



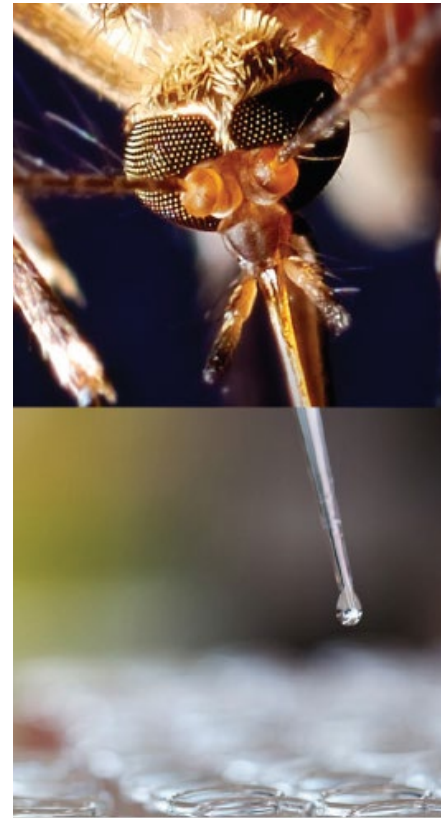
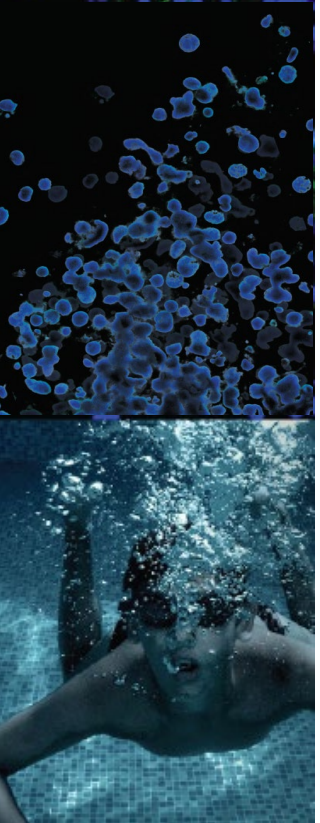
Genomic Data Quality

Connecting the Dots Between Bioinformatics
and Physical Materials

Jonathan Jacobs, PhD

Senior Director, Bioinformatics
Sequencing & Bioinformatics Center
ATCC

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Genomics data quality

Connecting the dots between bioinformatics and physical materials

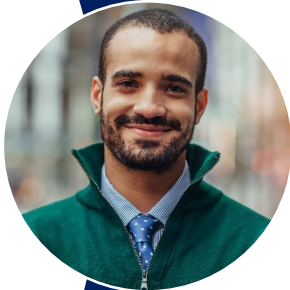
- Review challenges associated with genomics data quality and authenticity
- Discuss **why** ATCC is committed to **providing reference-quality genomes** for our materials
- Discuss our current efforts to produce standardized genomics reference data
- Explore the ATCC Genome Portal
- Explore the ATCC Cell Line Land



Challenges stemming from poor data quality...



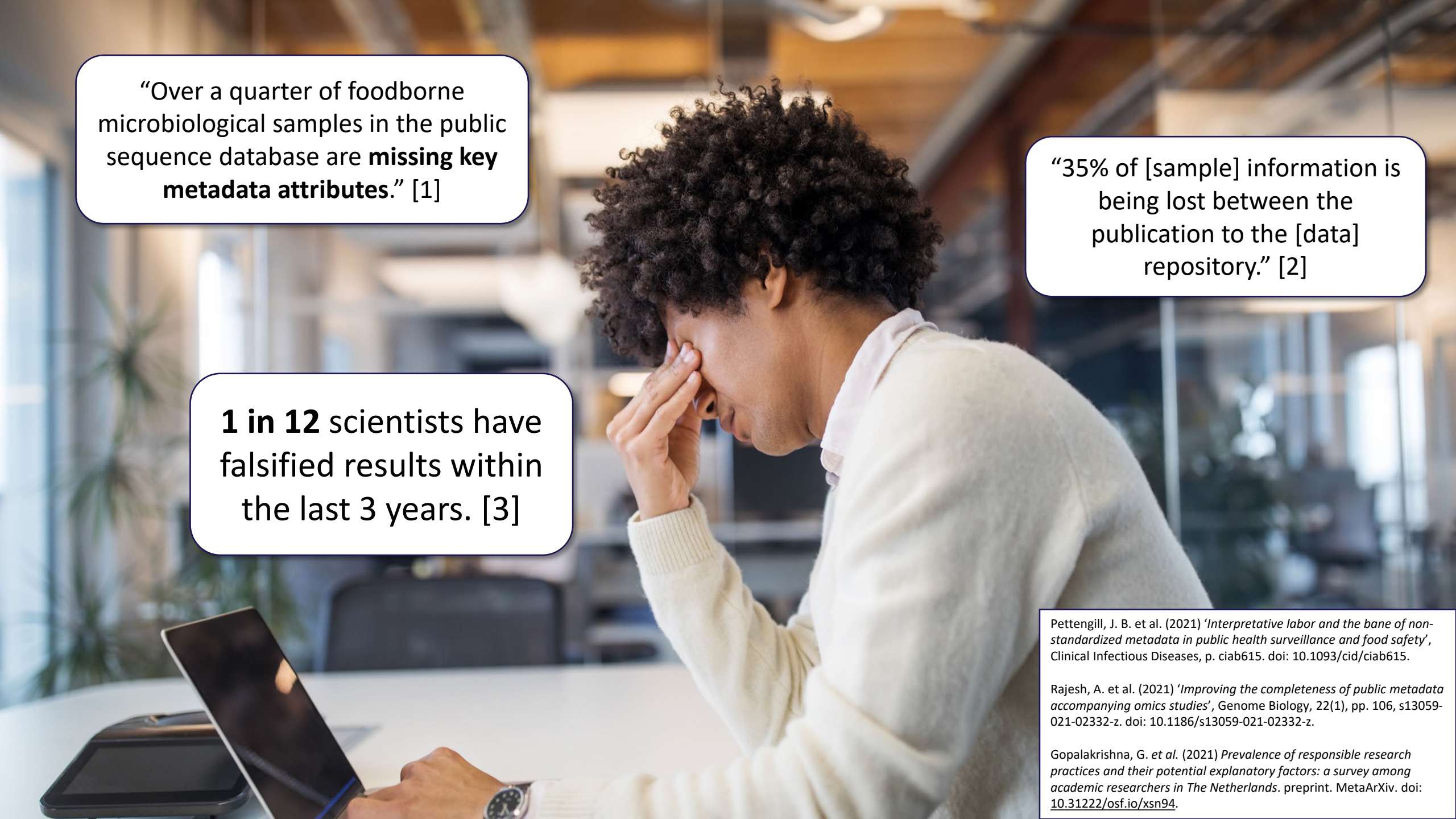
“Finding the right cell lines for my research is a challenge.”



“Many cell types are ***not good models*** for the disease I’m studying.”



“Pre-existing results are difficult to reproduce and often ***not reproducible.***”



“Over a quarter of foodborne microbiological samples in the public sequence database are **missing key metadata attributes.**” [1]

“35% of [sample] information is being lost between the publication to the [data] repository.” [2]

1 in 12 scientists have falsified results within the last 3 years. [3]

Pettengill, J. B. et al. (2021) *'Interpretative labor and the bane of non-standardized metadata in public health surveillance and food safety'*, *Clinical Infectious Diseases*, p. ciab615. doi: 10.1093/cid/ciab615.

Rajesh, A. et al. (2021) *'Improving the completeness of public metadata accompanying omics studies'*, *Genome Biology*, 22(1), pp. 106, s13059-021-02332-z. doi: 10.1186/s13059-021-02332-z.

Gopalakrishna, G. et al. (2021) *Prevalence of responsible research practices and their potential explanatory factors: a survey among academic researchers in The Netherlands*. preprint. MetaArXiv. doi: [10.31222/osf.io/xsn94](https://doi.org/10.31222/osf.io/xsn94).

Fake data was first discovered in GenBank in 1997



“Mr. Hajra, former graduate student, University of Michigan, engaged in scientific misconduct by falsifying and fabricating research data in five published research papers, two published review articles, one submitted but unpublished paper, in his doctoral dissertation, and in a submission to the GenBank computer data base.” – The Federal Register, v62, n135 (1997)

author of the application is identified and that person's role in the project is

years. In the event a consortium of applicants is proposed, the project history of prior joint work should be provided. The previous Federal assistance is identified by project number, Federal agency, and grants or contracting officer. 25 points

Components of a Complete Application

A complete application consists of the following items in this order:

1. Application for Federal Assistance (Standard Form 424, REV 4-88);
2. Budget Information—Non-construction Programs (Standard Form 424A, REV 4-88);
3. Assurances—Non-construction Programs (Standard Form 424B, REV 4-88);
4. Table of Contents;

Dated: July 9, 1997.

David F. Garrison,

Principal Deputy Assistant Secretary for Planning and Evaluation.

[FR Doc. 97-18528 Filed 7-14-97; 8:45 am]

BILLING CODE 4151-04-M

DEPARTMENT OF HEALTH AND HUMAN SERVICES

Office of the Secretary

Findings of Scientific Misconduct

AGENCY: Office of the Secretary, HHS.
ACTION: Notice.

SUMMARY: Notice is hereby given that the Office of Research Integrity (ORI) has made a final finding of scientific misconduct in the following case:

Amitav Hajra, University of Michigan: Based upon a report from the University of Michigan, information obtained by the Office of Research Integrity (ORI) during its oversight review, and Mr. Hajra's own admission, ORI found that Mr. Hajra, former graduate student, University of Michigan, engaged in scientific misconduct by falsifying and fabricating research data in five published research papers, two published review articles, one submitted but unpublished paper, in his doctoral dissertation, and in a submission to the GenBank computer data base. Mr. Hajra's doctoral training and research was supported by two Public Health Service (PHS) grants, and his experiments were conducted at and submitted for publication from the

• Wijmenga, C., Gregory, P.E., Hajra, A., Schröck, E., Ried, T., Eils, R., Liu, P.P., and Collins, F.S. "Core binding factor β -smooth muscle myosin heavy chain chimeric protein involved in acute myeloid leukemia forms unusual nuclear rod-like structures in transformed NIH 3T3 cells." *Proc. Natl. Acad. Sci. USA* 93(4):1630-1635, 1996; and

• Liu, P.P., Wijmenga, C., Hajra, A., Blake, T.B., Kelley, C.A., Adelstein, R.S., Bagg, A., Rector, J., Cotelingham, J., Willman, C.L., and Collins, F.S. "Identification of the chimeric protein product of the CBF β -MYH11 fusion gene in inv(16) leukemia cells." *Genes, Chromosomes, and Cancer* 16:77-87, 1996 (Erratum in *Genes, Chromosomes, and Cancer* 18(1):71, 1997).

Mr. Hajra included fabricated and falsified data in the following review articles:

• Hajra, A., Liu, P.P., and Collins, F.S. "Transforming properties of the leukemic Inv(16) fusion gene CBF β -MYH11." In *Molecular Aspects of Myeloid Stem Cell Development in Current Topics in Microbiology and Immunology* (L. Wolff and A.S. Perkins, Eds.) 211:289-298, 1996 (Review). Berlin and New York: Springer-Verlag; and

• Liu, P.P., Hajra, A., Wijmenga, C., and Collins, F.S. "Molecular pathogenesis of the chromosome 16 inversion in the M4Eo subtype of acute myeloid leukemia." *Blood* 85:2289-2302, 1995 (Review).

Mr. Hajra submitted a fabricated nucleotide sequence in computer data

24 years later, this falsified data still being cited...


Received: 25 March 2021 | Revised: 16 June 2021 | Accepted: 13 July 2021
DOI: 10.1002/humu.24261

REVIEW

Human Mutation

Pathogenic noncoding variants in the neurofibromatosis schwannomatosis predisposition genes

PEREZ-BECERRIL ET AL.

