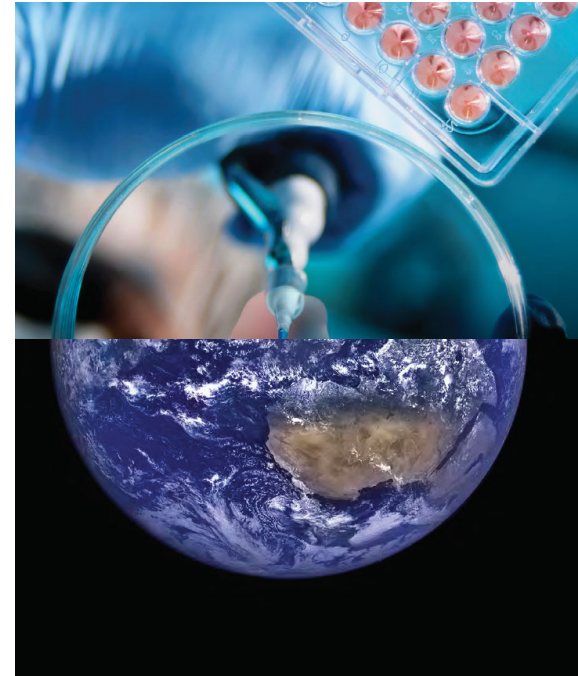
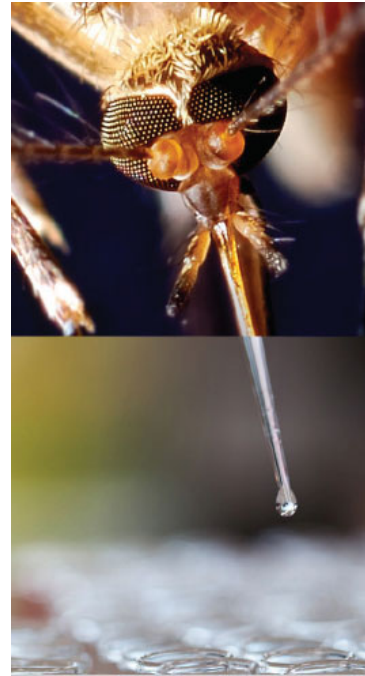
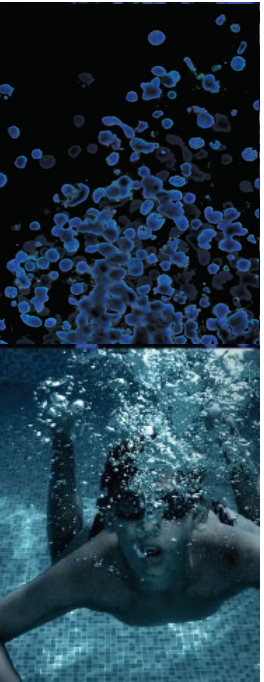




Skin Microbiome: Considerations, Applications, and Future Directions

Tasha M. Santiago-Rodriguez, PhD
Data Scientist III, Diversigen, Inc.

Credible Leads to Incredible™



About ATCC

- Founded in 1925, ATCC is a non-profit organization with HQ in Manassas, VA, and an R&D and Services center in Gaithersburg, MD
- World's largest, most diverse biological materials and information resource for cell culture – the “*gold standard*”
- Innovative R&D company featuring gene editing, microbiome, NGS, advanced models
- cGMP biorepository
- Partner with government, industry, and academia
- Leading global supplier of authenticated cell lines, viral and microbial standards
- Sales and distribution in 150 countries, 19 international distributors
- Talented team of 450+ employees, over one-third with advanced degrees



The skin microbiome: Considerations, applications and future directions

Tasha M. Santiago-Rodriguez, Ph.D.
Diversigen, Houston, TX
R&D Data Scientist III



Agenda

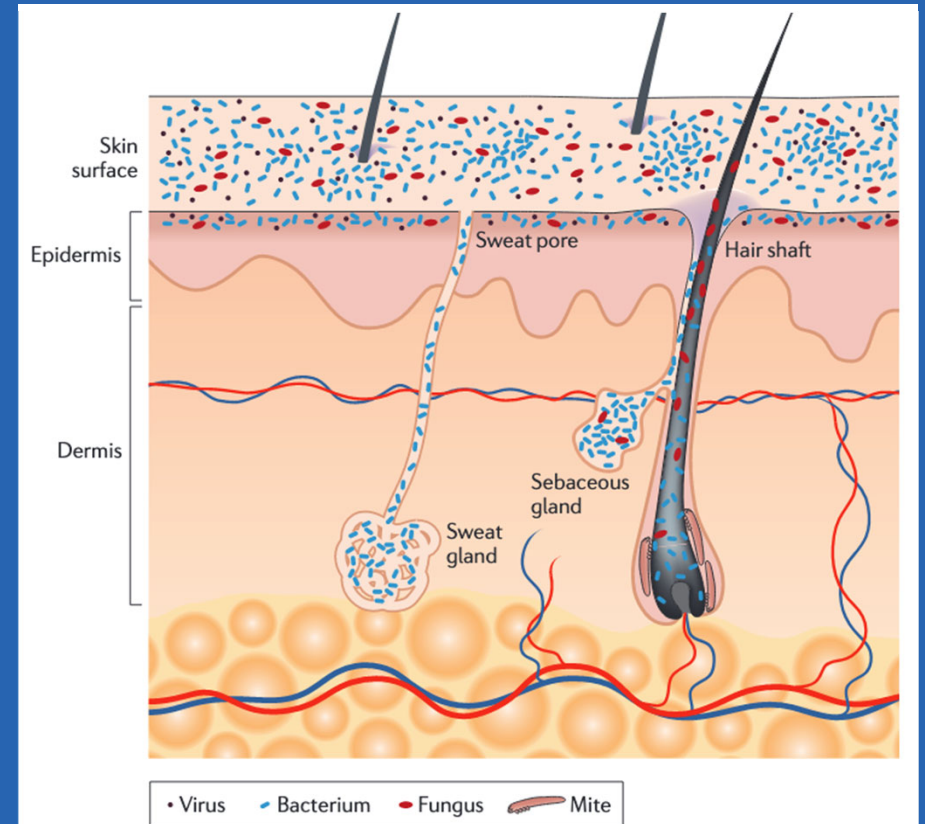
- Introduction to the skin microbiome
- Biases in skin microbiome research
 - Collection
 - Extraction and amplification
 - Database and annotation
- Future directions in skin microbiome research



Introduction to the skin microbiome

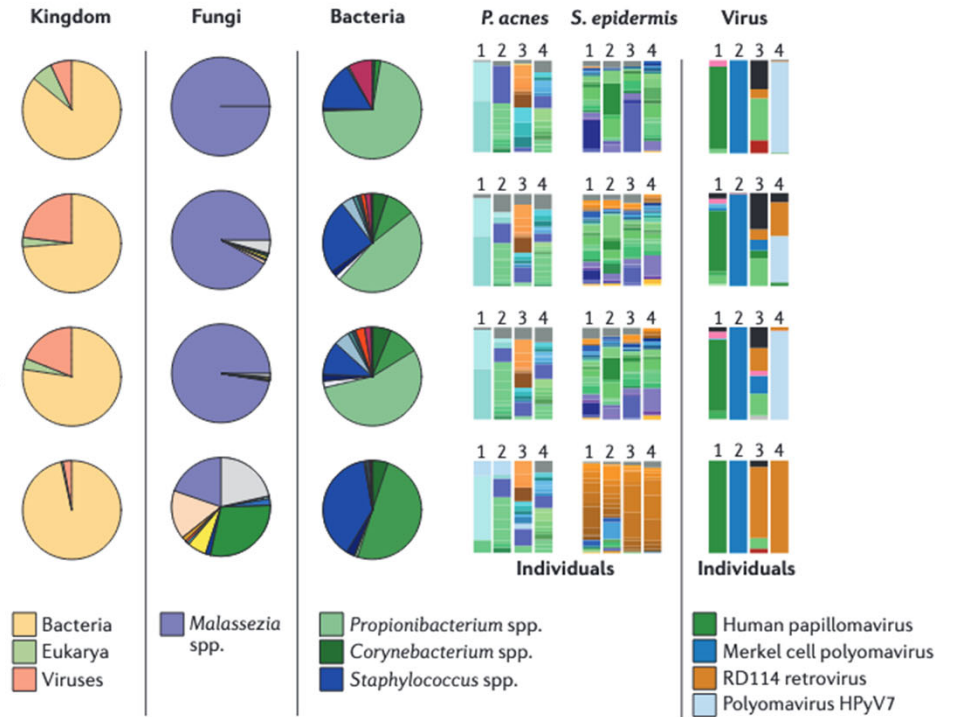
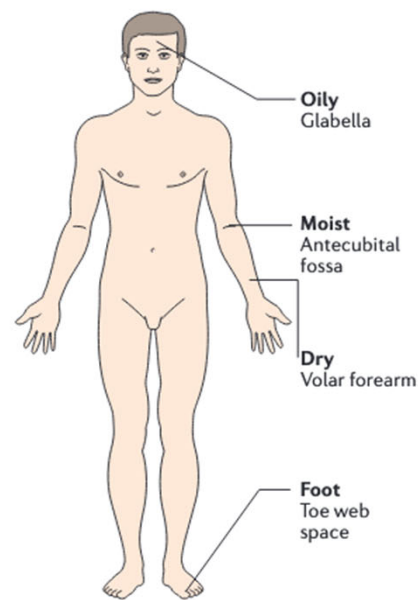
Skin is an ecosystem

- As the largest human organ, skin serves as a physical barrier and acts as an ecosystem with diverse micro environments (pH, light exposure, moisture and oil content)
- Bacteria fungi and viruses inhabit skin
- Most of these microorganisms are harmless, providing protection and modulation of the immune system



