-185IG EML4-ALK is sensitive to ALK inhibitors

EML4-ALK isogenic line (ATCC® CCL-185IG™) is more sensitive to ALK inhibitor than its parental line

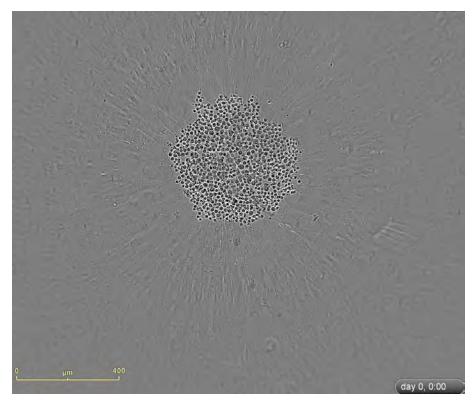


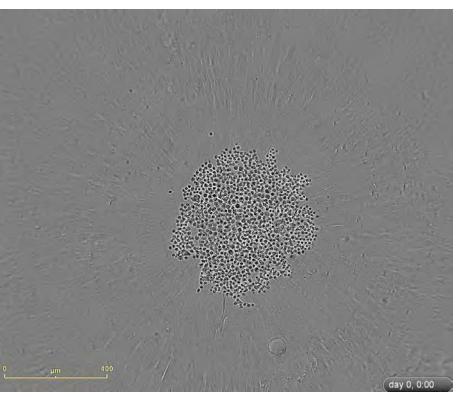


Isogenic pair in 3D culture

A549 parental cell line (ATCC® CCL-185 ™)



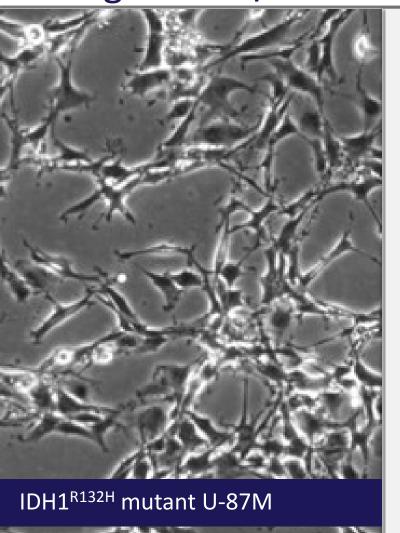




Video clips of the cell growth in 3D culture environment (1-9 days). The EML4-ALK isogenic line (right) grows more aggressively in 3D culture than the parental line (left).