Cristina Perez-Becerril 

Division of Evolution and Genomic Science, Manchester Centre for Genomic Medicine, St Mary's Hospital, Manchester Academic Health Science Centre, School of Biological Sciences, University of Manchester, Manchester, UK

Correspondence
Miriam J. Smith, Division of Evolution and Genomic Science, Manchester Centre for Genomic Medicine, St Mary's Hospital, Manchester Academic Health Science Centre, School of Biological Sciences, University of Manchester, Manchester M13 9WL, UK. Email: miriam.smith@manchester.ac.uk

comparison of the full human and murine neurofibromin sequences revealed a high degree of similarity (>98%) and high conservation levels across 5'- and 3'-UTRs (Bernards et al., 1993; Hajra et al., 1994). A subsequent *in silico* study compared the 5' upstream region and intron 1 of *NF1* and homologous genes in human, mouse, rat, and puffer fish (*Fugu rubripes*). The authors found high homology segments throughout the region across all species, including two exact

and *NF2* loci, respectively. To date, most variants associated with have been identified in the *SMARCB1* and *LZTR1* genes, and at the *DGCR8* gene was recently reported to predispose to schwannomatosis. The high detection rate for PVs in *NF1* and *NF2* (over 90% of variants can be identified by routine genetic screening) under a portion of clinical cases remain undetected. A higher proportion



author of the application is identified and that person's role in the project is identified. 20 points

4. *Organizational Experience.* The application identifies the qualifying experience of the organization to demonstrate the applicant's ability to effectively and efficiently administer this project. The application specifically identifies the applicant as a nationally-recognized organization, institution, or company with a record of study and analysis of rural and special transportation needs. Previous specific experience with work similar to the Tasks proposed is clearly and specifically described. The relationship between this project and other work planned, anticipated, or underway by the applicant is described, including a chart which lists all related Federal assistance received within the last five years. In the event a consortium of applicants is proposed, the project history of prior joint work should be provided. The previous Federal assistance is identified by project number, Federal agency, and grants or contracting officer. 25 points

- Components of a Complete Application*
- A complete application consists of the following items in this order:
1. Application for Federal Assistance (Standard Form 424, REV 4-88);
 2. Budget Information—Non-construction Programs (Standard Form 424A, REV 4-88);
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Dated: July 9, 1997.
David F. Garrison,
Principal Deputy Assistant Secretary for Planning and Evaluation.
[FR Doc. 97-18528 Filed 7-14-97; 8:45 am]
BILLING CODE 4151-04-M

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Office of the Secretary

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• Wijmenga, C., Gregory, P.E., Hajra, A., Schröck, E., Ried, T., Eils, R., Liu, P.P., and Collins, F.S. "Core binding factor β -smooth muscle myosin heavy chain chimeric protein involved in acute myeloid leukemia forms unusual nuclear rod-like structures in transformed NIH 3T3 cells." *Proc. Natl. Acad. Sci. USA* 93(4):1630-1635, 1996; and

• Liu, P.P., Wijmenga, C., Hajra, A., Blake, T.B., Kelley, C.A., Adelstein, R.S., Bagg, A., Rector, J., Cotelingham, J., Willman, C.L., and Collins, F.S. "Identification of the chimeric protein product of the CBF β -MYH11 fusion gene in *inv(16)* leukemia cells." *Genes, Chromosomes, and Cancer* 16:77-87, 1996 (Erratum in *Genes, Chromosomes, and Cancer* 18(1):71, 1997).

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• Liu, P.P., Hajra, A., Wijmenga, C., and Collins, F.S. "Molecular pathogenesis of the chromosome 16 inversion in the M4Eo subtype of acute myeloid leukemia." *Blood* 85:2289-2302, 1995 (Review).

Mr. Hajra submitted a fabricated nucleotide sequence in computer data



After 42 citations... the data is still in GenBank...

Article preview

DNA Sequences in the Promoter Region of the NF1 Gene Are Highly Conserved between Human and Mouse

Amitav Hajra, Antonia Martin-Gallardo, Susan A. Tarlé, Matthew Freedman, Susan Wilson-Gunn, Andre Bernards, Francis S. Collins

Abstract

The gene for type 1 neurofibromatosis (NF1) is most highly expressed in brain and spinal cord, although low levels of mRNA can be found in nearly all tissues. As a first step in investigating the regulation of NF1 gene expression, we have cloned and sequenced the promoter regions of the human and mouse NF1 genes and mapped the transcriptional start sites in both species. We report here that the 5' ends of the human and murine NF1 genes are highly conserved. While no discernable TATA or CCAAT box sequences are seen, transcription initiates at identical sites in both species, 484 nucleotides upstream of the ATG initiation codon in the human gene. The human and mouse NF1 genes share particularly high sequence homology (95%) between nucleotides -33 and +261 and contain several perfectly conserved transcription factor binding site motifs, including a cAMP response element, several AP2 consensus binding sites, and a serum response element. The high conservation of these sequences indicates that they are likely to be significant in the regulation of NF1 gene expression.

References (0)

Cited by (42)

Synthetic promoter for efficient and muscle-specific expression of exogenous genes 2019, Plasmid

Human neurofibromin (NF1) gene, promoter region and partial cds

GenBank: U17084.1

FASTA Graphics

LOCUS HSU17084 3953 bp DNA linear PRI 07-DEC-1994

DEFINITION Human neurofibromin (NF1) gene, promoter region and partial cds.

ACCESSION U17084 U09106

VERSION U17084.1

KEYWORDS .

SOURCE Homo sapiens (human)

ORGANISM *Homo sapiens*

Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Mammalia; Eutheria; Euarchontoglires; Primates; Haplorrhini; Catarrhini; Hominidae; Homo.

REFERENCE 1 (bases 2943 to 3953)

AUTHORS Hajra,A., Martin-Gallardo,A., Tarle,S.A., Freedman,M., Wilson-Gunn,S., Bernards,A. and Collins,F.S.

TITLE DNA sequences in the promoter region of the NF1 gene are highly conserved between human and mouse

JOURNAL Genomics 21 (3), 649-652 (1994)

PUBMED 7959746

REFERENCE 2 (bases 1 to 3953)

AUTHORS Marchuk,D.A., Saulino,A.M., Tavakkol,R., Swaroop,M., Wallace,M.R., Andersen,L.B., Mitchell,A.L., Gutmann,D.H., Boguski,M. and Collins,F.S.

TITLE cDNA cloning of the type 1 neurofibromatosis gene: complete sequence of the NF1 gene product

JOURNAL Genomics 11 (4), 931-940 (1991)

PUBMED 1783401

REFERENCE 3 (bases 1 to 3953)

AUTHORS Hajra,A.

TITLE Direct Submission

JOURNAL Submitted (10-NOV-1994) Amitav Hajra, Laboratory of Gene Transfer, National Center for Human Genome Research, NIH, Building 49, Room 3A23, 49 Convent Drive, MSC 4470, Building 49, Room 3A23, 9000

Falsified sequencing data to support a false phylogeny



Biochemical Systematics and Ecology

Volume 96, June 2021, 104263



Scientific data laundering: Chimeric mitogenomes of a sparrowhawk and a nightjar covered-up by forged phylogenies

George Sangster^a, Jolanda A. Luksenburg^{b, c}

Show more

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*“The evidence indicates that Liu et al. (2017) published phylogenies that were not based on existing data **but were fabricated to reflect preconceived ideas about phylogenetic relationships.**” – Sangster & Luksenburg (2021)*

Liu and colleagues in a paper in *Biochemical Systematics and Ecology* in 2017 is not an authentic sequence of this species but represents a chimera of three different species (a

Sangster, G. and Luksenburg, J.A. (2021) ‘Scientific data laundering: Chimeric mitogenomes of a sparrowhawk and a nightjar covered-up by forged phylogenies’, *Biochemical Systematics and Ecology*, 96, p. 104263. doi:[10.1016/j.bse.2021.104263](https://doi.org/10.1016/j.bse.2021.104263).

Unfortunately, the falsified mitogenome is still in GenBank...

UNVERIFIED: *Accipiter gularis* mitochondrion sequence

GenBank: KX585864.1

[FASTA](#) [Graphics](#)

Go to:

LOCUS KX585864 17918 bp DNA linear VRT 31-AUG-2021
DEFINITION UNVERIFIED: *Accipiter gularis* mitochondrion sequence.
ACCESSION KX585864
VERSION KX585864.1
KEYWORDS UNVERIFIED; UNVERIFIED_ORGANISM.
SOURCE mitochondrion *Accipiter gularis* (Japanese sparrowhawk)
ORGANISM [Accipiter gularis](#)
Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi;
Archelosauria; Archosauria; Dinosauria; Saurischia; Theropoda;
Coelurosauria; Aves; Neognathae; Accipitriformes; Accipitridae;
Accipitrinae; *Accipiter*.
REFERENCE 1 (bases 1 to 17918)
AUTHORS Liu,G.
TITLE The complete mtDNA of *Accipiter gularis*
JOURNAL Unpublished
REFERENCE 2 (bases 1 to 17918)
AUTHORS Liu,G.
TITLE Direct Submission
JOURNAL Submitted (21-JUL-2016) School of life science, Anhui Medical
University, 81 Meishan Rd, Hefei, Anhui 230032, China
COMMENT GenBank staff is unable to verify source organism and sequence
and/or annotation provided by the submitter.
FEATURES Location/Qualifiers
source 1..17918

Labeled as “Unverified”, but the sequence still remains in GenBank and, for example, will come up in a BLAST search...

Intentional falsification is rare... but accidents happen right?

(>2 million times) ...

Mukherjee et al. *Standards in Genomic Sciences* 2015, **10**:18
<http://www.standardsingenomics.com/content/10/1/18>



COMMENTARY

Open Access

Large-scale contamination of microbial isolate genomes by Illumina PhiX control

Supratim Mukherjee^{1*}, Marcel Huntemann¹, Natalia Ivanova¹, Nikos C Kyrpides^{1,2} and Amrita Pati¹

Abstract

With the rapid growth and development of sequencing technologies, we are exploring solutions to some of the world's biggest challenges such as sequencing and exploring genomic dark matter. However, progress in sequencing has been hampered by that can occur during template or library preparation, sequencing, and imaging. We screened over 18,000 publicly available microbial isolate genome sequences in a database and identified more than 1000 genomes that are contaminated during Illumina sequencing runs. Approximately 10% of these genomes are contaminated genomes were sequenced under the Human Microbiome Project. Contamination from various sources and are usually eliminated during decontamination of PhiX contaminated genomes indicates a lapse in either the application of measures. The presence of PhiX contamination in several publicly available genomes when such data are used in comparative genomics analyses. Such contamination has far-reaching consequences in the form of erroneous data interpretation and measures to proofread raw sequences before releasing them to the broad community.

Keywords: Next-generation sequencing, PhiX, Contamination, Comparative genomics

Background

The ability to produce large numbers of high-quality, low-cost reads has revolutionized the field of microbiology [1-3]. Starting from a meager 1575 registered projects in September 2005, there has been a steady increase in the number of sequencing projects according to the Genomes OnLine Database [4]. As of November 17th 2014, there were 41,553 bacterial and archaeal isolate genome sequencing projects reported in GOLD [4,5]. This explosion of genome sequencing projects especially during the last 5 years has been largely cata-

Despite its need to be One such ch used as a qu runs. PhiX i with a single 5386 nucleot sequenced b defined gen used as a co

Steinegger and Salzberg *Genome Biology* (2020) 21:115
<https://doi.org/10.1186/s13059-020-02023-1>

METHOD

Open Access

Terminating contamination: large-scale search identifies more than 2,000,000 contaminated entries in GenBank

Martin Steinegger^{1,2,3*} and Steven L. Salzberg^{2,4,5}

*Correspondence: martin.steinegger@snu.ac.kr
¹School of Biological Sciences, Seoul National University, Seoul, 08826, South Korea
²Center for Computational Biology, Whiting School of Engineering, Johns Hopkins University, 21218 Baltimore, Maryland, USA
Full list of author information is available at the end of the article

Abstract

Genomic analyses are sensitive to contamination in public databases caused by incorrectly labeled reference sequences. Here, we describe Conterminator, an efficient method to detect and remove incorrectly labeled sequences by an exhaustive all-against-all sequence comparison. Our analysis reports contamination of 2,161,746, 114,035, and 14,148 sequences in the RefSeq, GenBank, and NR databases, respectively, spanning the whole range from draft to "complete" model organism genomes. Our method scales linearly with input size and can process 3.3 TB in 12 days on a 32-core computer. Conterminator can help ensure the quality of reference databases. Source code (GPLv3): <https://github.com/martin-steinegger/conterminator>

Downloaded from genome.cshlp.org on October 20, 2021 - Published by Cold Spring Harbor Laboratory Press

Research

Human contamination in bacterial genomes has created thousands of spurious proteins

Florian P. Breitwieser,¹ Mihaela Pertea,^{1,2} Aleksey V. Zimin,^{1,3} and Steven L. Salzberg^{1,2,3,4}

¹Center for Computational Biology, McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins School of Medicine, Baltimore, Maryland 21205, USA; ²Department of Computer Science, Whiting School of Engineering, Johns Hopkins University, Baltimore, Maryland 21218, USA; ³Department of Biomedical Engineering, Johns Hopkins University, Baltimore, Maryland 21218, USA; ⁴Department of Biostatistics, Bloomberg School of Public Health, Johns Hopkins University, Baltimore, Maryland 21205, USA

Genome Biology



shed genomes can cause numerous problems for downstream analyses, particularly in metagenomics projects. Our large-scale scan of complete and draft bacterial and archaeal genomes that 2250 genomes are contaminated by human sequence. The contaminant human repeat regions, which themselves are not adequately represented in the reference genome. The absence of the sequences from the human assembly offers a likely explanation. In some cases, the contaminating contigs have been erroneously annotated over time have propagated to create spurious protein "families" across multiple databases. As a result, 3437 spurious protein entries are currently present in the widely used protein databases. We report here an extensive list of contaminant sequences in bacterial genome assemblies. We found that nearly all contaminants occurred in small contigs in draft genome assemblies may mitigate the issue of contaminant genomic sequences.