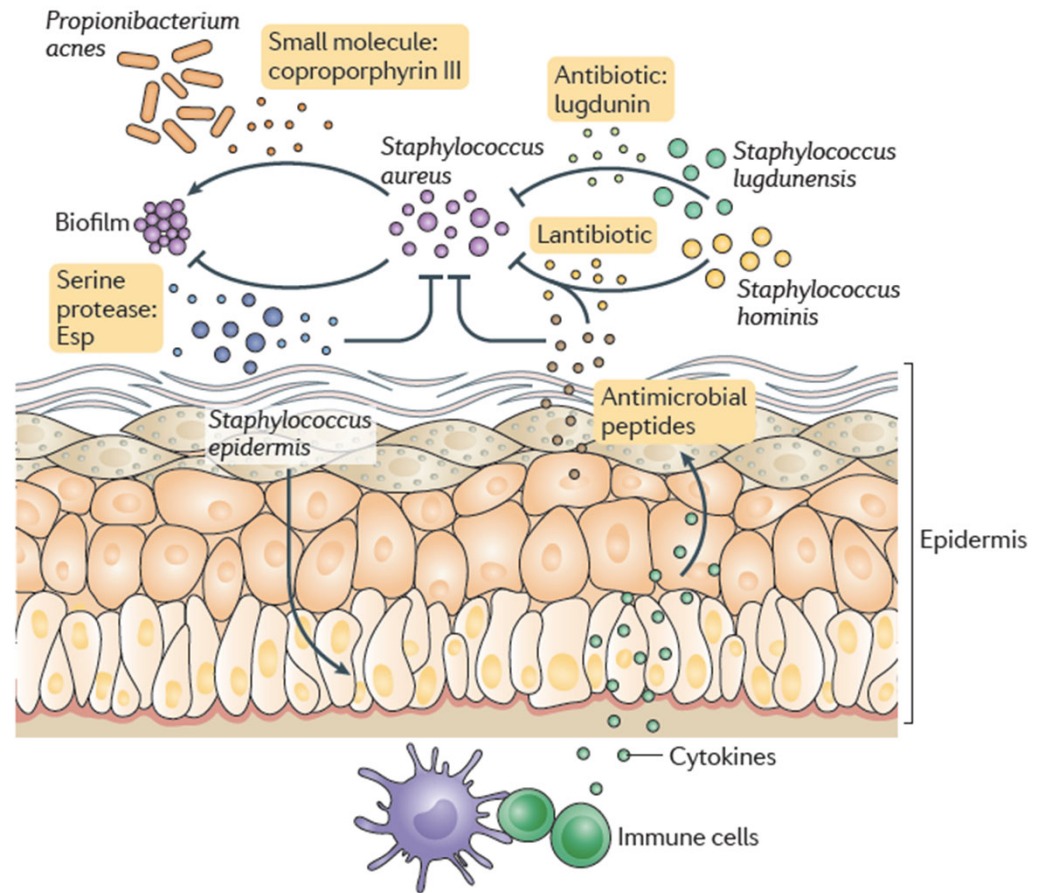
Grice, E. A., & Segre, J. A. (2011). The skin microbiome. *Nature reviews microbiology*, 9(4), 244-253.

Skin contains diverse micro environments



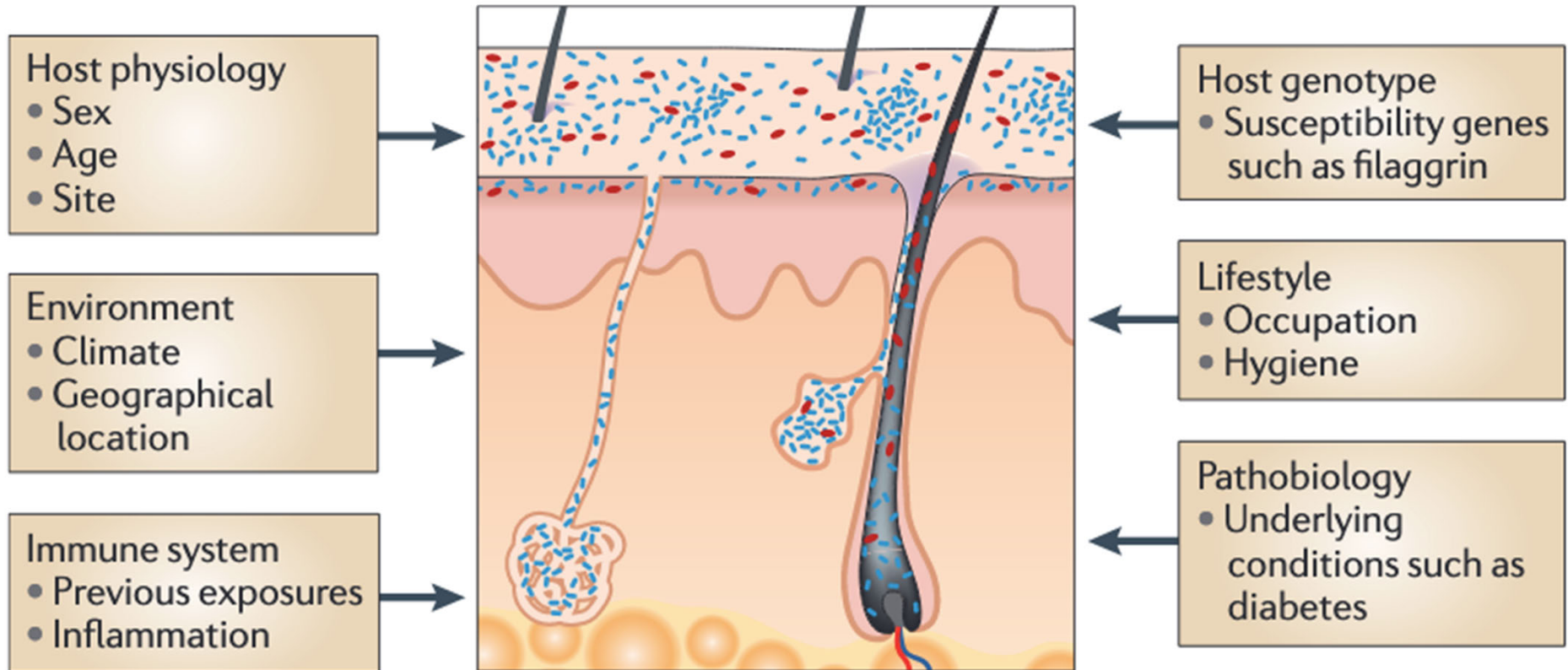
Left: Eisenstein, M. (2020). The skin microbiome and its relationship with the human body explained. *Nature*, 588(7838), S211-S211.
Right: Byrd, A. L., Belkaid, Y., & Segre, J. A. (2018). The human skin microbiome. *Nature Reviews Microbiology*, 16(3), 143-155.

The skin microbiome protects us from pathogens and modulates the immune system



Byrd, A. L., Belkaid, Y., & Segre, J. A. (2018). The human skin microbiome. *Nature Reviews Microbiology*, 16(3), 143-155.

Different factors are associated with the skin microbiome

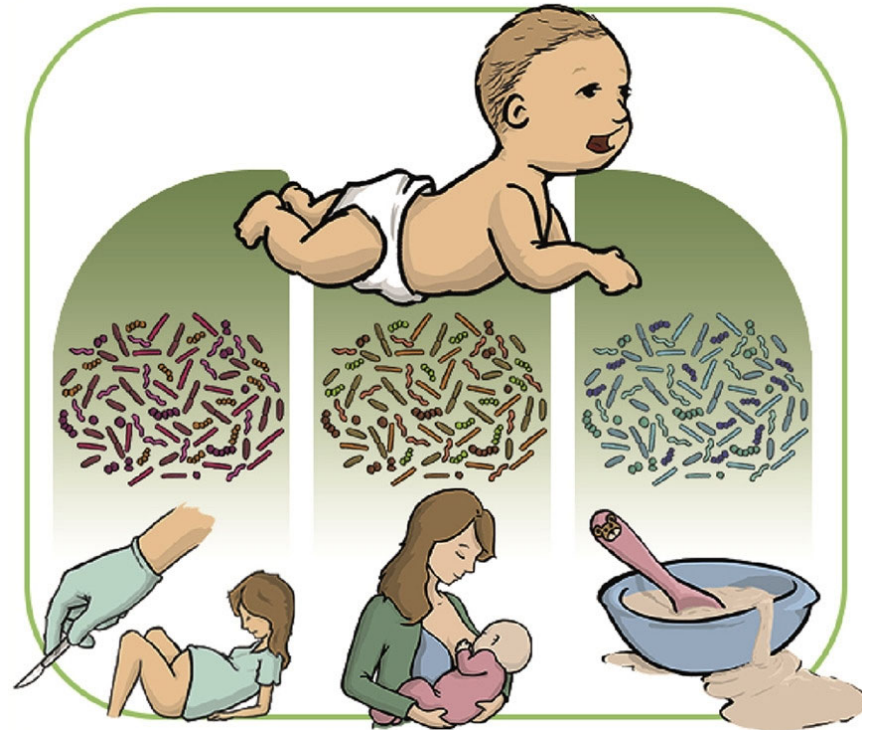


Grice, E. A., & Segre, J. A. (2011). The skin microbiome. *Nature reviews microbiology*, 9(4), 244-253.

The skin microbiome is acquired at birth

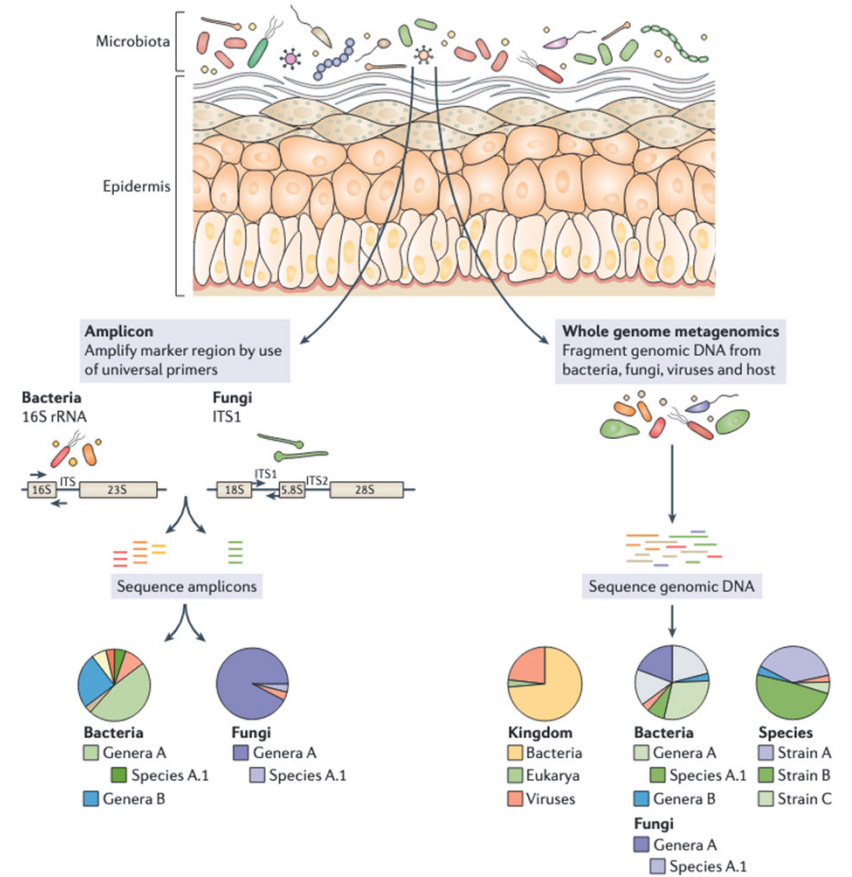
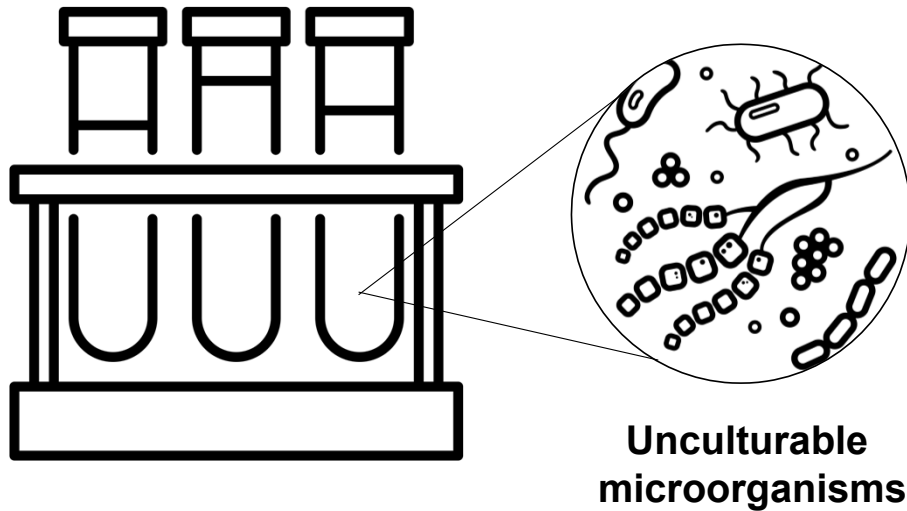
Skin microbiome composition at birth and early on in life is influenced by:

- Delivery mode
- Maternal microbiota
- Antibiotic use
- Soaps and detergents
- Nutritional factors
- Housing
- Animal contact
- Outdoor play



Galazzo, G., van Best, N., Bervoets, L., Dapaah, I. O., Savelkoul, P. H., Hornef, M. W., ... & Penders, J. (2020). Development of the microbiota and associations with birth mode, diet, and atopic disorders in a longitudinal analysis of stool samples, collected from infancy through early childhood. *Gastroenterology*, 158(6), 1584-1596.