Case study #2: Creation of driver gene mutations using CRISPR/Cas9



IDH1^{R132H} mutant glioma isogenic cell model in U-87MG

ATCC® HTB-14IG™

IDH2^{R140Q} mutant leukemia isogenic cell model in AML TF-1

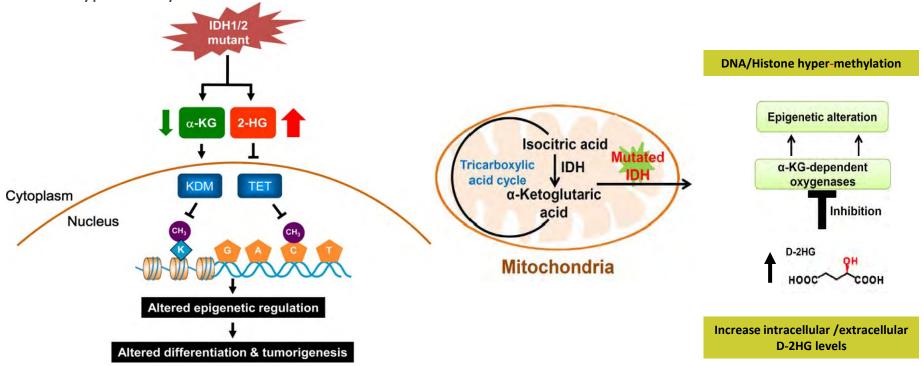
■ ATCC® CRL-2003IG™



IDH1/2 mutations in cancer

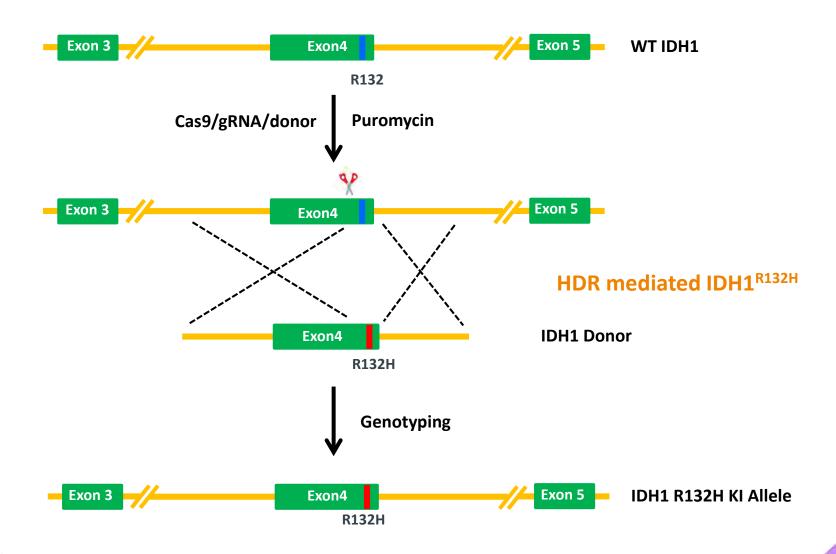
The Role of IDH1^{R132H} & IDH2^{R140Q} Gene Mutations in Cancer

- IDH1 and IDH2 genes are mutated in glioma, acute myeloid leukemia, and other types of human cancer.
- •IDH1/2 mutations cause a gain-of-function in cancer cells resulting in accumulation and secretion of the oncometabolite (2HG). This causes dysregulation of demethylases leading to histone and DNA hypermethylation.





Generation of the IDH1^{R132H} isogenic glioma U-87MG line



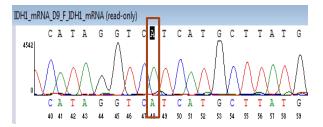


Generation of the IDH1^{R132H} isogenic glioma U-87MG line

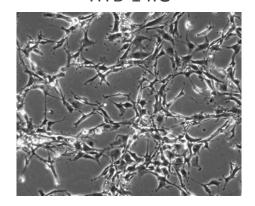
IDH1^{R132Q} mutant TF-1 Isogenic Cell Line (ATCC[®] HTB-14IG[™])

- Sequence gDNA to confirm IDH1^{R132H} knock-in
- mRNA level validation
- Off-target screening
- Cell morphology
- Cell growth kinetics
- Cell STR profile
- Bio-functional assays