[article]

available ge- to well over il resources for g microbiome plex samples ence databases but for practi- today are still gs or scaffolds chromosomes eor "finished" y chromosome he human ger animal gean assembly, folds that con- sequence has ive regions are g to problems es vary widely

assemblies in the NCBI and UCSC Genome Browser databases were contaminated with the primate-specific *AhrY* repeats (Longo et al. 2011). Although validation pipelines have improved substantially since then (Tatusova et al. 2016; Haft et al. 2018), some contaminants still remain, as we describe below. Furthermore, when open reading frames (ORFs) in the contaminated contigs get annotated as protein-coding genes, their protein sequence may be added to other databases. Once in those databases, these spurious proteins may in turn be used in future annotation, leading to the so-called "transitive catastrophe" problem where errors are propagated widely (Karp 1998; Salzberg 2007; Danchin et al. 2018). Indeed, one study found that the percentage of misannotated entries in the NCBI nonredundant (nr) protein collection, which is used for thousands of BLAST searches every day, has been increasing over time (Schnoes et al. 2009).

Contamination of genomic sequences can be particularly problematic for metagenomic studies. For example, if a genome is labeled as species X contains fragments of the human genome, then any sample containing human DNA might erroneously be identified as also containing species X. Since human DNA is virtually al-

Poor quality genomes result in taxonomic misclassification

Multiple papers (more than the two listed here) have found widespread misclassification in GenBank

Bioinformatics, 36(18), 2020, 4699–4705
doi: 10.1093/bioinformatics/btaa586
Advance Access Publication Date: 24 June 2020
Original Paper



Sequence analysis

Detecting and correcting misclassified sequences in the large-scale public databases

Hamid Bagheri^{1,*}, Andrew J. Severin² and Hridesh Rajan¹

¹Department of Computer Science and ²Genome Informatics Facility, Iowa State University, Ames, IA 50011, USA

*To whom correspondence should be addressed.
Associate Editor: Arne Elofsson

Received on April 2, 2020; revised on June 10, 2020; editorial decision on June 16, 2020

Abstract

Motivation: As the cost of sequencing has decreased, the amount of DNA sequences deposited into public repositories is increasing rapidly. Public databases rely on user input and do not have methods for identifying errors in the provided metadata. This leads to the propagation of errors. Previous research on a small subset of the NR database analyzed the amount of misclassification in the database. To the best of our knowledge, the amount of misclassification in the database is increasing. We propose a heuristic method to detect potentially misclassified taxonomic assignments. The method uses a heuristic and quality control to find the most probable taxonomic assignment for each annotation from manually and computationally derived taxonomic assignments.

Results: We found more than two million misclassified sequences in the NR database. Using simulated data, we show a high precision of 95% for the proposed method. The misclassified proteins. The proposed approach and findings could also be applied to other public databases.

Availability and implementation: Source code, dataset, documentation and Docker container are available at <https://github.com/boalang/nr>.

Contact: hbagheri@iastate.edu

Supplementary information: Supplementary data are available at <https://doi.org/10.1093/bioinformatics/btaa586>.

1 Introduction

Researchers use BLAST on the non-redundant (NR) database on a daily basis to identify the source and function of a protein/DNA sequence. The NR database encompasses protein sequences from

are deposited into public repositories. Sequences were deposited by a researcher who obtained them from a soybean roots, but the taxonomic metadata will be labeled with the organism name *Glycine max*. If the researcher had in fact been working on *Glycine soja* then this would result in a misclassification.

~7.8% of genomes misclassified at the species level

~4% at the genus level

PLOS ONE

RESEARCH ARTICLE

Large-scale k -mer-based analysis of the informational properties of genomes, comparative genomics and taxonomy

Yuval Bussi^{1,2,3}, Ruti Kapon¹, Ziv Reich^{1*}

¹ Department of Biomolecular Sciences, Weizmann Institute of Science, Rehovot, Israel, ² Department of Computer Science and Applied Mathematics, Weizmann Institute of Science, Rehovot, Israel, ³ Department of Molecular Cell Biology, Weizmann Institute of Science, Rehovot, Israel

* ziv.reich@weizmann.ac.il



Abstract

Information theoretic approaches are widely used in a variety of bioinformatics applications. In comparative genomics, the analysis of short DNA words, or k -mers, are particularly useful. Analyzing k -mer lengths for genome comparison is a common task. In 2005, 1605 genomes in the KEGG GENES database were analyzed using k -mers spanning the relevant range of k values. In this study, we analyze representative genomes using k -mers to study the relationship between a phylogenetic/taxonomic distance and k -mer similarity. Analyzing high subtree similarity for k -mers in a large dataset of 14.2M prokaryotic genomes, we detected many potential misclassifications. At the genus level, we detected many potential misclassifications, demonstrating the need for whole-genome similarity-based taxonomic assignments.

~7% of genomes misclassified at genus or higher

OPEN ACCESS

Citation: Bussi Y, Kapon R, Reich Z (2021) Large-scale k -mer-based analysis of the informational properties of genomes, comparative genomics and taxonomy. PLoS ONE 16(10): e0258693. <https://doi.org/10.1371/journal.pone.0258693>

Editor: Orni Finkel, University of North Carolina at Chapel Hill, UNITED STATES

Received: April 30, 2021

Accepted: October 2, 2021

Published: October 14, 2021

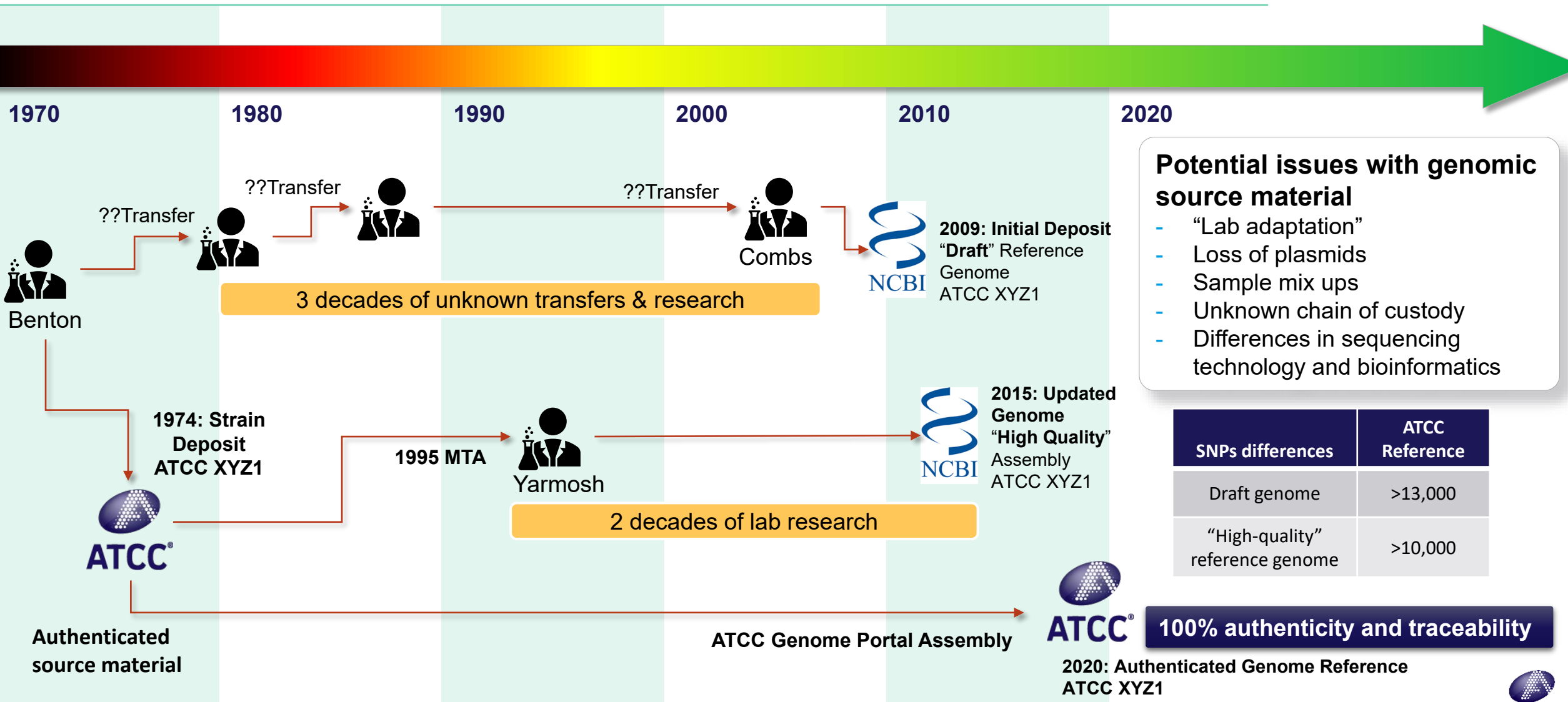
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Data Availability Statement: Data and code for

Introduction

Information theory, initially developed for the mathematical analysis of communication systems by Shannon [1], has been applied to molecular biology for decades. Gattin's pioneering works in the late 1960s were the first to define life as an information processing system [2, 3].