Culture-independent methods for studying the skin microbiome



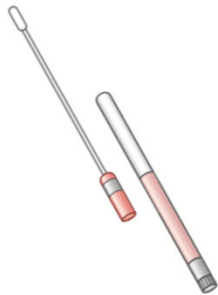
Left: <http://teachthemicrobiome.weebly.com/uploads/2/6/4/3/26438968/1392600741.jpg>

Right: Byrd, A. L., Belkaid, Y., & Segre, J. A. (2018). The human skin microbiome. *Nature Reviews Microbiology*, 16(3), 143-155.

Skin microbiome pipeline: from sample collection to analysis



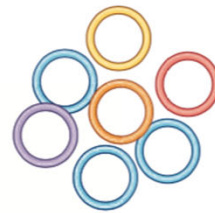
Sample
processing
steps



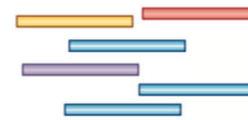
Sample
collection



Sample
storage



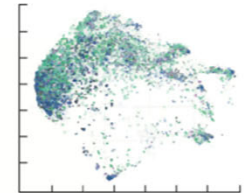
DNA
extraction



Sequencing
library preparation

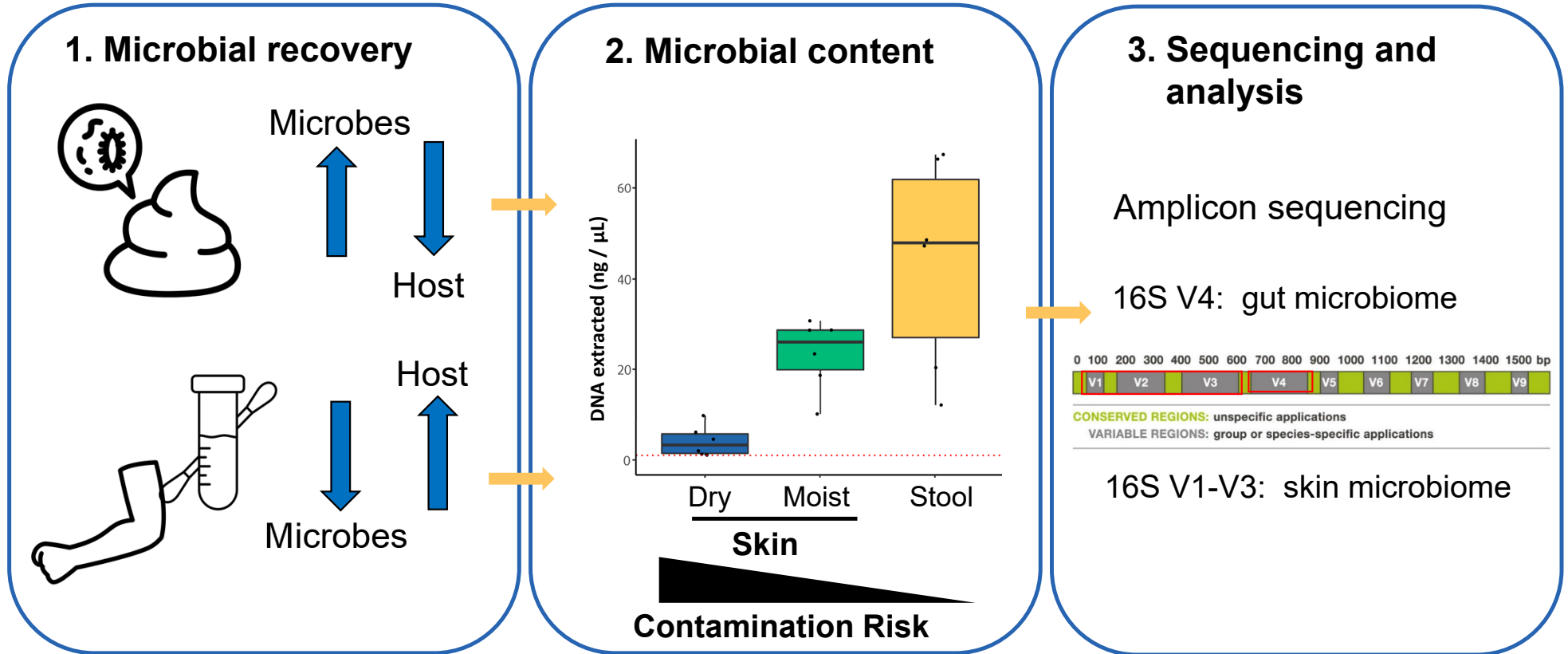


DNA
sequencing

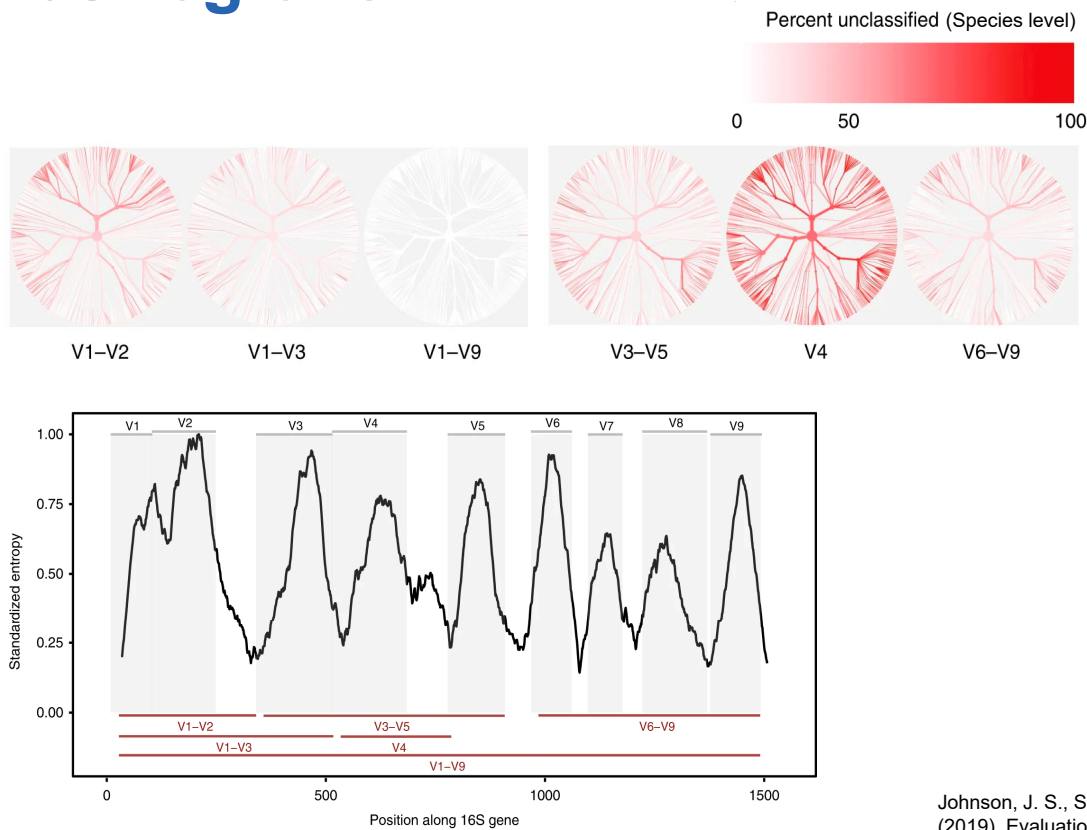


Computational
analysis

Considerations for studying the skin microbiome



Trade-offs between primers targeting V4 and V1-V3 16S regions



V4 16S region

- Inability to capture certain species without primer editing
- Ability to compare to a broad literature and across body sites
- Improved sequence quality

V1-V3 16S region

- Challenges in bioinformatic analysis due to long amplicon
- Improved species-level resolution for skin samples
- Better correlation with whole genome sequencing results