Isogenic line IDH1^{R132H} transcript



HTB-14IG



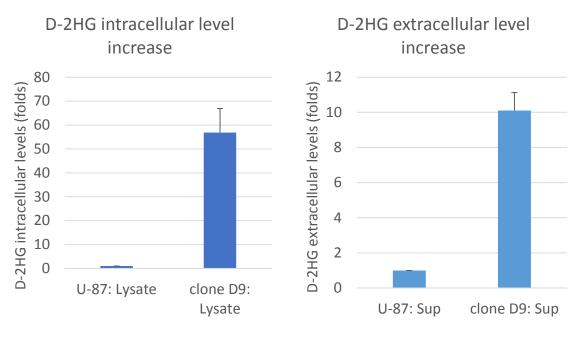
Doubling time = 25.6 h

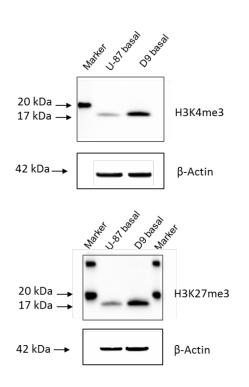


Bio-functional validation of the IDH1^{R132H} isogenic line

Significantly increased intracellular and extra-cellular 2-HG levels in HTB-14IG

Histone hyper-methylation in HTB-14IG





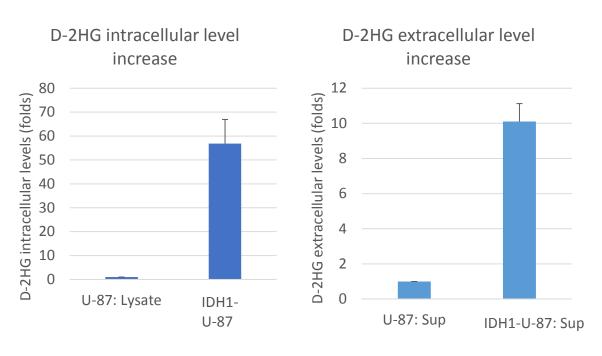
Pico-Probe assay was used to detect D-2HG level.

IDH1 mutations cause a gain-of-function in cancer cells, resulting in accumulation and secretion of the oncometabolite D-2-Hydroxyglutarate (D-2HG). This causes inhibition of proteins involved in epigenetic regulation leading to DNA and histone hypermethylation.

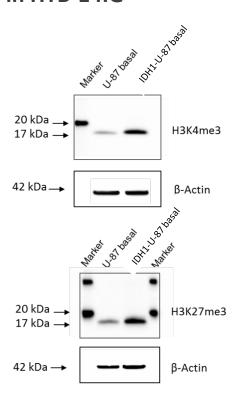


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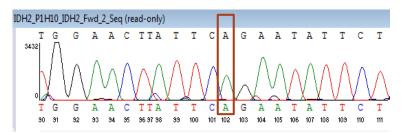


Generation and validation of the IDH2^{R140Q} isogenic TF-1 line

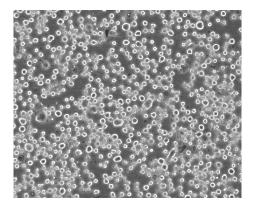
IDH2^{R140Q} mutant TF-1 Isogenic Cell Line (ATCC[®] CRL-2003IG[™])

- Sequence gDNA to confirm IDH2^{R140Q} knock-in
- mRNA level validation
- Off-target screening
- Cell morphology
- Cell growth kinetics
- Cell STR profile
- Cell response to therapeutics

Isogenic line IDH2^{R140Q} transcript



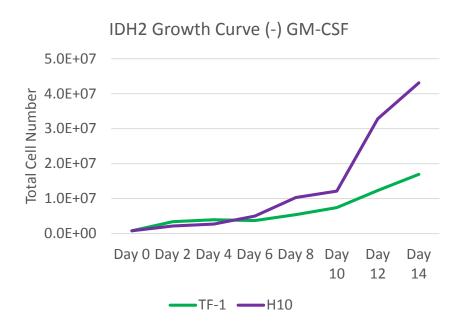
CRL-2003IG



Doubling time = 21.4 h



Impact of the IDH2^{R140Q} mutation on cell growth and differentiation



TF-1 Parental IDH2^{R140Q} (ATCC® CRL-2003IG™)

TF1 cells are GM-CSF-dependent, while IDH2^{R140Q} mutants exhibit GM-CSF-independent proliferation

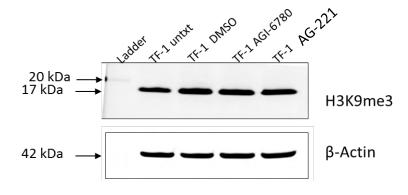
IDH2^{R140Q} mutant cells attach to plate, exhibit spindle-like (undifferentiated mesenchymal) morphology