Challenging traceability of most public genomics data



Potential issues with genomic source material

- "Lab adaptation"
- Loss of plasmids
- Sample mix ups
- Unknown chain of custody
- Differences in sequencing technology and bioinformatics

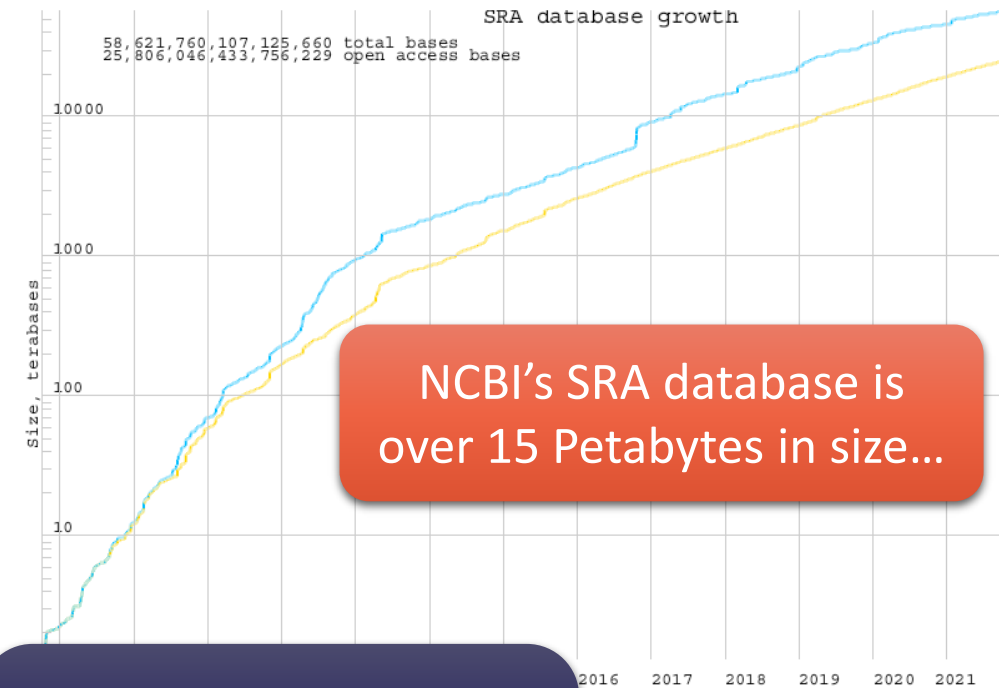
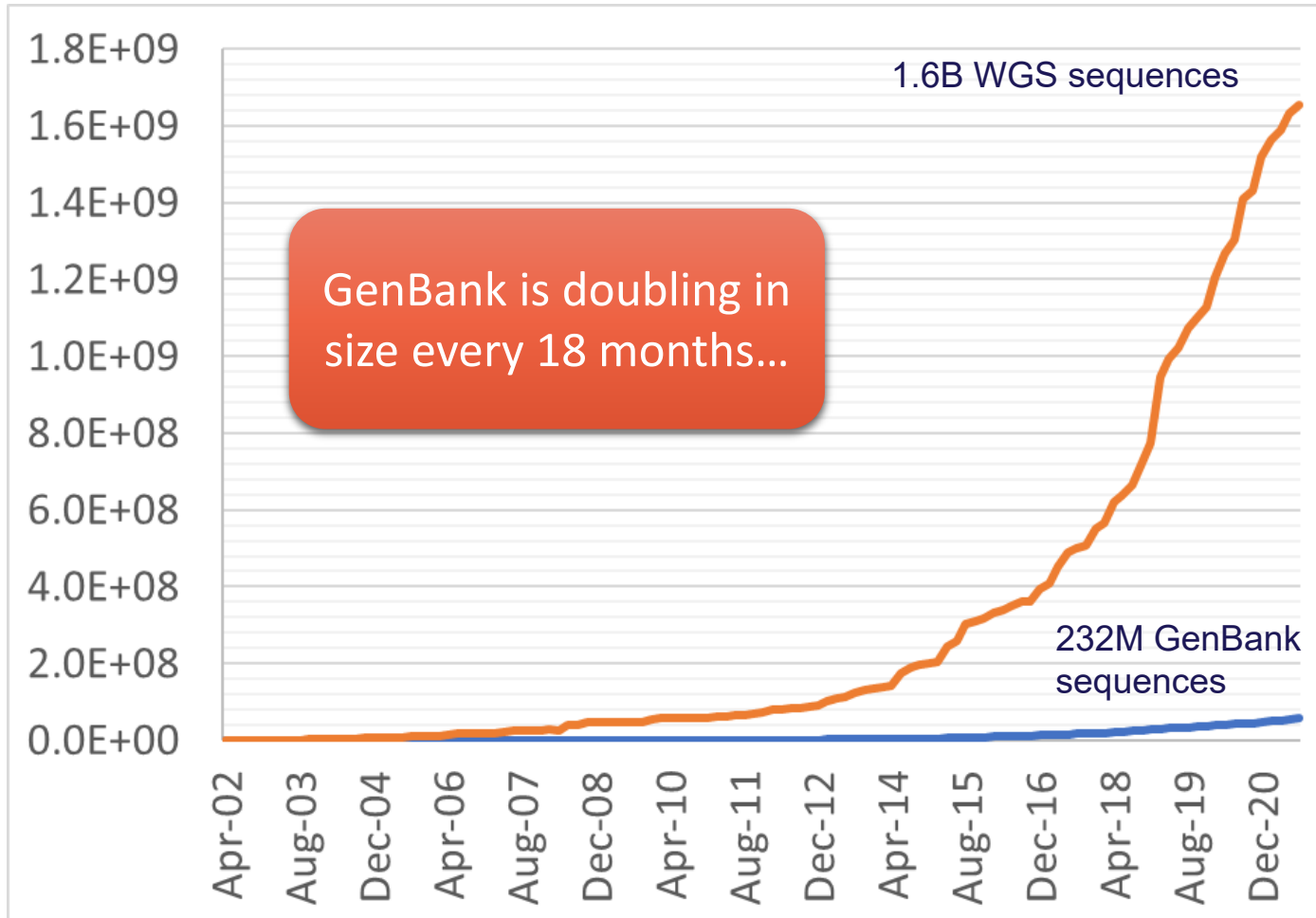
SNPs differences	ATCC Reference
Draft genome	>13,000
"High-quality" reference genome	>10,000



A reminder on the growth of public genomics data

1.6B sequences in WGS

232M sequences in GenBank



Data curation is a huge challenge

Genomics data quality issues impact many disciplines

FACTORS

- Misclassification of sequences
- Chimeric genome assemblies
- Sample contamination
- Sequencing errors
- Mislabeling or data errors
- Data omission
- Data obfuscation
- Intentional misconduct



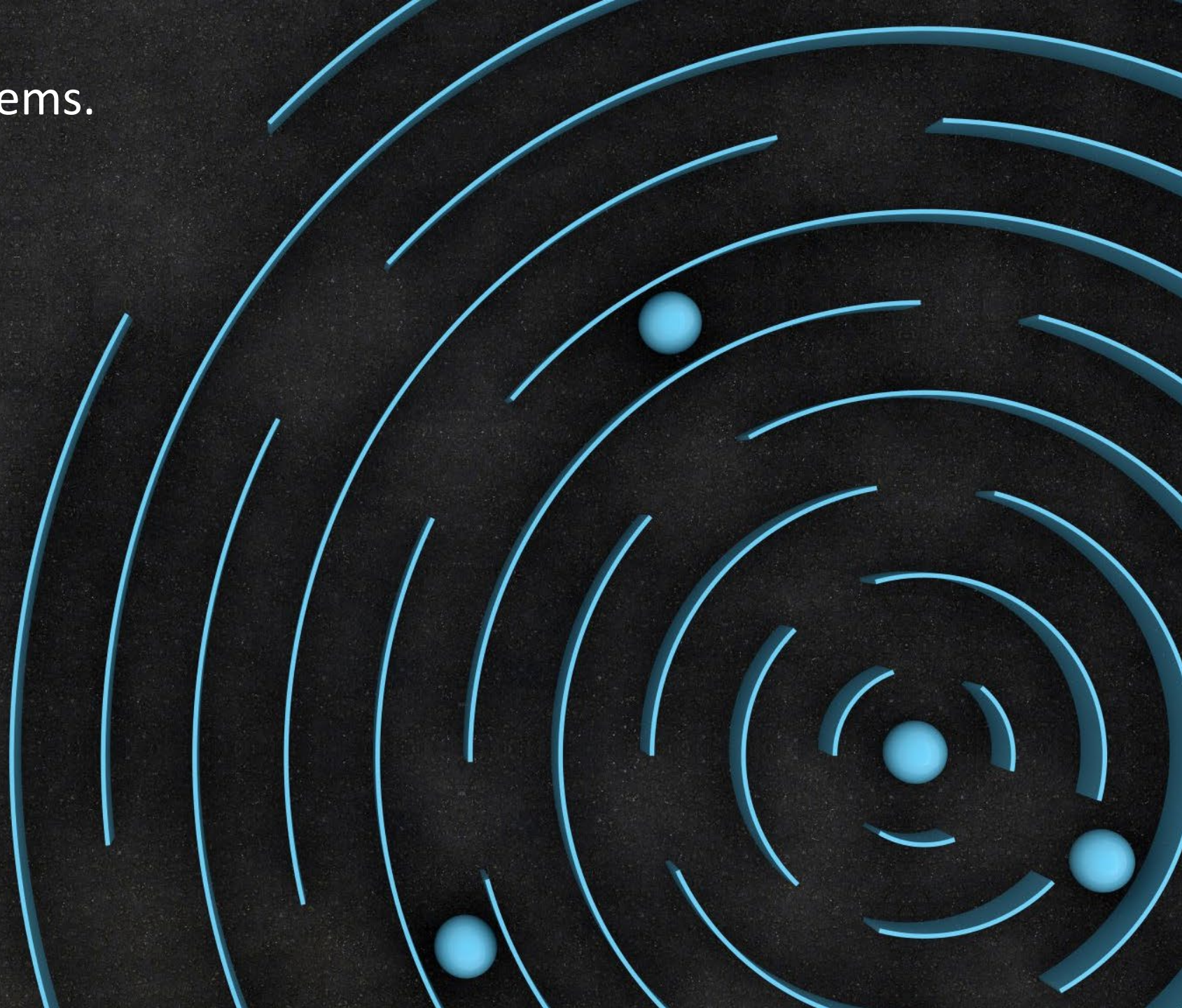
Critically Impacted Areas

- Basic research (hypothesis generation)
- Biodiversity and environmental sciences
- Diagnostics & epidemiology
- Forensics
- Food safety
- Biodefense
- Many other areas...

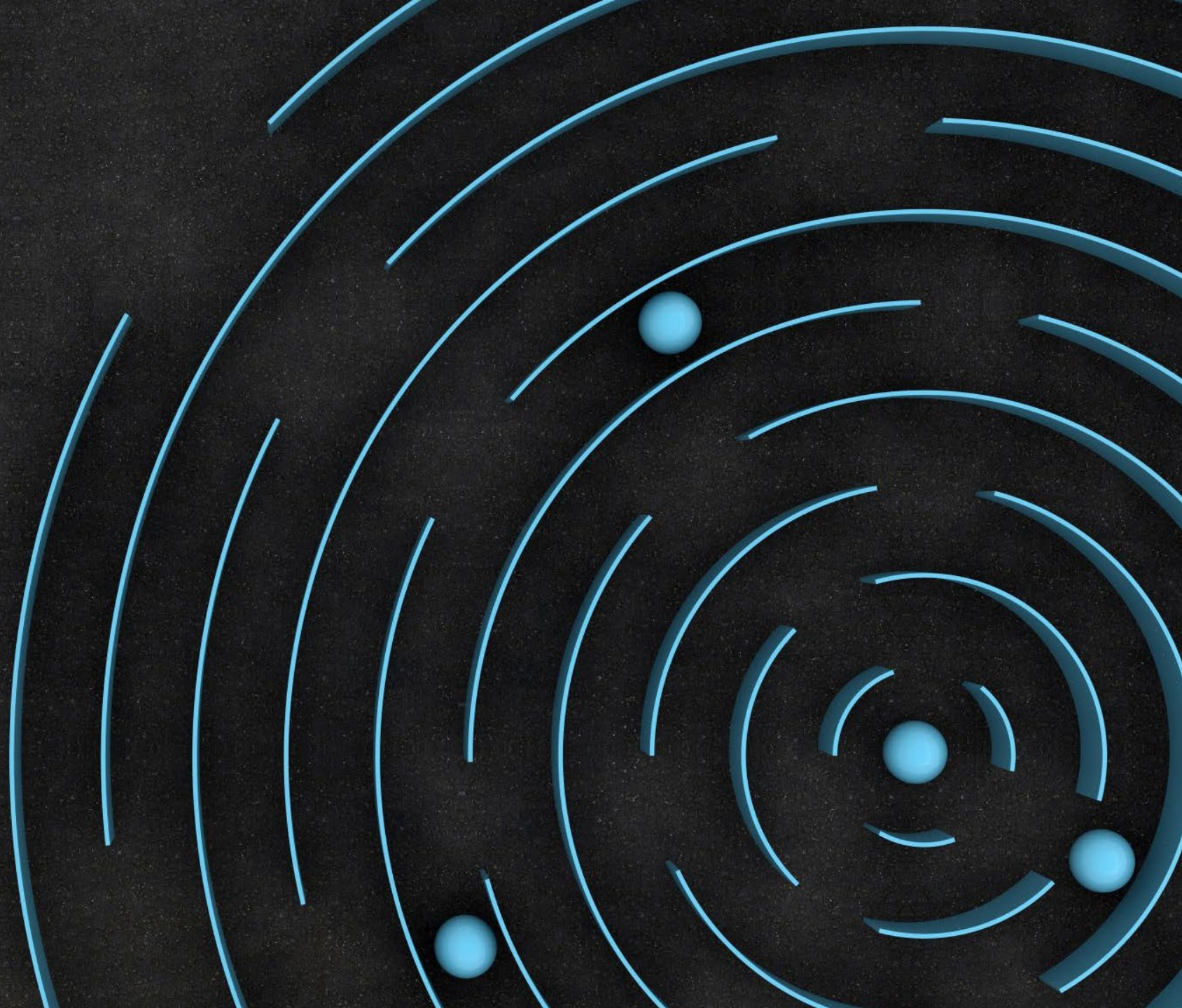
These are not “new” problems.