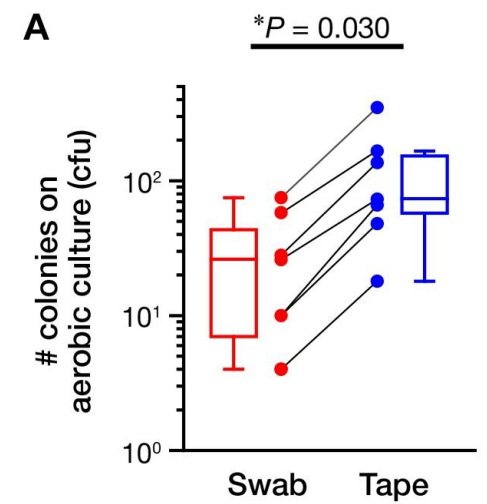
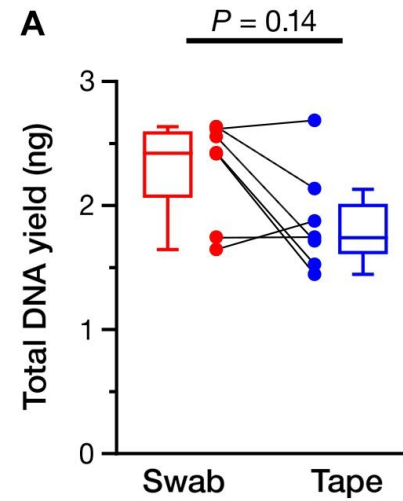
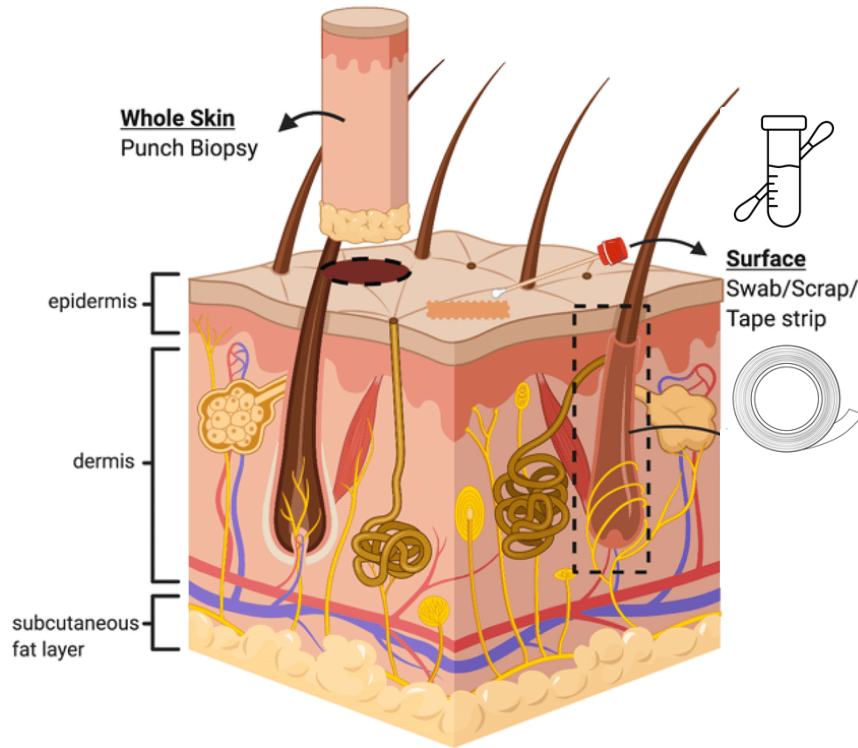
Johnson, J. S., Spakowicz, D. J., Hong, B. Y., Petersen, L. M., Demkowicz, P., Chen, L., ... & Weinstock, G. M. (2019). Evaluation of 16S rRNA gene sequencing for species and strain-level microbiome analysis. *Nature communications*, 10(1), 1-11.

Biases in skin microbiome r

Sample collection



Human skin microbiome collection methods



Left: McLoughlin, I. J., Wright, E. M., Tagg, J. R., Jain, R., & Hale, J. D. (2021). Skin Microbiome—The Next Frontier for Probiotic Intervention. *Probiotics and Antimicrobial Proteins*, 1-18.

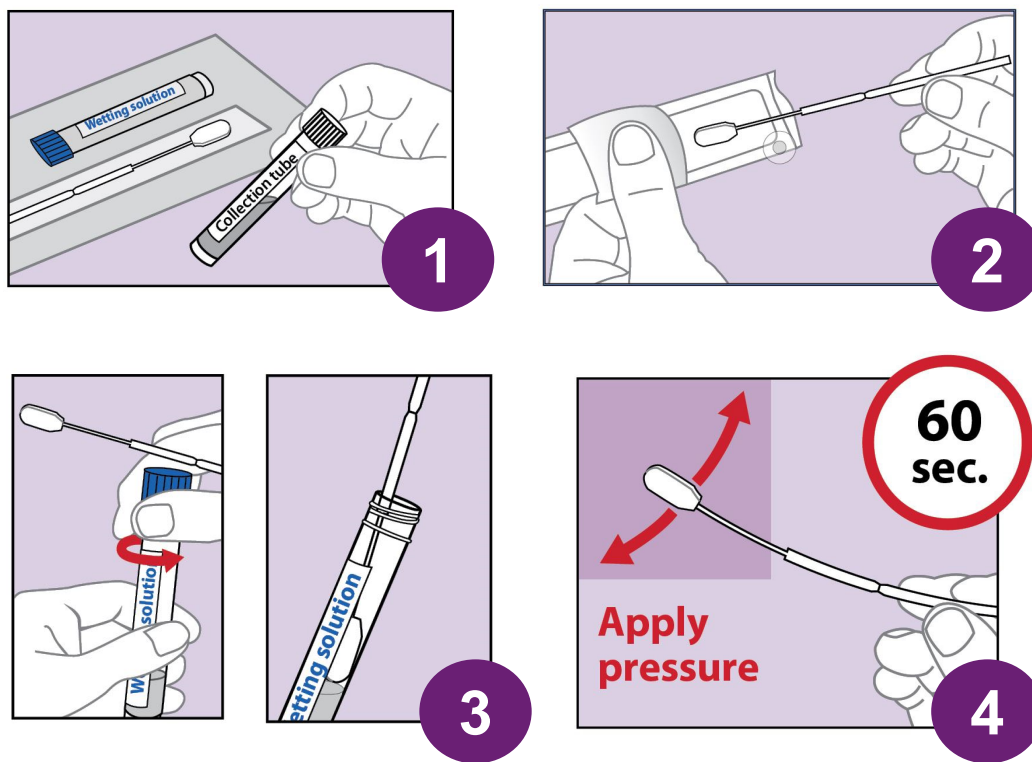
Right: Ogai, K., Nagase, S., Mukai, K., Iuchi, T., Mori, Y., Matsue, M., ... & Okamoto, S. (2018). A comparison of techniques for collecting skin microbiome samples: swabbing versus tape-stripping. *Frontiers in microbiology*, 2362.



A swab-based microbiome collection device



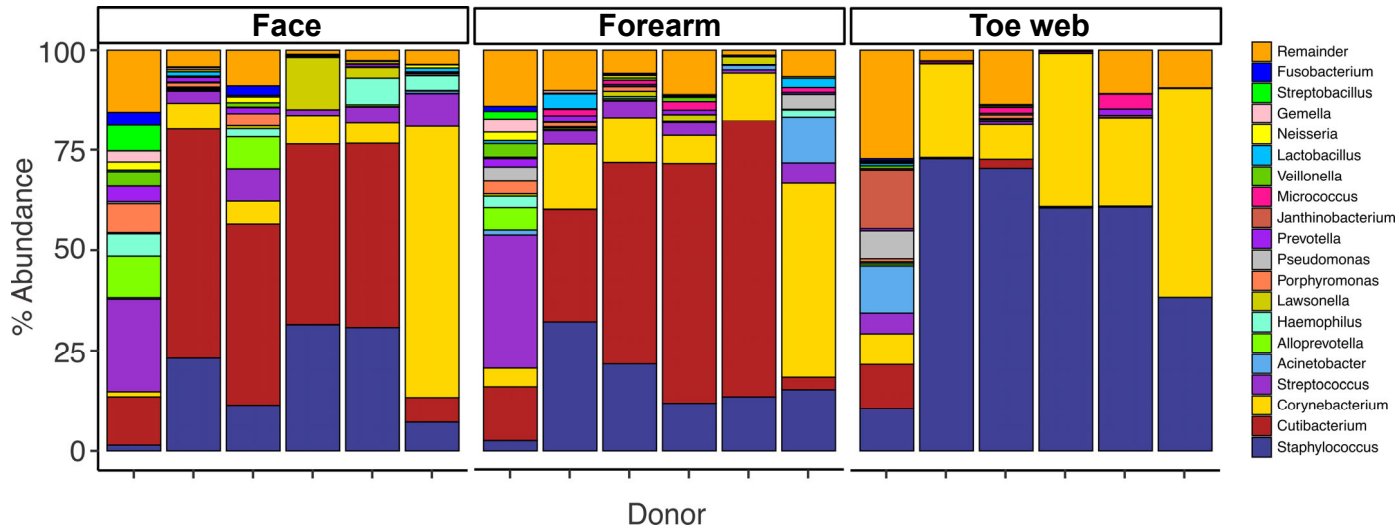
Optimized Instructions for Use (IFU)



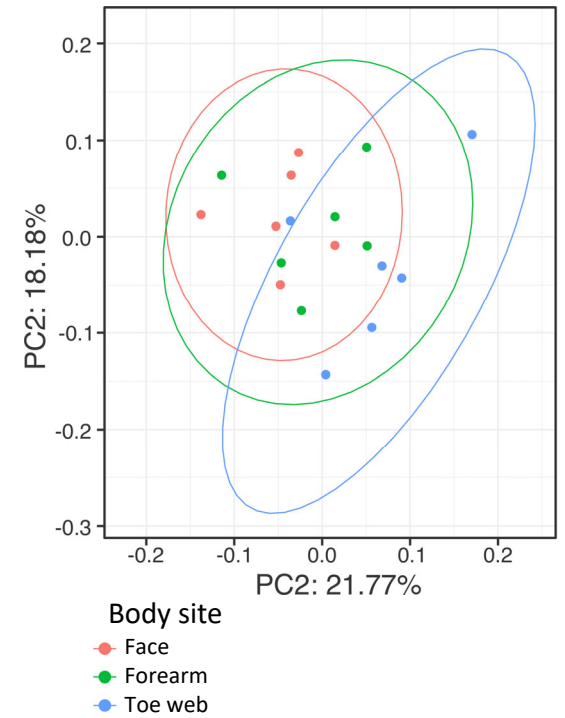


Capture site-specific bacterial profiles

V1-V3 16S taxonomic profiles across skin sites



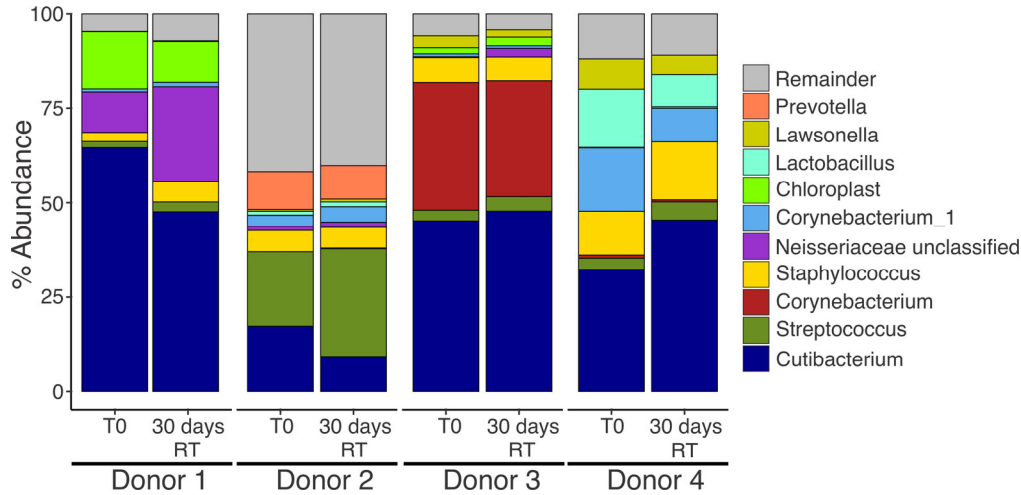
PCoA plot of skin microbiome body site profiles