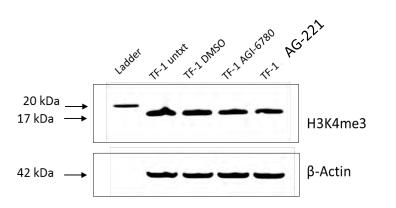


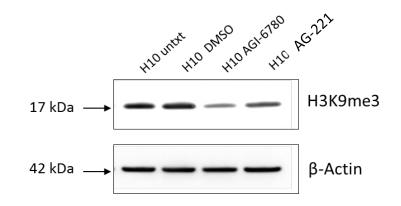
IDH2 inhibitors decrease histone methylation in the IDH2^{R140Q} isogenic line

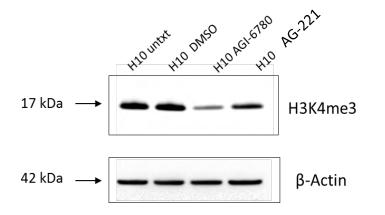
TF-1 Parental line (ATCC® CRL-2003™)

IDH2^{R140Q} Isogenic line (ATCC® CRL-2003IG™)



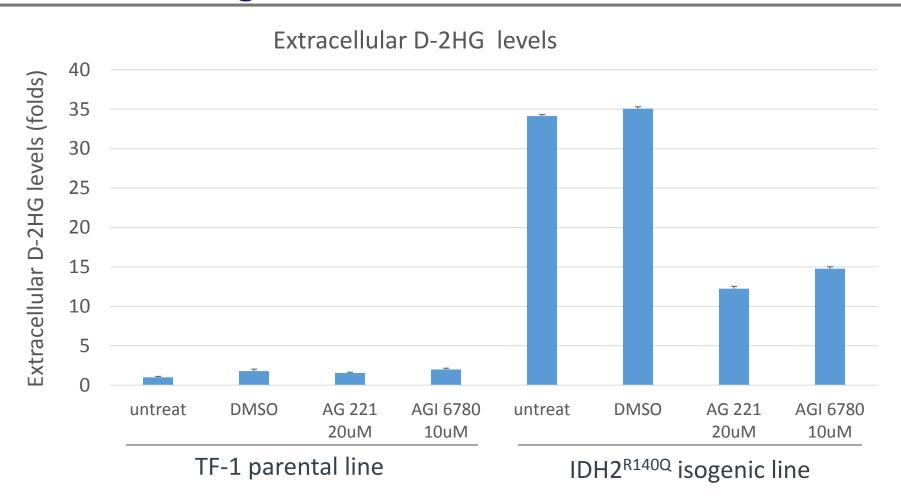








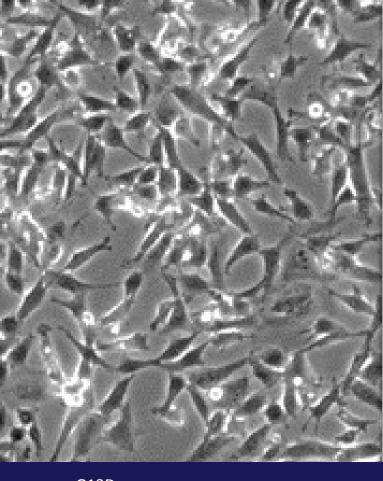
IDH2 inhibitors decrease 2-HG levels in the IDH2^{R140Q} isogenic line





Parental and IDH2 isogenic cell lines were cultured with or without IDH2-specific inhibitors (AG221 and AGI6780) in triplicate for 3 days. Pico-ProbeTM D-2HG assay kit (BioVision) was used to detect D-2HG levels days post drug treatment showing several fold reduction in extracellular D-2HG levels.