Many groups have sought solutions.

None, however, have sought to create
Authenticated Genomics Data



What is
“Authenticated Genomics
Data”?



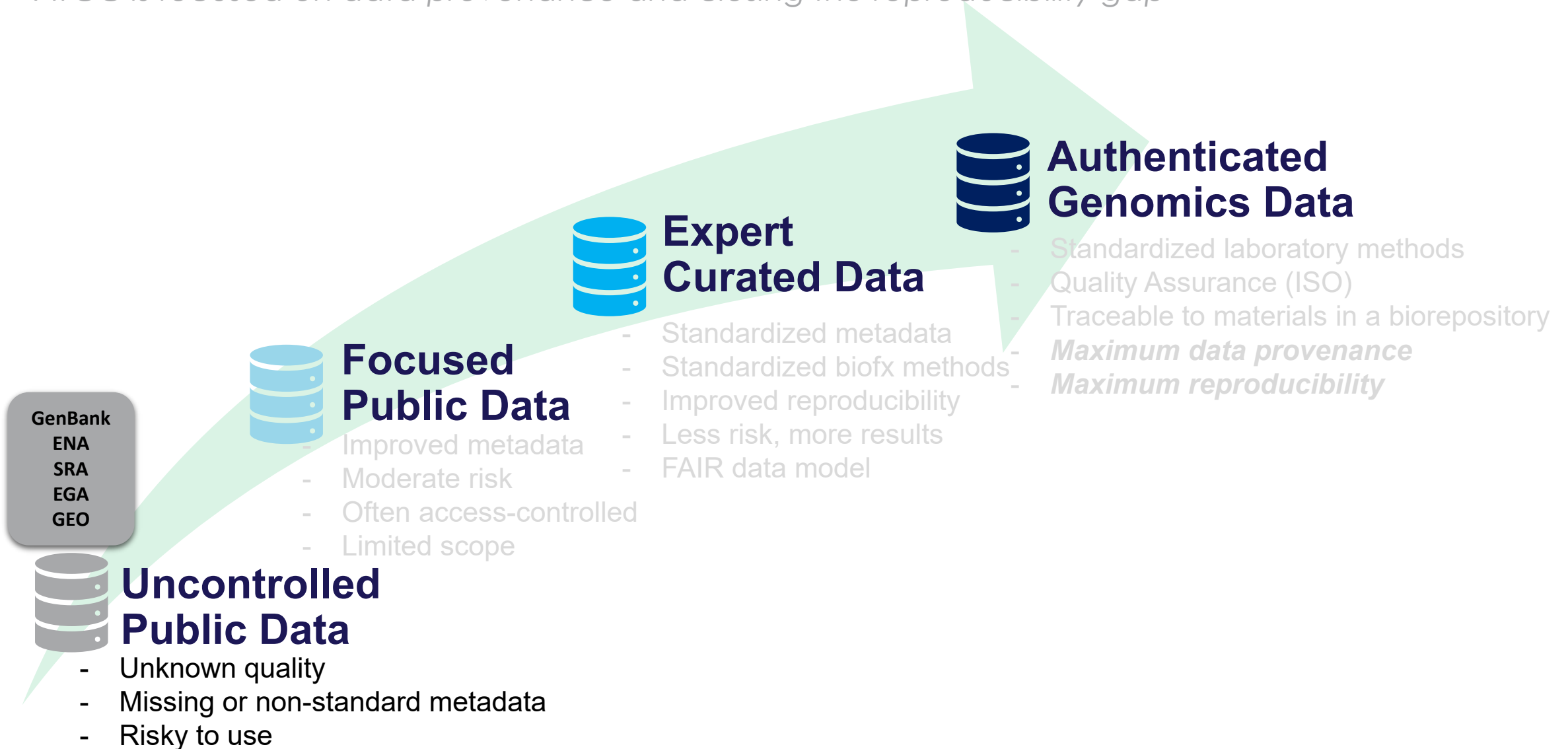


Authenticated Genomics Data:

- 1. Traceable to physical materials*
- 2. Produced with defined quality assurance metrics*
- 3. Reproducible across multiple tests*