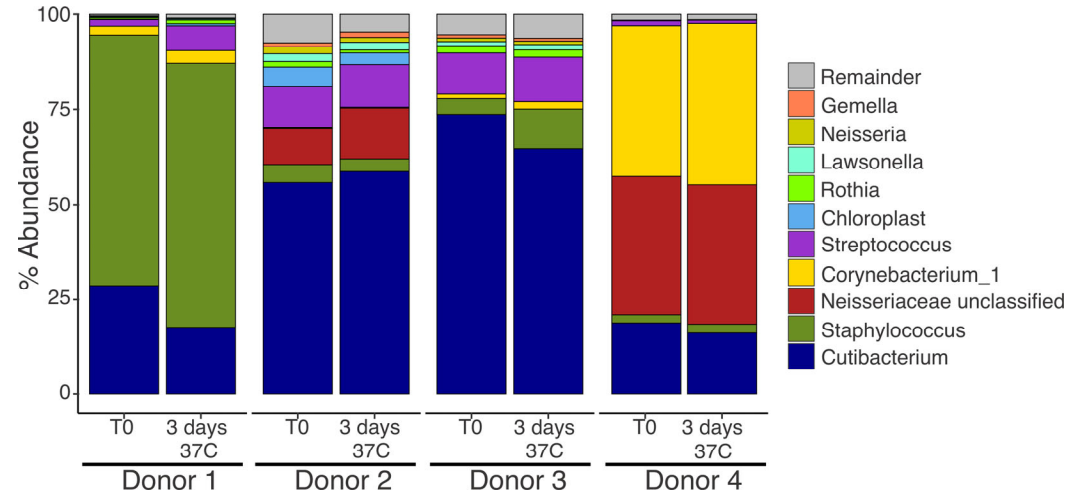


Stabilize the skin microbial profile during shipping and storage

Room temperature, 30 days



37°C, 3 days



Considerations for selecting a skin microbiome collection device

- Validated across various skin sites (i.e., dry, oily and moist)
- Optimized IFU for improved collection/sample performance
- Low bioburden
- Captures and stabilizes the skin microbiome during storage or shipping at ambient temperatures
- Consistent performance



Biases in skin microbiome r

Extraction and amplification



DNA extraction can add biases to skin microbiome studies

- Commercial DNA extraction kits are the most common for skin microbiome analyses
- Different approaches exist, including enzymatic treatment and/or mechanical lysis
- DNA concentrations and success of library preparation are associated with extraction method

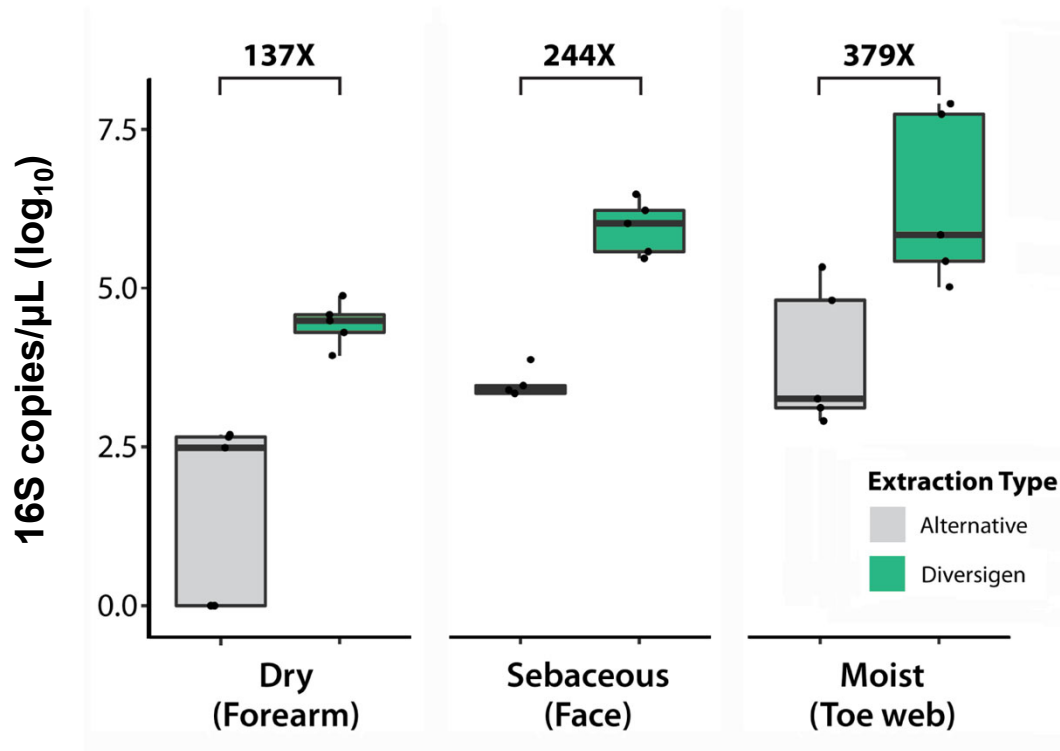
Mechanical + enzymatic treatment
(high salt)

Mechanical + enzymatic treatment

Kit number	DNA concentration (ng/μl) Skin samples average
1	0.05
2	0.02
3	0.01
4	0.00
5	0.00
6	0.03
7	0.02
8	0.99
9	0.01
10	0.03
11	0.02
12	0.04

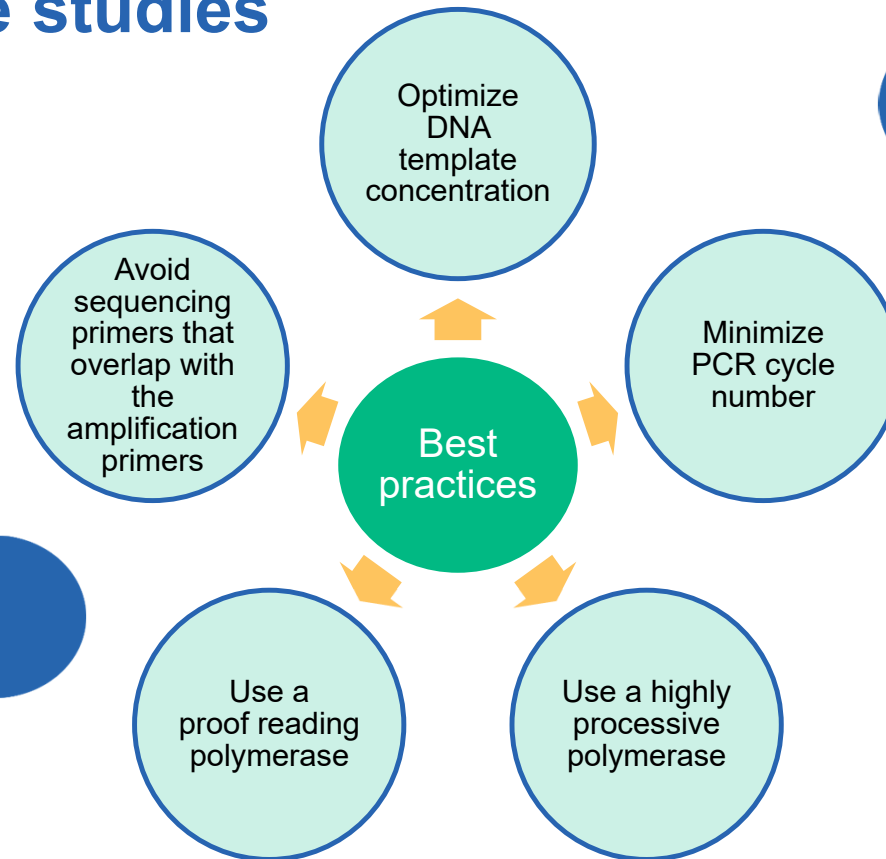
Modified from Bjerre, R. D., Hugerth, L. W., Boulund, F., Seifert, M., Johansen, J. D., & Engstrand, L. (2019). Effects of sampling strategy and DNA extraction on human skin microbiome investigations. *Scientific reports*, 9(1), 1-11.

DNA extraction optimization is essential for skin microbiome studies



- Optimized collection kits and extraction workflows maximize DNA yields from low biomass sites
- Increases rate of successful collection (fewer sample dropouts)
- Ensured success of downstream applications, including amplicon generation

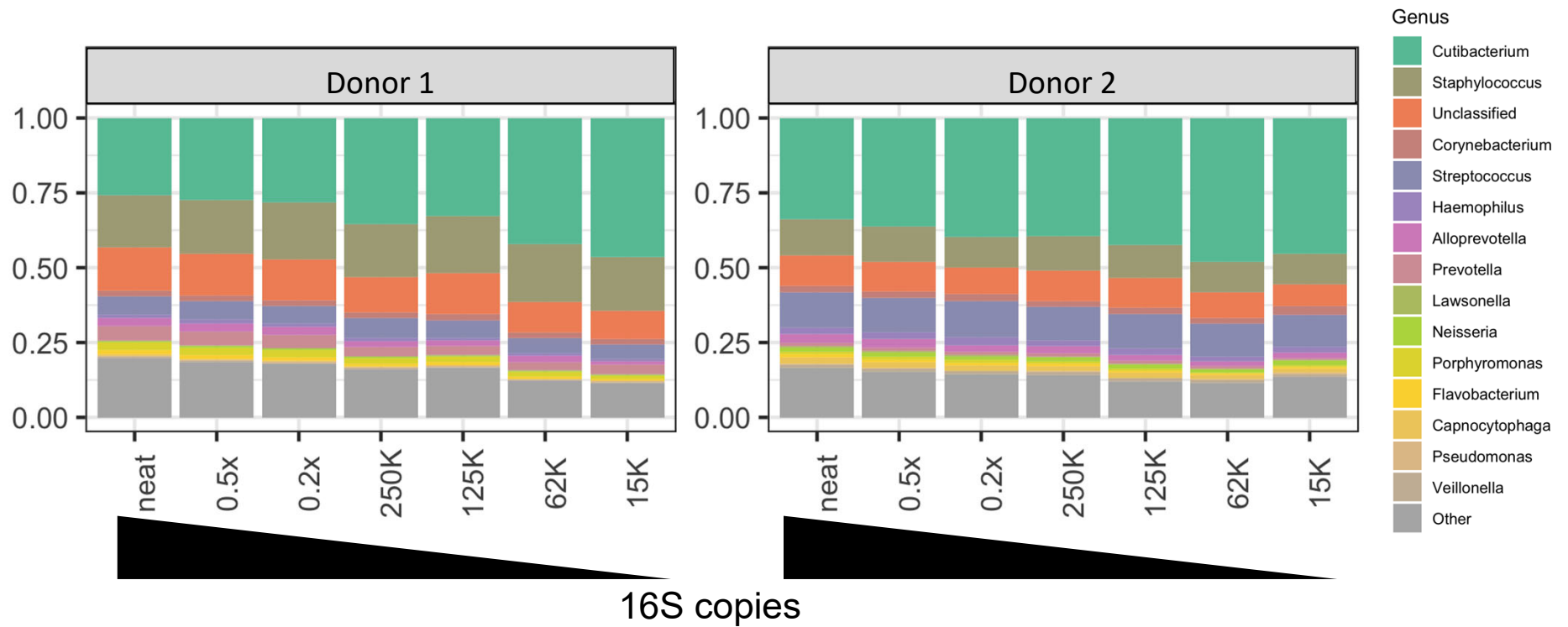
Best practices for generating amplicons for microbiome studies