Case study #3: Creation of drug-resistant mutations using CRISPR/Cas9



KRAS^{G13D} mutant A375

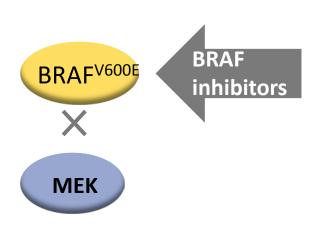
BRAF inhibitor-resistant melanoma models (ATCC® CRL-1619IG-1™, CRL-1619IG-2™)

- NRAS^{Q61K} mutation in A375 BRAF^{V600E} melanoma line
 - Available now
- KRAS^{G13D} mutation in A375 BRAF^{V600E} melanoma line
 - Coming soon



Therapeutic targeting of BRAFV600E in melanoma

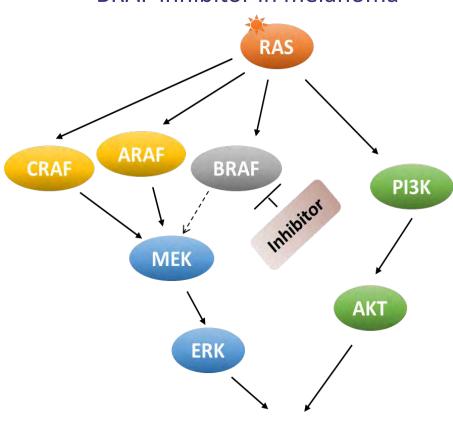
Role of mutant BRAF^{V600E} in driving cell survival: Oncogenic BRAF signaling





Decrease cell growth and survival

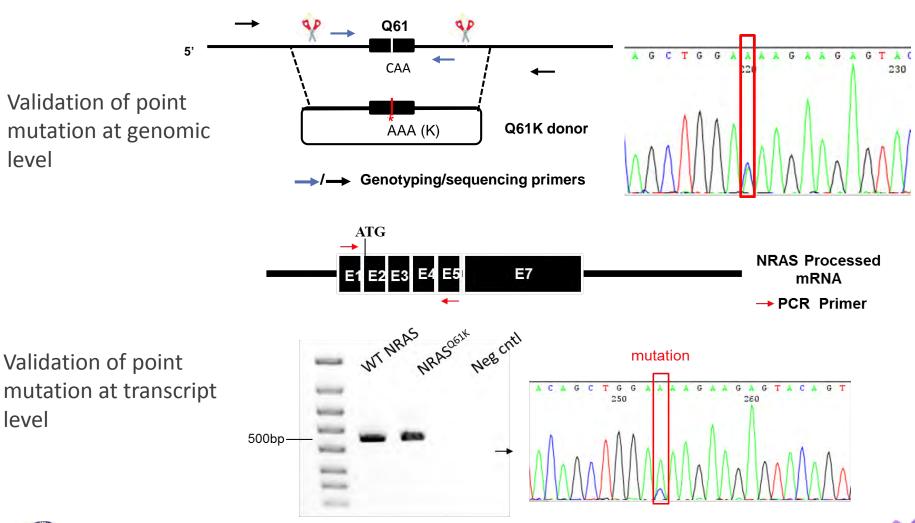
RAS mediated resistance to BRAF inhibitor in melanoma



Cell proliferation, cell survival



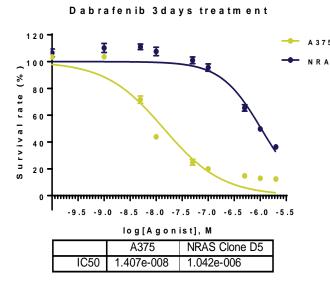
Generation of NRAS^{Q61K} isogenic melanoma A375 line

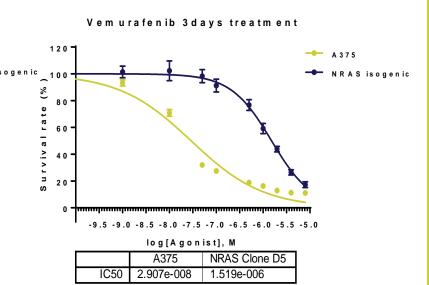




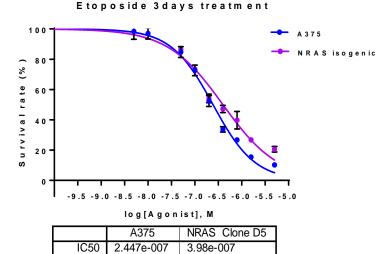
Drug resistance of NRAS^{Q61K} isogenic melanoma A375 line CRL-1619IG-2