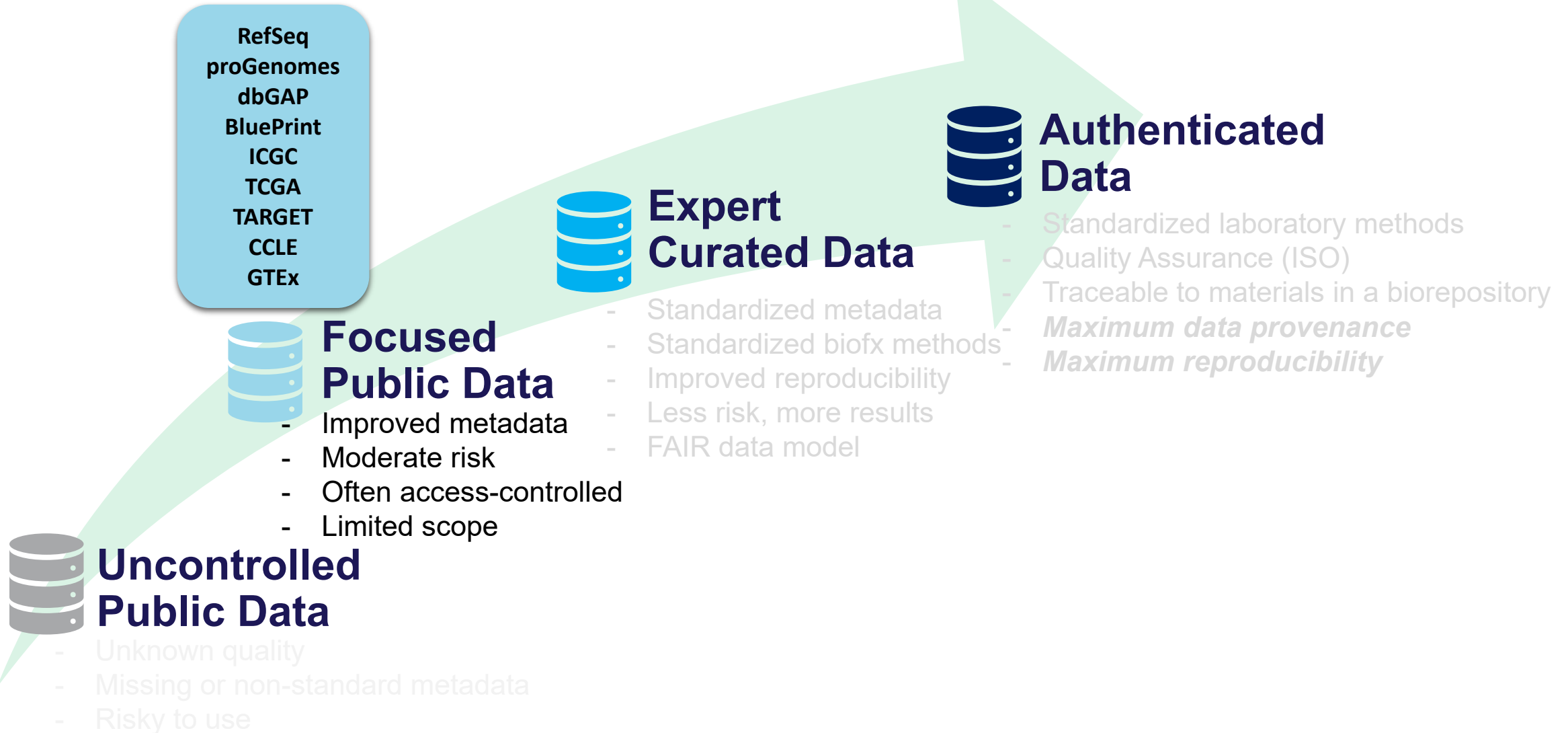
Authenticated genomics data at ATCC

ATCC is focused on data provenance and closing the reproducibility gap



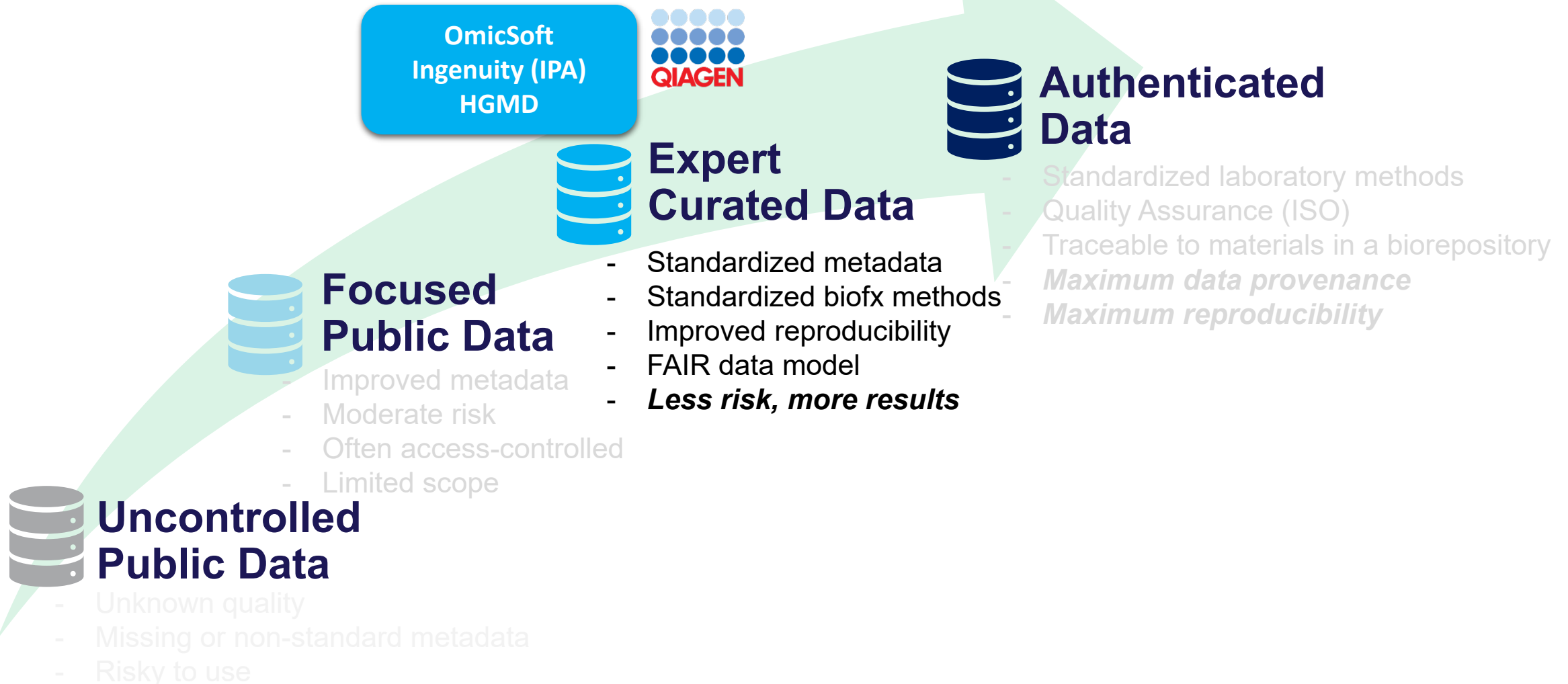
Authenticated genomics data at ATCC

ATCC is focused on data provenance and closing the reproducibility gap



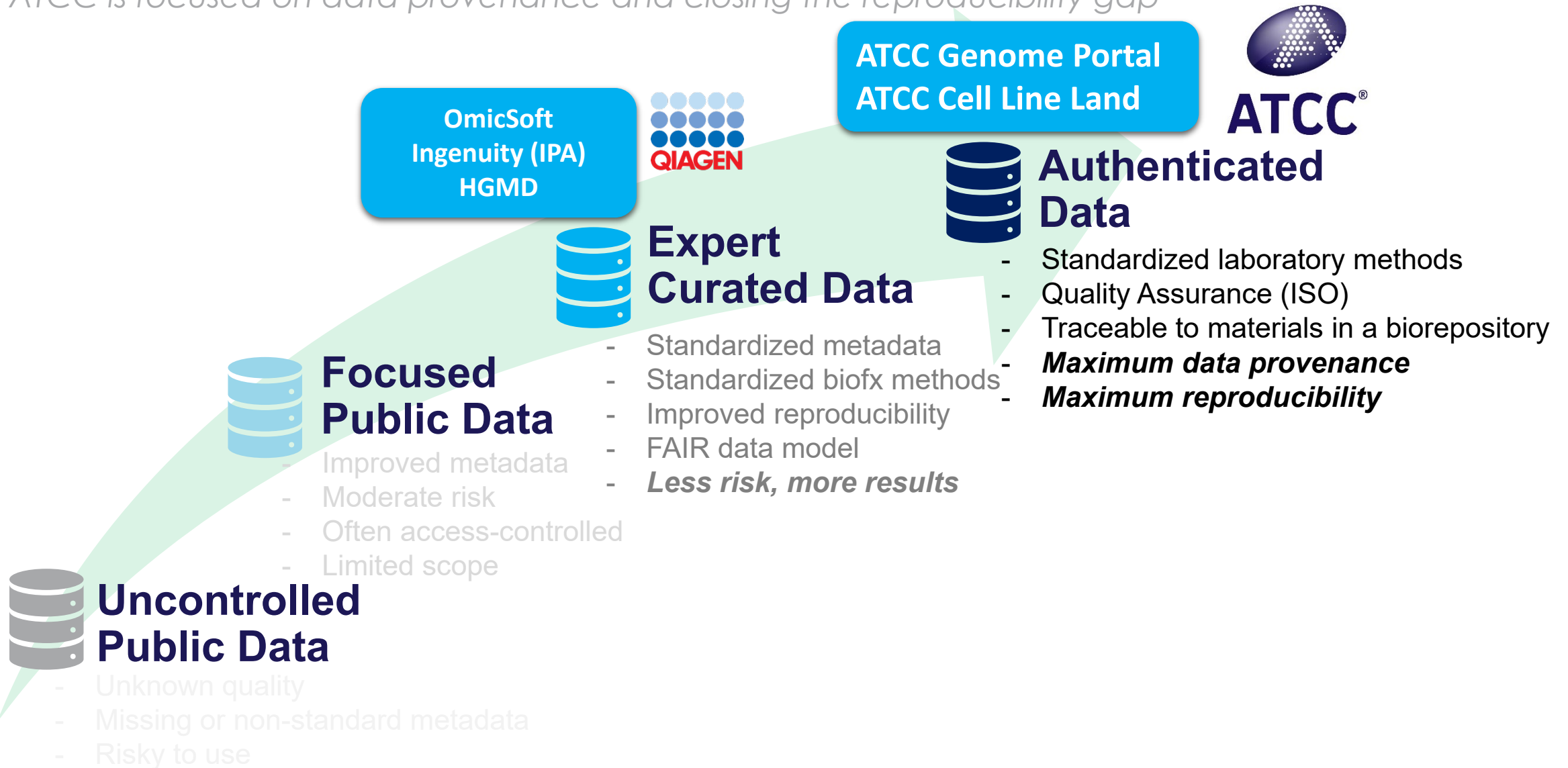
Authenticated genomics data at ATCC

ATCC is focused on data provenance and closing the reproducibility gap



Authenticated genomics data at ATCC

ATCC is focused on data provenance and closing the reproducibility gap





The ATCC Genome Portal

Tackling the reproducibility gap in microbial genomics

ATCC Genome Portal

The ATCC Genome Portal is a cloud-based platform that enables users to easily browse genomic data and metadata by simply logging into the portal