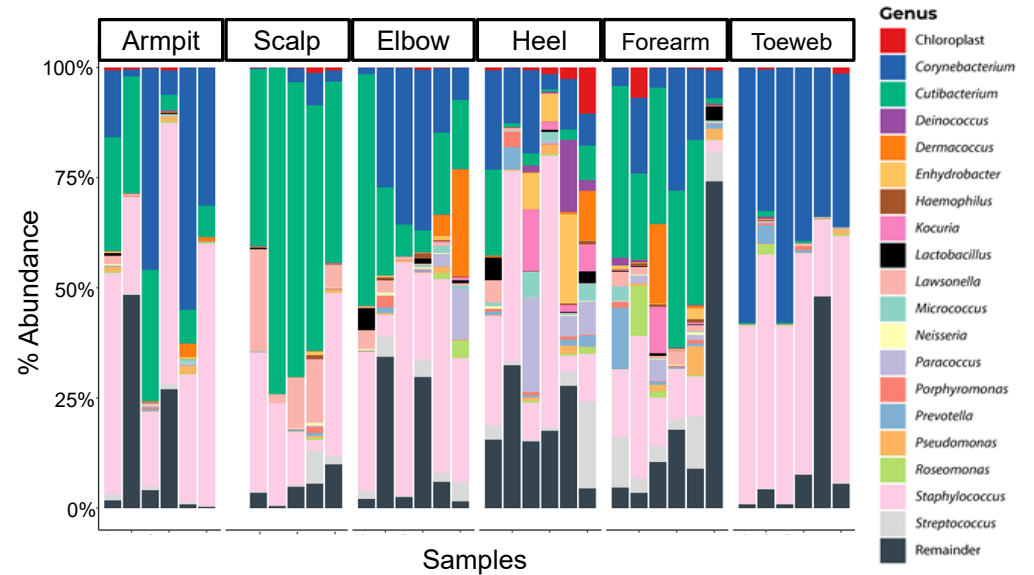
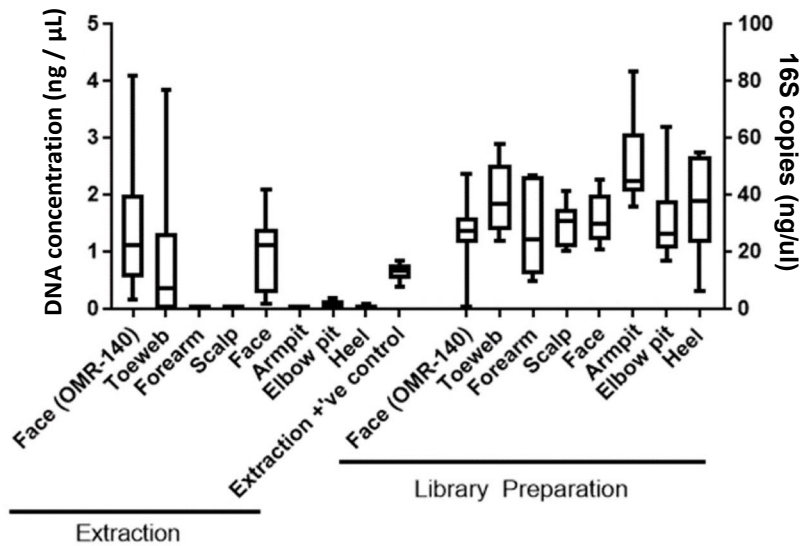
Improved quantitative accuracy

Improved qualitative accuracy

Reproducible skin profiles were generated, regardless of sample concentration



Library preparation provides consistent DNA concentrations for sequencing and bioinformatic analyses



- Sufficient library yields are achieved, regardless of extracted biomass
- Unique microbial profiles can be generated across various skin sites

Considerations for skin sample processing

- Validated across various skin sites (i.e., dry, oily and moist)
- Optimized to maximize DNA yields and 16S rRNA gene copy number
- Capture expected taxonomic profiles of the skin microbiome across a range of DNA concentrations
- Provides consistent performance, which will depend on the number of PCR cycles, polymerases, as well as amplification and sequencing primers



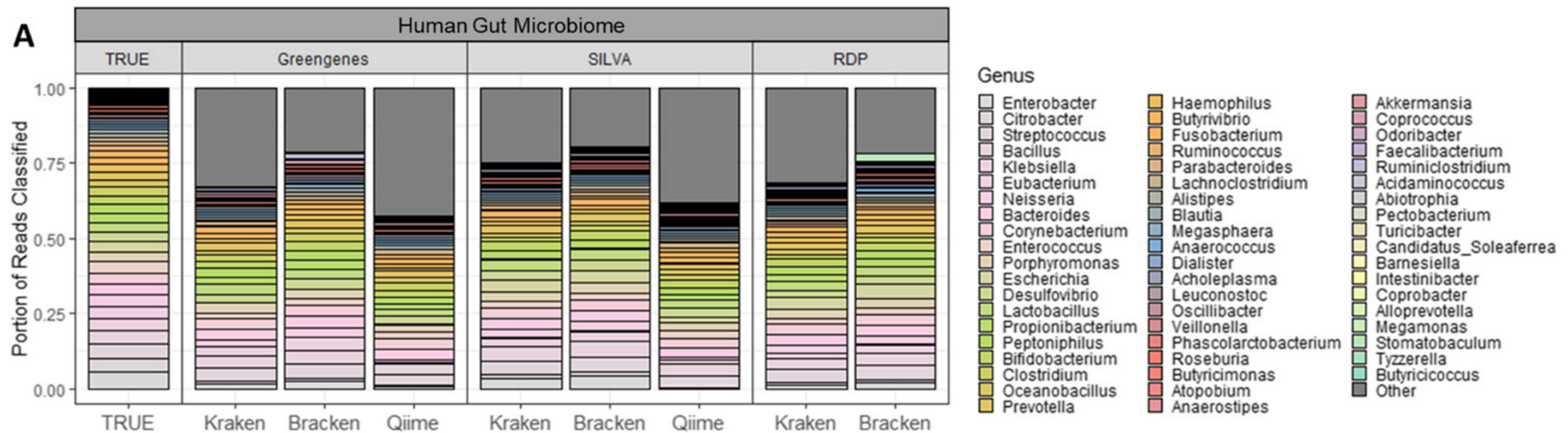
Biases in skin microbiome r

Database and annotation



Effects of different databases and taxonomic annotations on microbiome

- Annotation tools are known to affect the taxonomic composition of the human microbiome

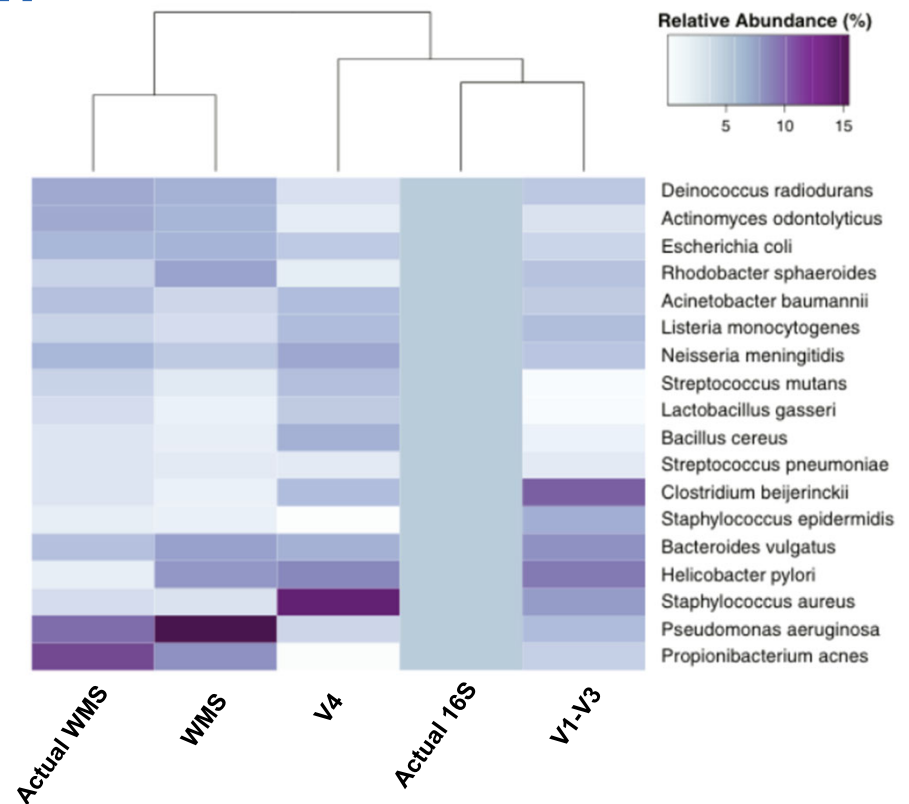


- Limited literature on the effect of different annotation tools on the skin microbiota composition

Lu, J., & Salzberg, S. L. (2020). Ultrafast and accurate 16S rRNA microbial community analysis using Kraken 2. *Microbiome*, 8(1), 1-11.

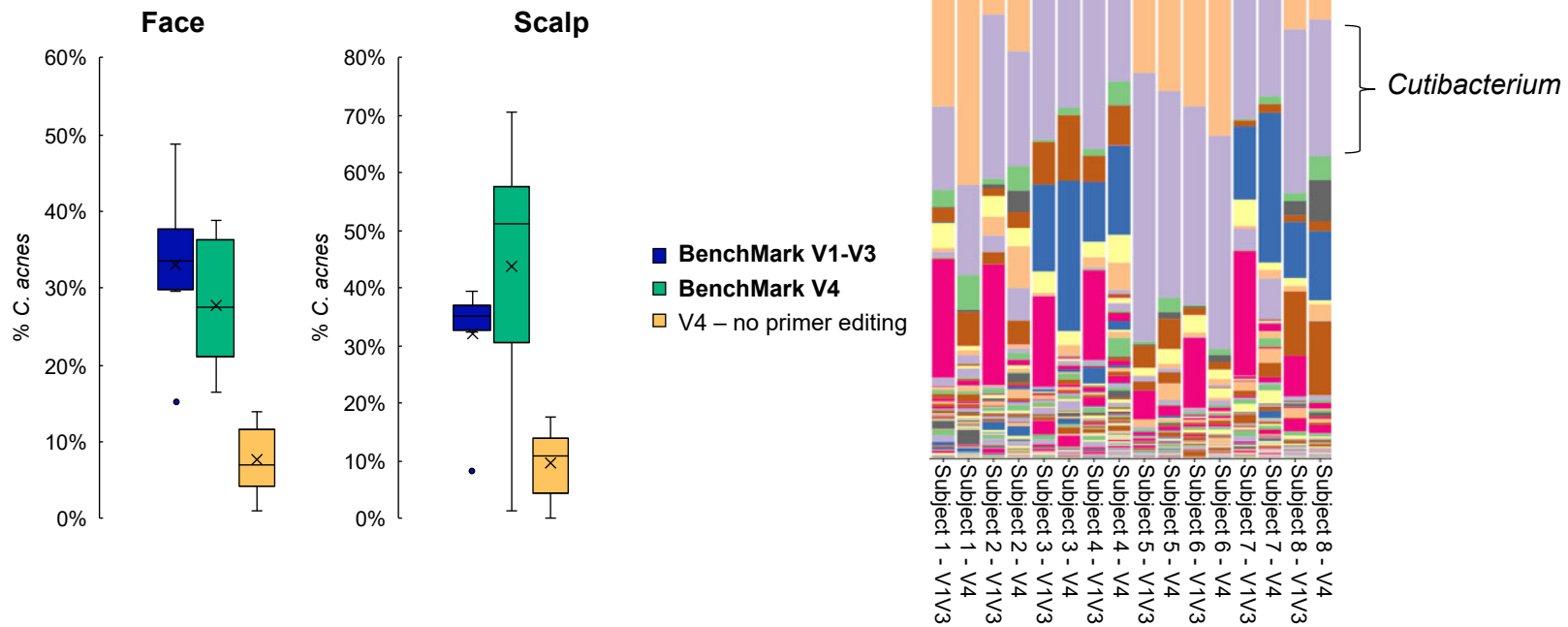
Amplification of 16S rRNA gene V4 vs. V1-V3 regions affects taxonomic classification