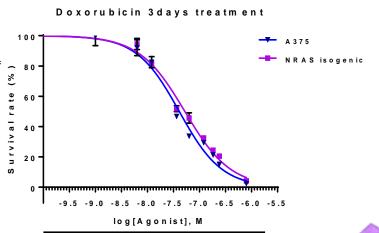
BRAF specific inhibitor





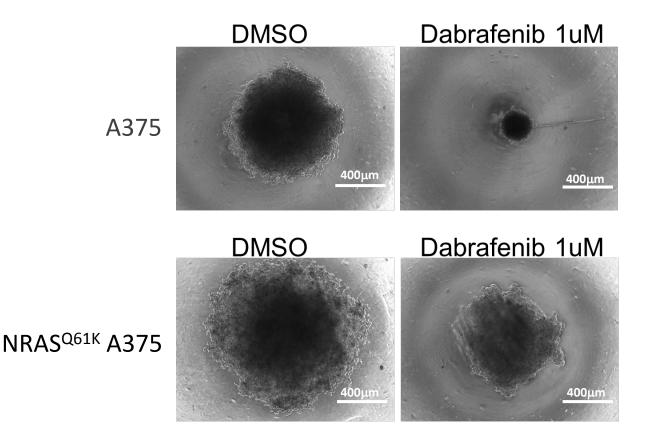
Non-specific chemotherapy drug





A375 NRAS Clone D5
IC50 4.093e-008 5.238e-008

3D culture evaluation of isogenic pair

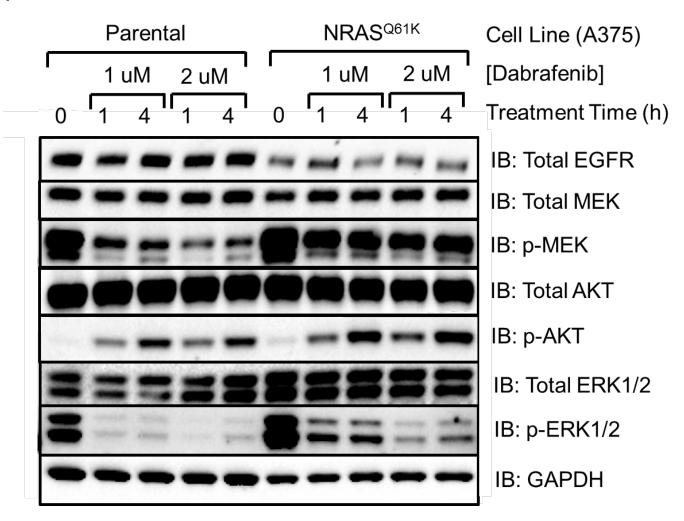


3D culture spheroids were generated by seeding cells on an ultra-low attachment plate and culturing for 4 days. Spheroids were then incubated for 10 days in the presence or absence of 1 μ M BRAF inhibitor debrafenib and then imaged. The NRAS Q61K isogenic line displays resistance to BRAF inhibitor, mimicking clinical emergence of BRAF-inhibitor resistant cancers.