Download whole-genome sequences and annotations of ATCC materials



Search for nucleotide sequences or genes within genomes



View genome assembly metadata and quality metrics

genomes.atcc.org

2,522 Authenticated Reference Genomes

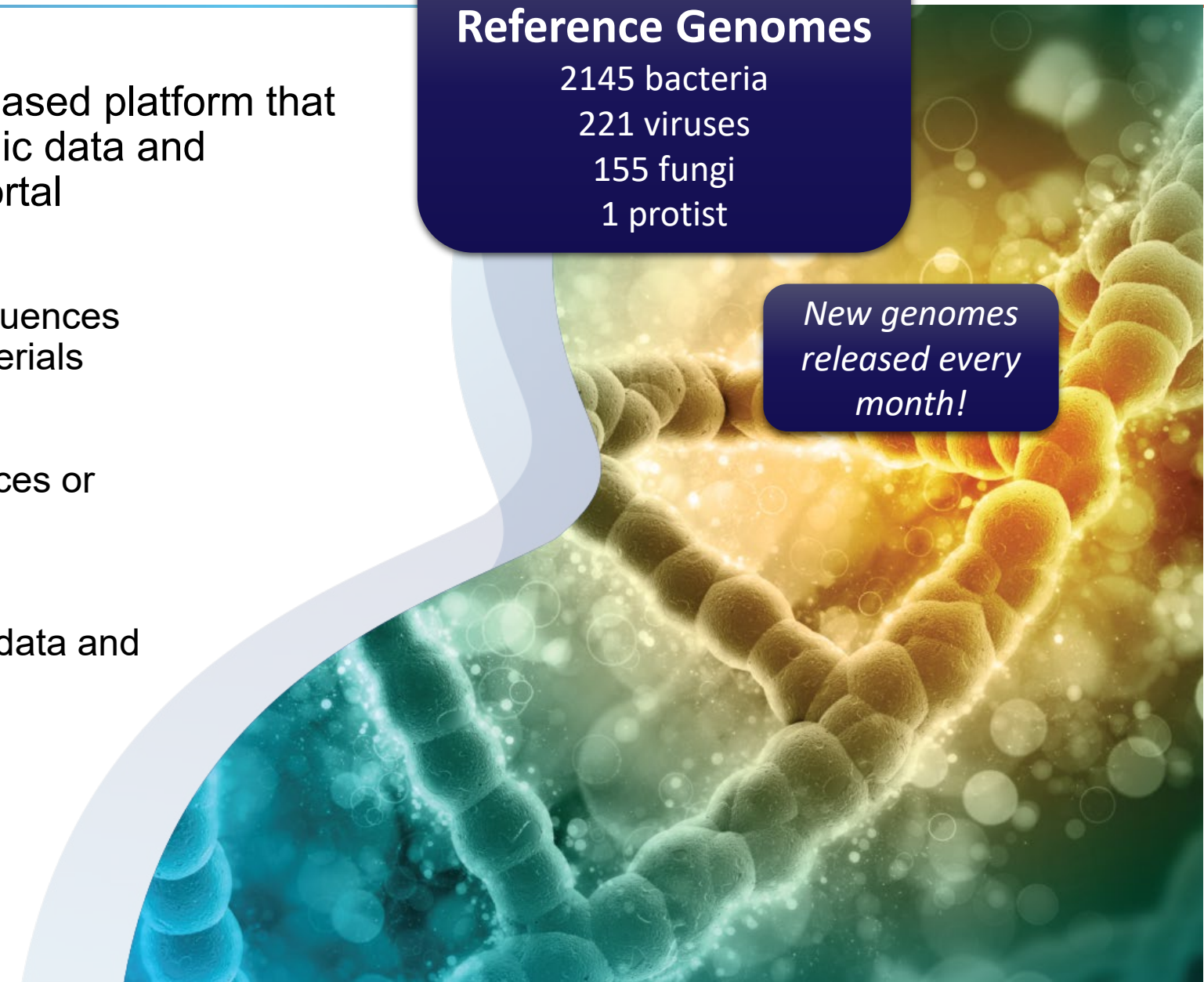
2145 bacteria

221 viruses

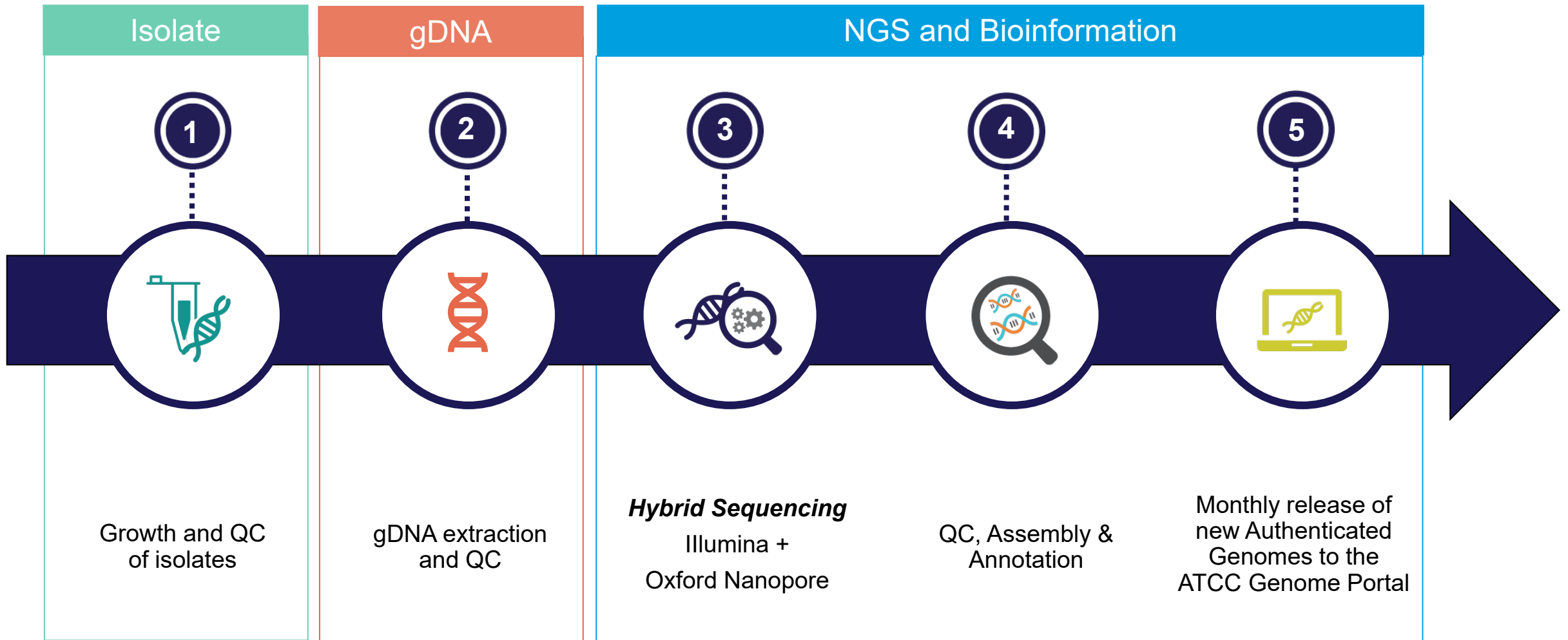
155 fungi

1 protist

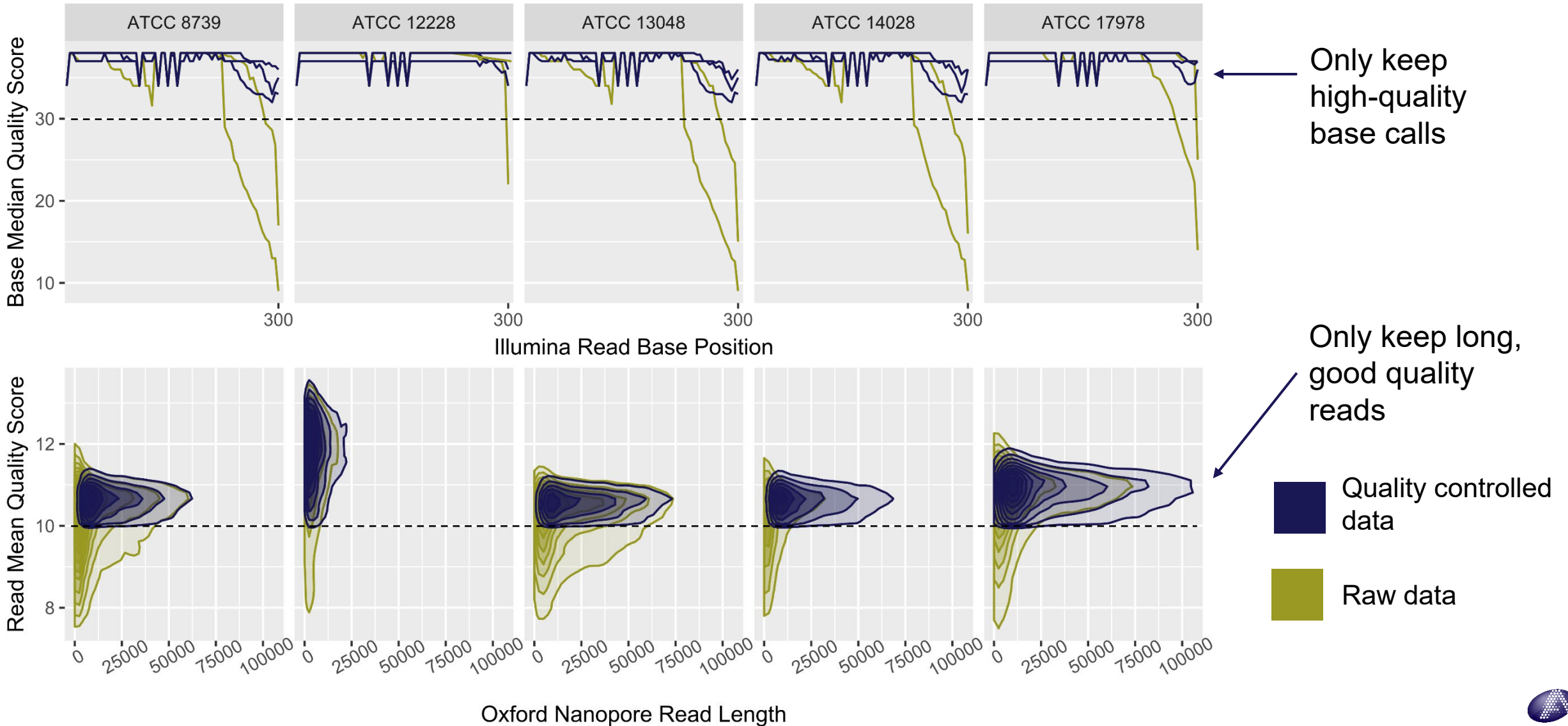
New genomes released every month!



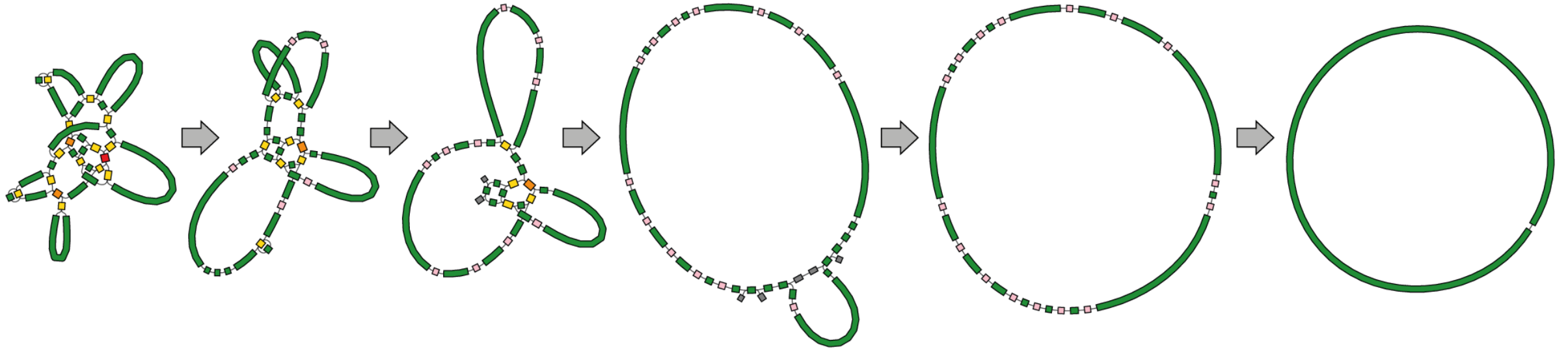
Authenticated physical material coupled with reference-quality genome sequences



Sequencing QC – Read trimming/filtering



Hybrid genome assembly



**Illumina-only
genome
assembly**
150 bp reads

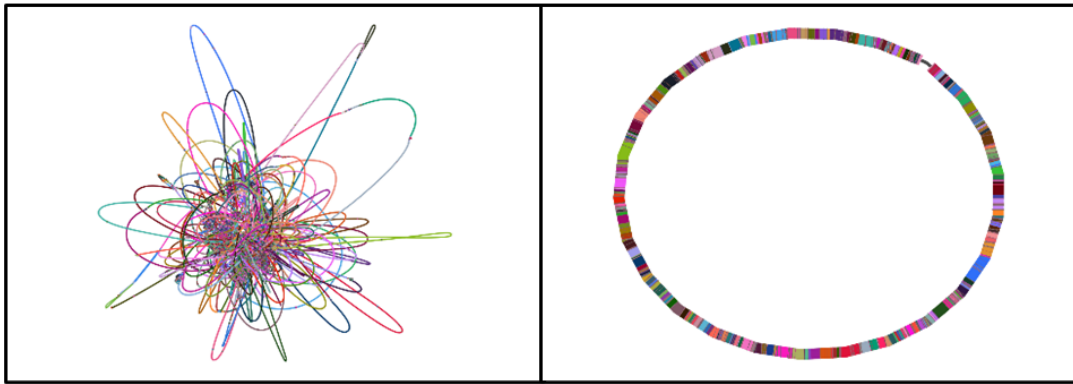
Long reads mapped to a tangled region creates a resolved bridge
Successively applying bridges resolves the structure of the genome

**Completed
hybrid assembly**

Advantage of hybrid assemblies

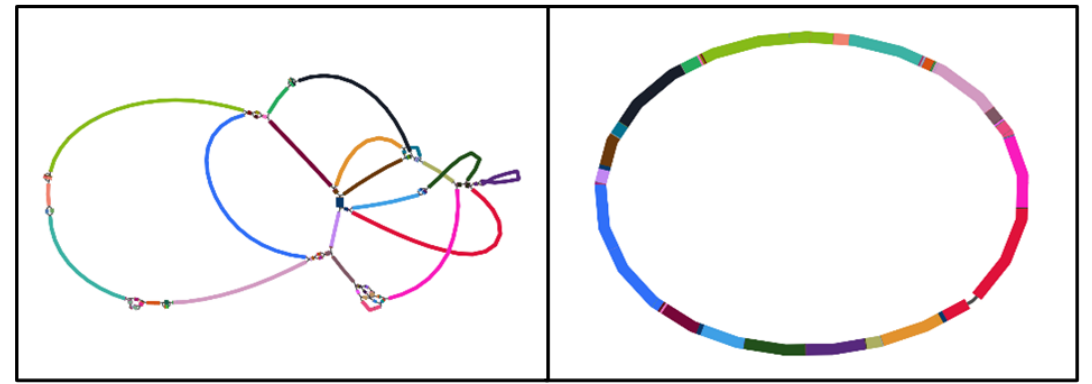
Illumina-only assembly Hybrid assembly

Neisseria meningitidis (ATCC® 53417™)

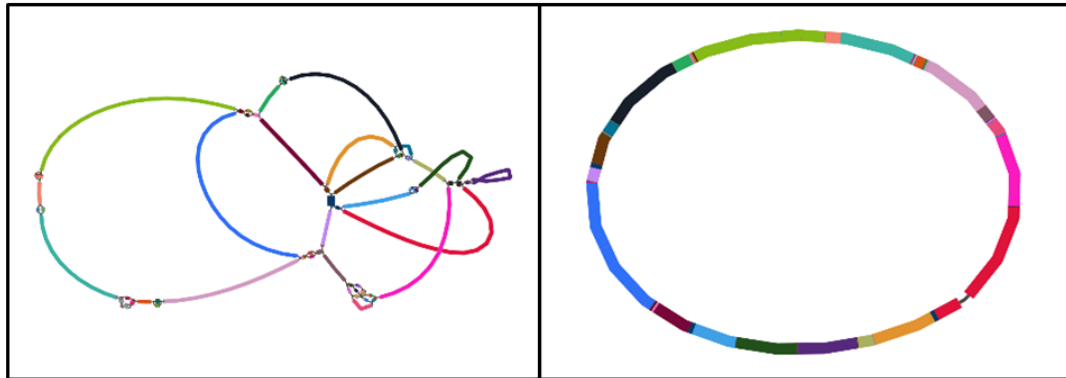


Illumina-only assembly Hybrid assembly

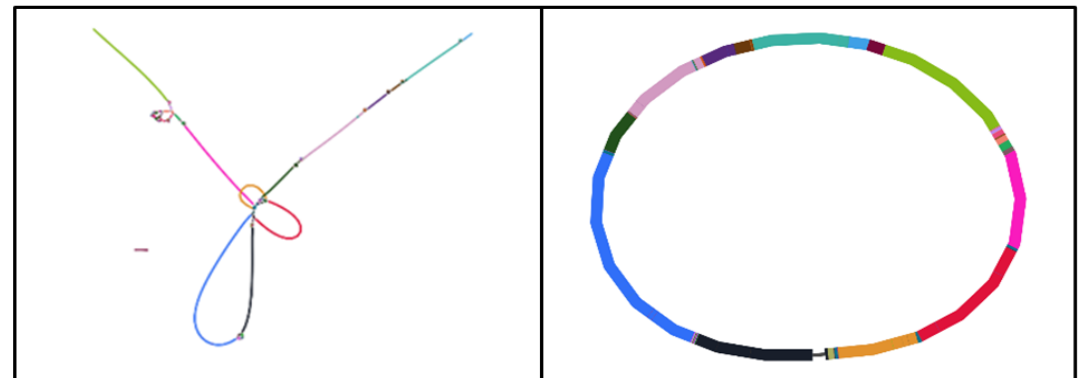
Campylobacter jejuni subsp. *jejuni* (ATCC® 43446™)