- Amplification of 16S V4 region resulted in an over representation of *Staphylococcus aureus* and an under-representation of *Cutibacterium acnes* and *Staphylococcus epidermidis*
- Amplification of the 16S V1-V3 region better recapitulated the expected proportions of mock communities



Meisel, J. S., Hannigan, G. D., Tyldsley, A. S., SanMiguel, A. J., Hodkinson, B. P., Zheng, Q., & Grice, E. A. (2016). Skin microbiome surveys are strongly influenced by experimental design. *Journal of Investigative Dermatology*, 136(5), 947-956.

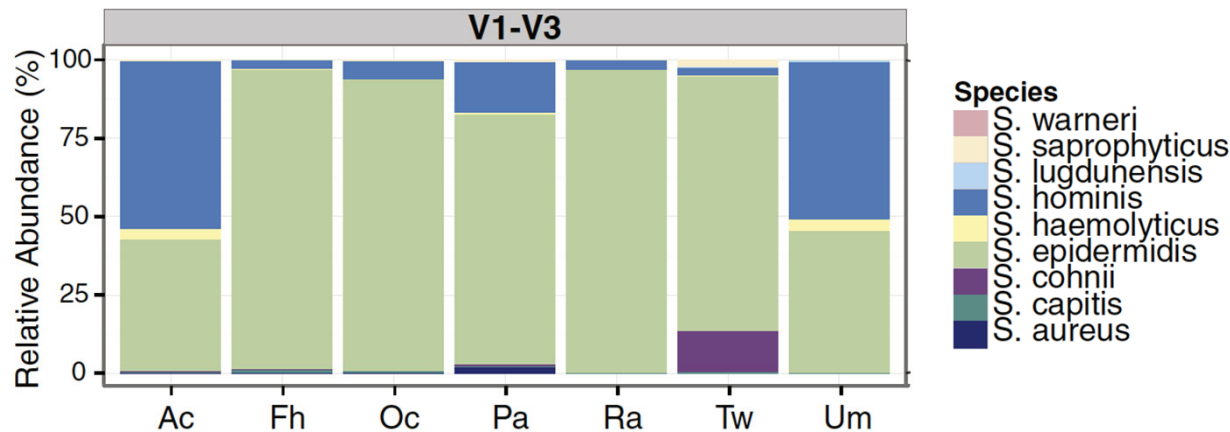
Primer editing targeting the 16S rRNA V4 region can improve recovery of *Cutibacterium acnes*



Gohl, D. M., Auch, B., Certano, A., LeFrançois, B., Bouevitch, A., Doukhanine, E., ... & Beckman, K. B. (2021). Dissecting and tuning primer editing by proofreading polymerases. *Nucleic acids research*, 49(15), e87-e87.

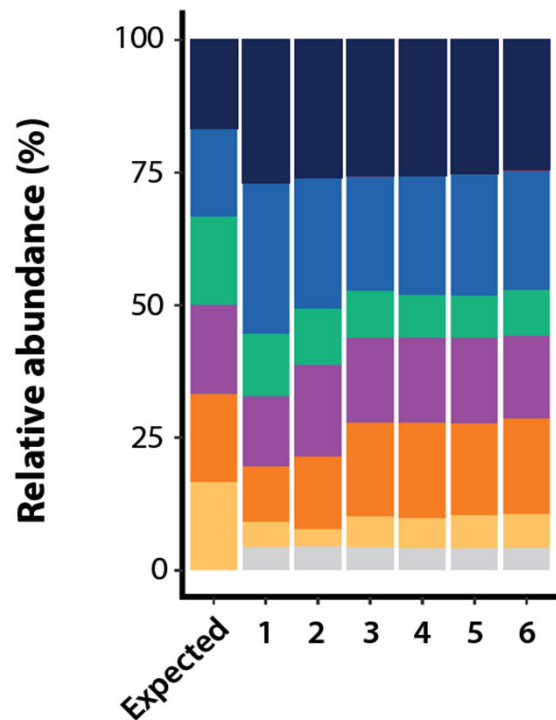
Limitations of 16S rRNA gene amplicon annotations to identify species

- Usually provide genus-level taxonomic resolution



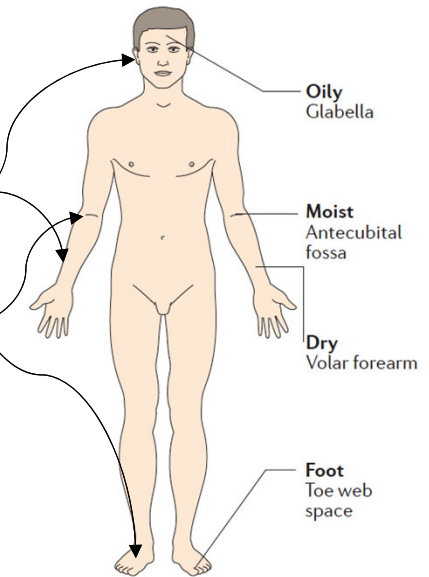
- Species-level resolution of skin microbiota is important
 - Pathogens vs. Commensals
- Phylogenetic placement algorithms have been employed with little success

Leveraging 16S amplicon information to accurately identify species



Bacterial species

- Acinetobacter johnsonii
- Corynebacterium striatum
- Cutibacterium acnes
- Micrococcus luteus
- Staphylococcus epidermidis
- Streptococcus mitis
- Remainder



- All bacterial species in ATCC® MSA-1005 skin mock community were detected across replicates

Accurate species-level resolution among same genera

ATCC® MSA-1002

Expected (ATCC MSA1002)	Alternative	Diversigen
<i>Streptococcus agalactiae</i>	Yellow	Green
<i>Lactobacillus gasseri</i>	Yellow	Green
<i>Cutibacterium acnes</i>	Yellow	Green
<i>Staphylococcus epidermidis</i>	Green	Green
<i>Streptococcus mutans</i>	Green	Green
<i>Staphylococcus aureus</i>	Yellow	Green
<i>Acinetobacter baumannii</i>	Green	Green
<i>Bacteroides vulgatus</i>	Green	Green
<i>Bifidobacterium adolescentis</i>	Green	Green
<i>Clostridium beijerinckii</i>	Yellow	Green
<i>Deinococcus radiodurans</i>	Yellow	Green
<i>Escherichia coli</i>	Green	Green
<i>Neisseria meningitidis</i>	Green	Green
<i>Porphyromonas gingivalis</i>	Yellow	Green
<i>Pseudomonas aeruginosa</i>	Yellow	Green
<i>Helicobacter pylori</i>	Green	Green
<i>Enterococcus faecalis</i>	Yellow	Green
<i>Rhodobacter sphaeroides</i>	Green	Green
<i>Bacillus pacificus</i>	Yellow	Green
<i>Schaalia odontolytica</i> (former <i>Actinomyces</i>)	Yellow	Green

Species-level identification
Genus-level identification
Not identified

	Alternative	Diversigen
Classified species	7/20	18/20
Unclassified species	12/20	1/20
False negatives	1/20	1/20

Toe web

Alternative	Diversigen
<i>Prevotella colorans</i>	<i>Prevotella colorans</i>
<i>Prevotella buccalis</i>	<i>Prevotella buccalis</i>
<i>Cutibacterium unclassified</i>	<i>Cutibacterium acnes</i>
<i>Corynebacterium unclassified</i>	<i>Corynebacterium tuberculostearicum</i>
<i>Methylobacterium-Methylorubrum</i>	<i>Methylobacterium brachiatum</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus capitis</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus pettenkoferi</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus caprae</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus epidermidis</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus hominis</i>
<i>Staphylococcus unclassified</i>	<i>Staphylococcus lugdunensis</i>

Considerations for selection of databases and annotation tools for skin microbiome research

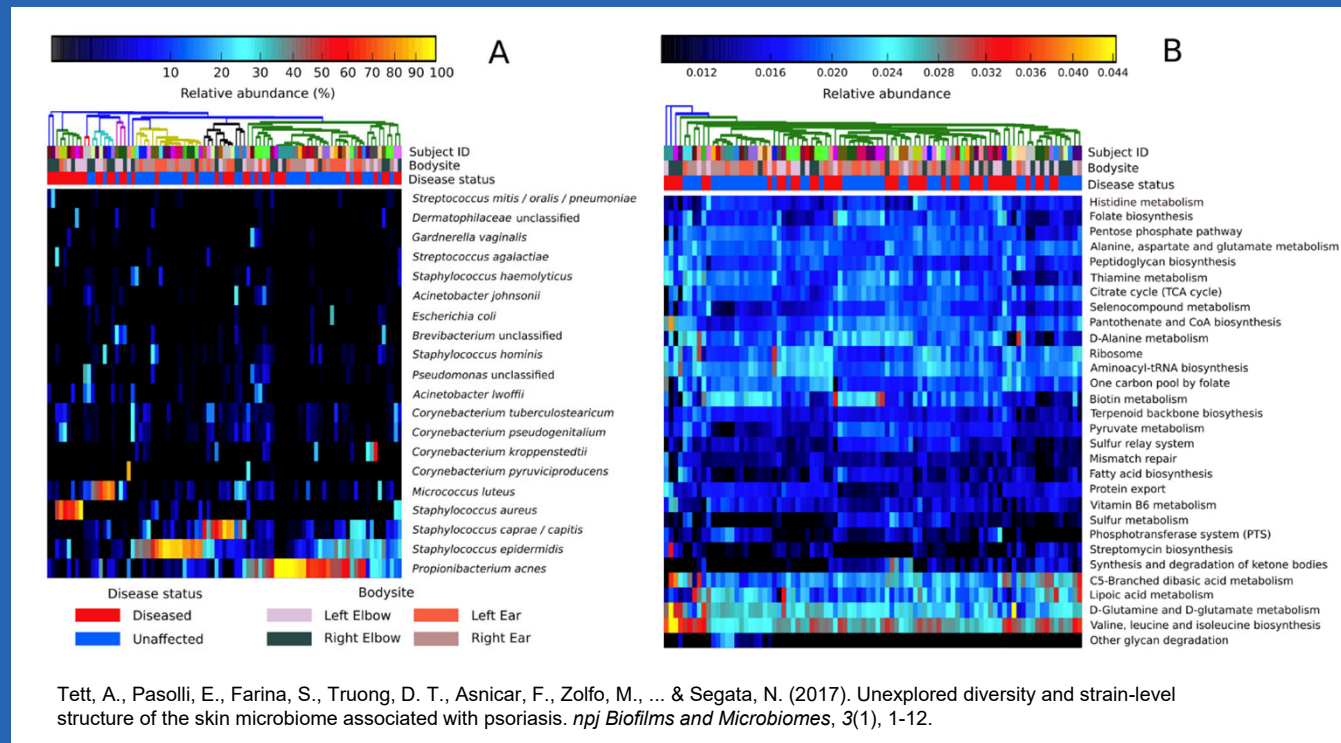
- Databases should be updated and curated on a regular basis
- Databases should provide reliable classifications at the genus and species level when possible
- Annotation tools should provide consistent results across skin sites and conditions
- Number of false positives, false negatives and ambiguous classifications should be minimized