ERK and AKT activation within NRAS^{Q61K} Mutant A375 Isogenic Cell Line

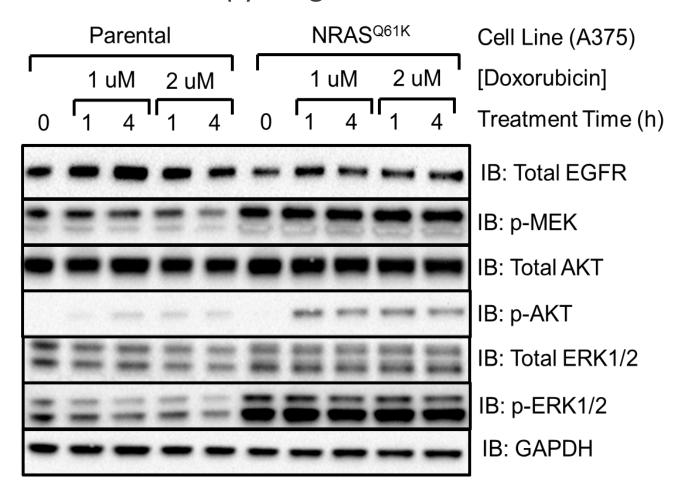
BRAF specific inhibitor treatment





ERK and AKT activation within NRAS^{Q61K} Mutant A375 Isogenic Cell Line

Non-specific chemotherapy drug treatment

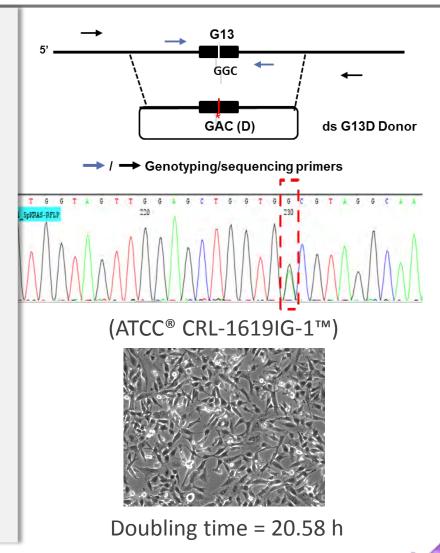




Generation and validation of the KRAS^{G13D} mutant A375 isogenic cell line

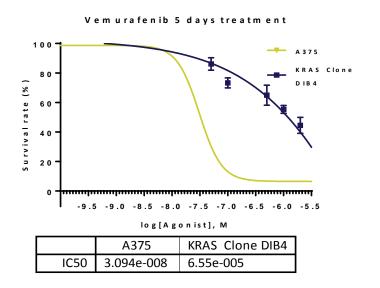
KRASG13D isogenic A375 melanoma line (ATCC® CRL-1619IG-1™)

- Sequence gDNA to confirm KRASG13D knock-in
- mRNA level validation
- Off-target screening
- Cell morphology
- Cell growth kinetics
- Cell STR profile
- Cell response to therapeutics

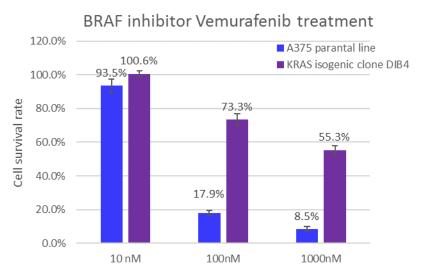


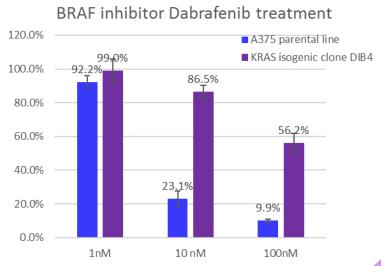


KRAS^{G13D} mutant A375 Cell Line exhibits significant resistance to BRAF inhibitors



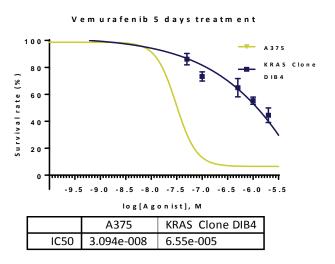
Dabrafenib 5 days treatment									
	1 0	0 7				375			
_	8	o –		1	、 	KRAS	Clon		
rate (%)	6	o –		_	1	DIB 4			
	4	o -		Ĭ					
Survival									
9.5 -9.0 -8.5 -8.0 -7.5 -7.0 -6.5 -6.0 -5.5 -5.0									
log[Agonist], M									
				A375	KRAS Clone DIB4	4			
			IC50	4.056e-009	1.184e-007				
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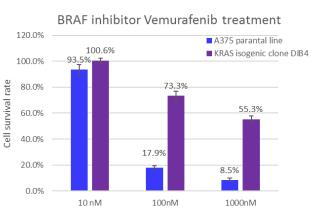