Next steps and future directions in skin microbiome research

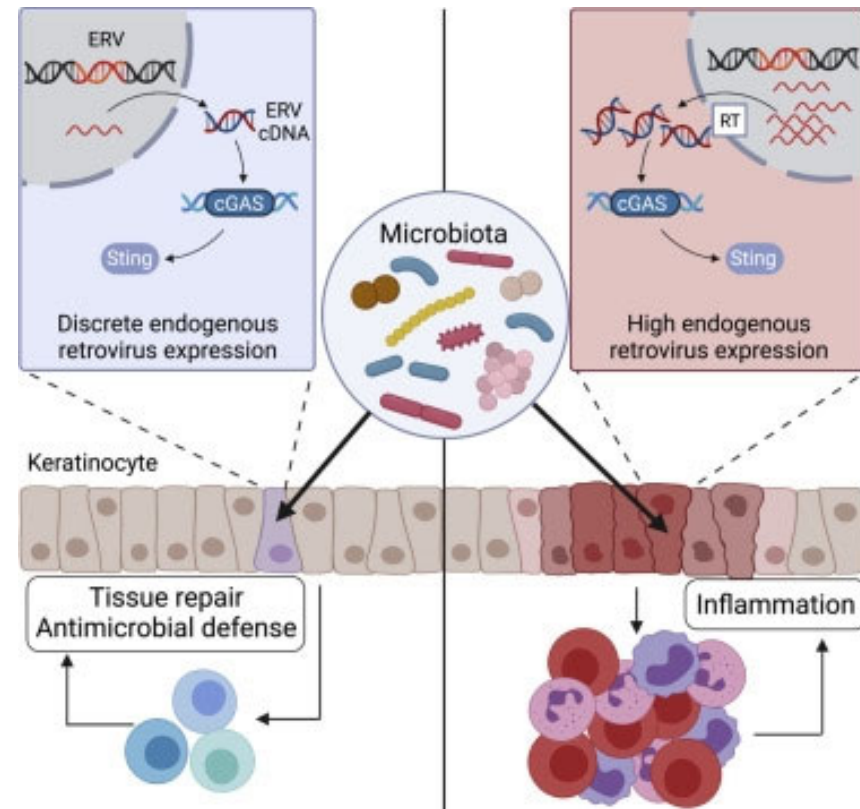
Standardization of whole genome sequencing methods

- Enable bacterial strain identification and identification of other microbes (e.g., viruses, fungi)
- Identify genes with functions related to metabolism, antibiotic resistance and virulence



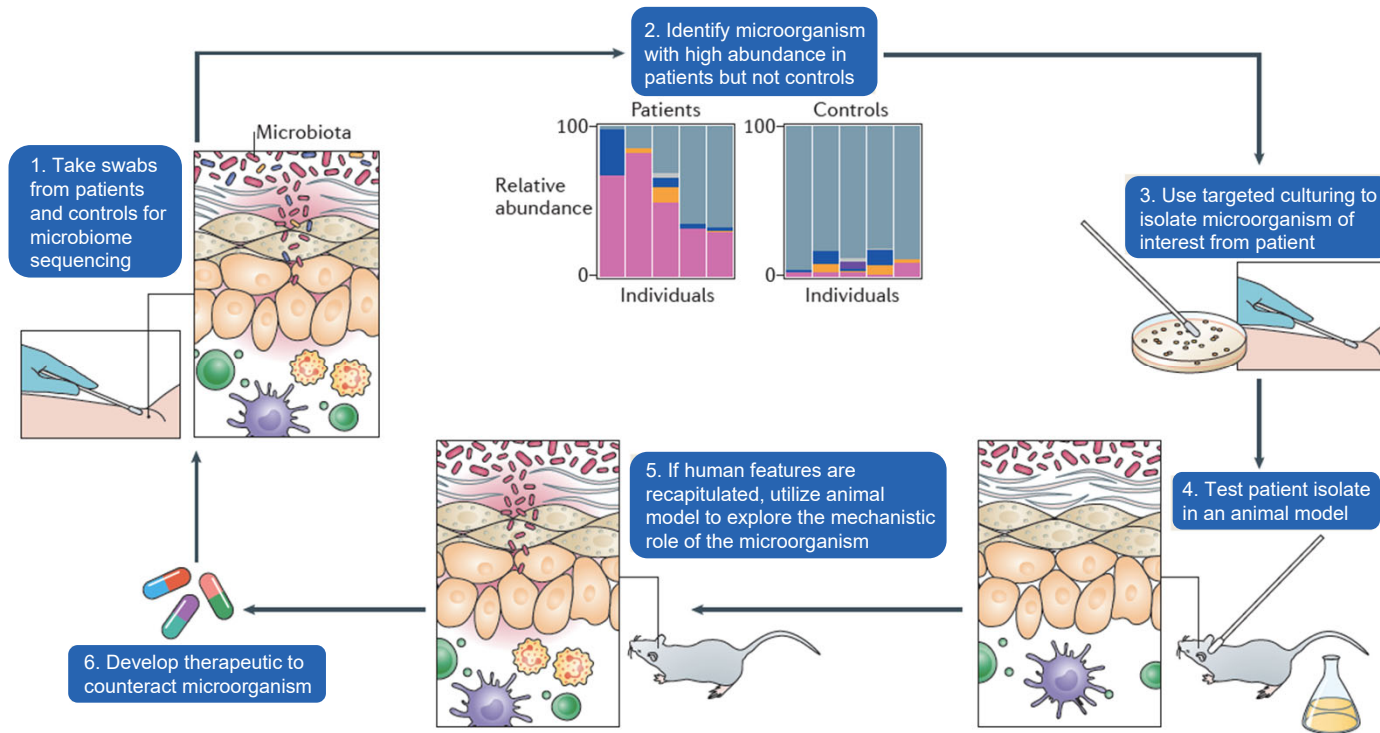
Deciphering multi-kingdom interactions in skin microbiome

- Skin health and disease may be a function of interactions across organisms from different kingdoms, or multi-kingdom interactions
- Bacteria, fungi and viruses interact with the host and immune system to maintain health or promote disease



Lima-Junior, D. S., Krishnamurthy, S. R., Bouladoux, N., Collins, N., Han, S. J., Chen, E. Y., ... & Belkaid, Y. (2021). Endogenous retroviruses promote homeostatic and inflammatory responses to the microbiota. *Cell*, 184(14), 3794-3811.

Developing therapeutic approaches in skin microbiome



Byrd, A. L., Belkaid, Y., & Segre, J. A. (2018). The human skin microbiome. *Nature Reviews Microbiology*, 16(3), 143-155.

Future directions in skin microbiome research



Expand understanding of the skin microbiome in health and disease



Continue improvements related to collection, stabilization, extraction and detection for skin microbiome assays



Layer information onto metagenomic, metatranscriptomic and metabolomic studies to provide new biological and clinical insights



Understand potential effects to skin conditions and the skin microbiome from novel treatments and microbial-based therapeutics



Questions?



Thank you and questions



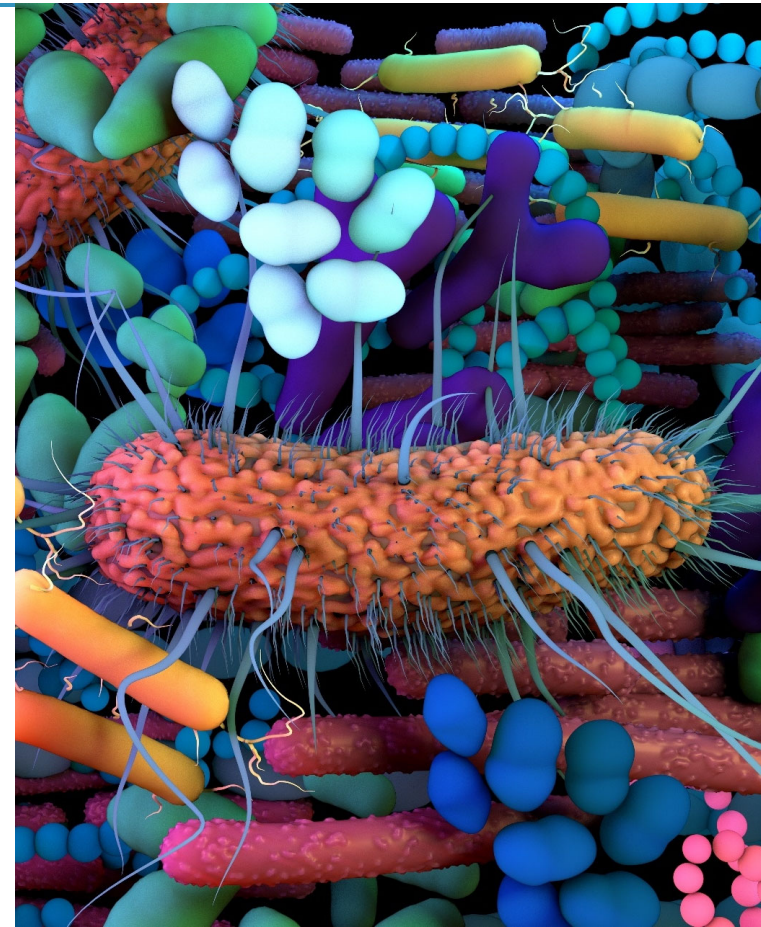
For more information about microbiome resources:

www.atcc.org/microbiome

info@diversigen.com

<https://www.diversigen.com/services/skinmicrobiome/>

Be sure to like and subscribe to ATCC's new podcast, *Behind the Biology*



©2022 American Type Culture Collection. The ATCC trademark and trade name, and any other trademarks listed in this publication are trademarks owned by the American Type Culture Collection unless indicated otherwise.