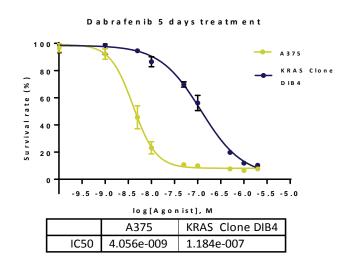


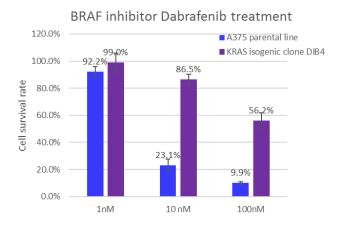


KRAS^{G13D} mutant A375 Cell Line exhibits significant resistance to BRAF inhibitors









The KRAS isogenic cell line is more resistant to BRAF inhibitors than the parental A375 cell line. A375 cells (ATCC® CRL-1619™ and KRAS Mutant-A375 Isogenic Cells (ATCC® CRL-1619IG-1™) were treated with the indicated concentrations of either dabrafenib of vemurafenib for three days. Cell survival was monitored via cell viability assay.



Use of CRISPR system to create cancer disease models

ATCC precision cell models for the development of new anti-cancer therapeutics

■EML4-ALK fusion A549 (ATCC® CCL-185IG™) **Now Available!**



Driver gene mutations

- IDH1^{R132H} mutant U-87 (ATCC® HTB-14IG™)
- IDH2R140Q mutant TF-1 (ATCC® CRL-2003IG™)

- NRAS^{Q61K} mutant A375 (ATCC[®] CRL-1619IG-1™)
- KRAS^{G13D} mutant A375 (ATCC® CRL-1619IG-2™)



Drugresistant mutants

Conclusion

Clinically relevant cancer cell models are critical both for studies of molecular and cellular mechanisms of tumorigenesis and for the design and screening of novel cancer therapeutics. With new genome editing tools such as CRISPR/Cas9, ATCC can now use its extensive cell-banking resources to generate novel isogenic disease model cell lines. We have engineered isogenic lines with mutations in key oncogenes that are ideally suited for the identification of novel, personalized treatment regimens.

Key features of ATCC isogenic cell lines:

- Parental lines are carefully selected to be highly relevant to diseases and drug targets
- Precisely edited isogenic cell lines have been thoroughly validated at genomic, transcript, and protein levels
- Additional bio-functional characterization with specific inhibitors to demonstrate drugscreening applicability
- Together with authenticated parental lines, CRISPR/Cas9-edited isogenic lines provide useful in vitro models for both basic and translational research



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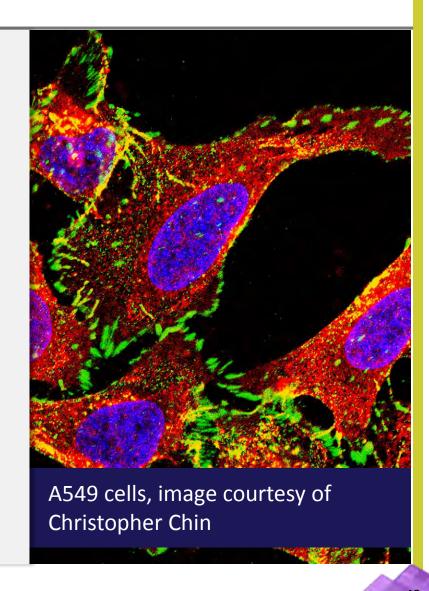
Monica Wood





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