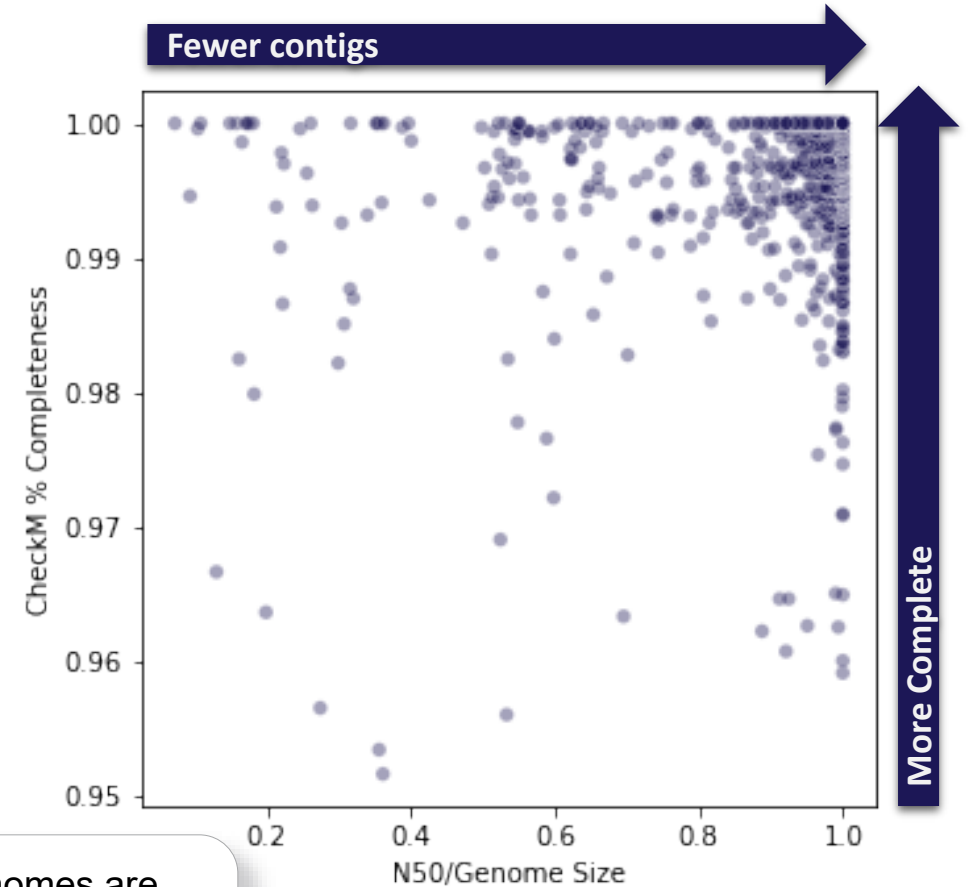
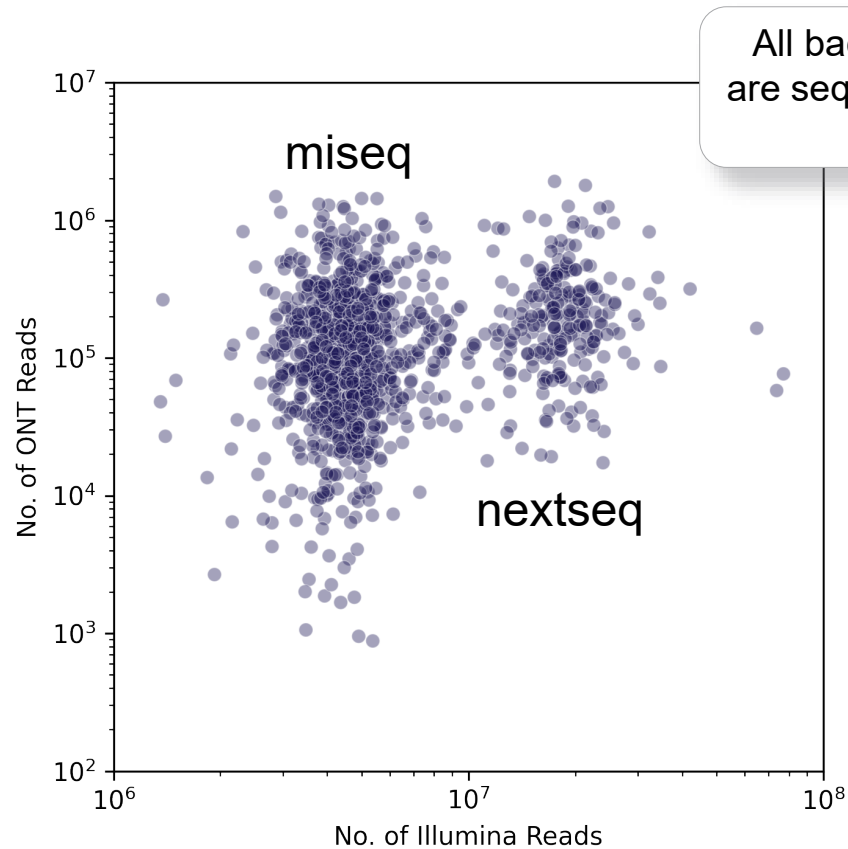
Campylobacter jejuni subsp. *jejuni* (ATCC® 43446™)



Streptococcus gordonii (ATCC® 35105™)



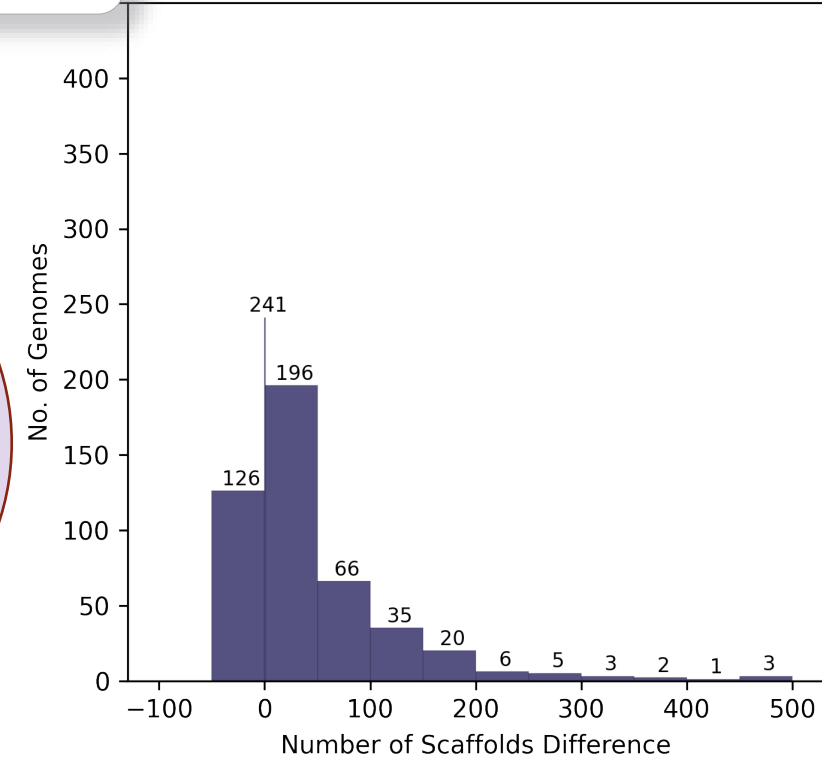
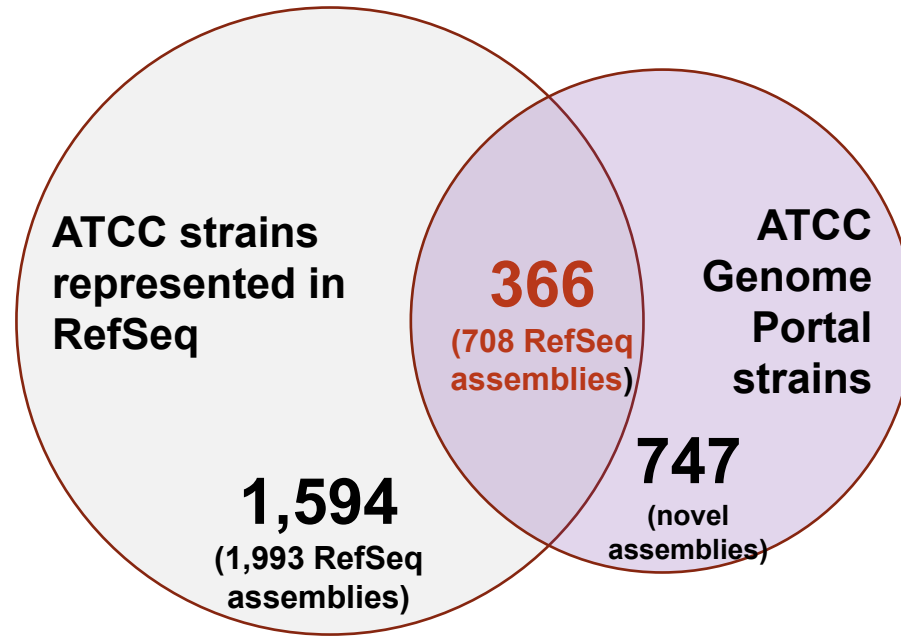
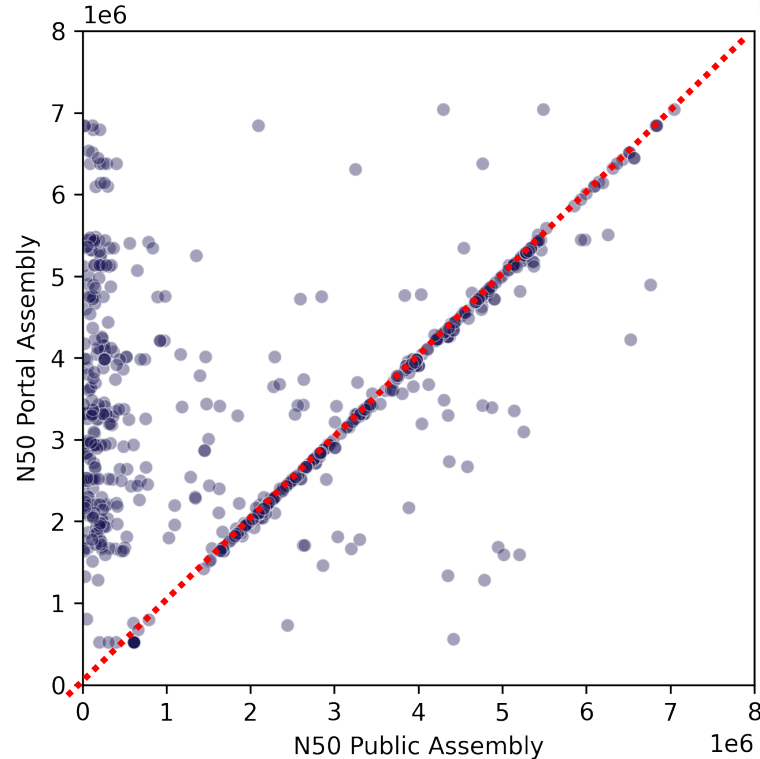
Quality of ATCC Genome Portal assemblies



All genomes are required to be at least 95% complete (CheckM/Busco)

Comparison of ATCC vs. RefSeq bacterial assemblies

>98% of our assemblies are more complete and of higher quality than RefSeq



Yarmosh DA et. al. *Comparative Analysis and Data Provenance for 1,113 Bacterial Genome Assemblies*. *mSphere* **2022**, e00077-22. <https://doi.org/10.1128/msphere.00077-22>.



ATCC Cell Line Land

A partnership with QIAGEN Digital Insights

ATCC cell biology collection

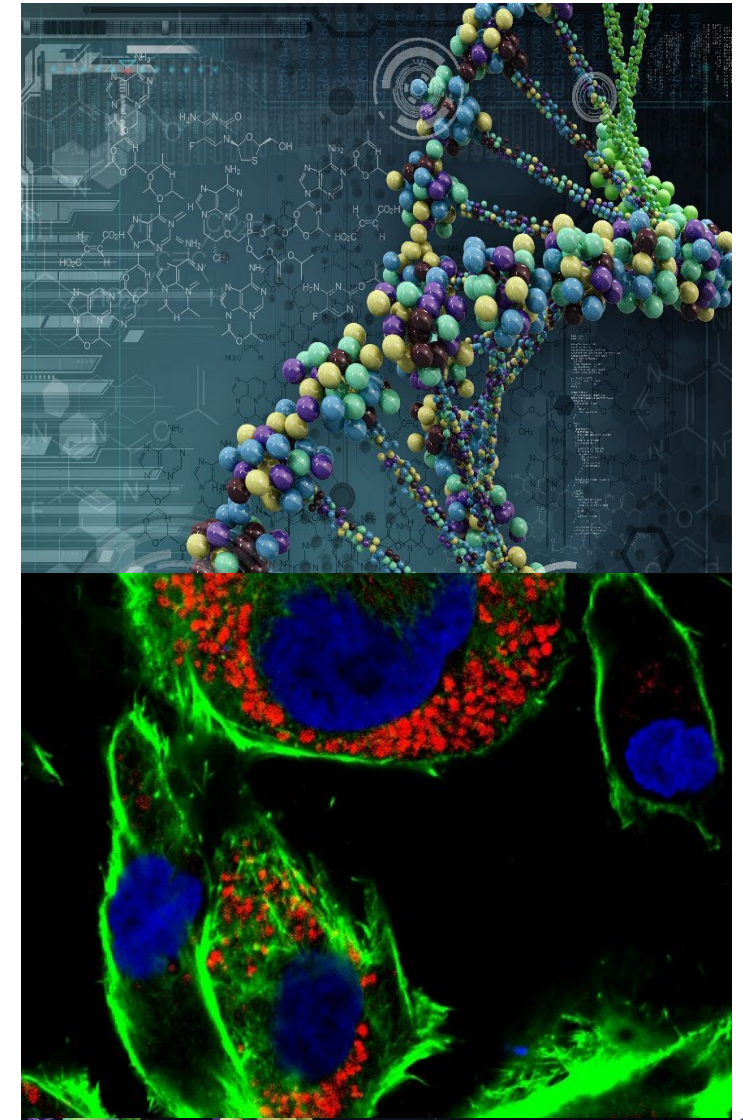
ATCC has **3,000+ authenticated** mammalian cell lines, genetic engineered cell lines, primary cells, stem cells, iPSCs, hTERT-immortalized cells, and tumor organoids representing various species, cell types, tissues origins, and diseases.

70+
Species

100+
Cell types

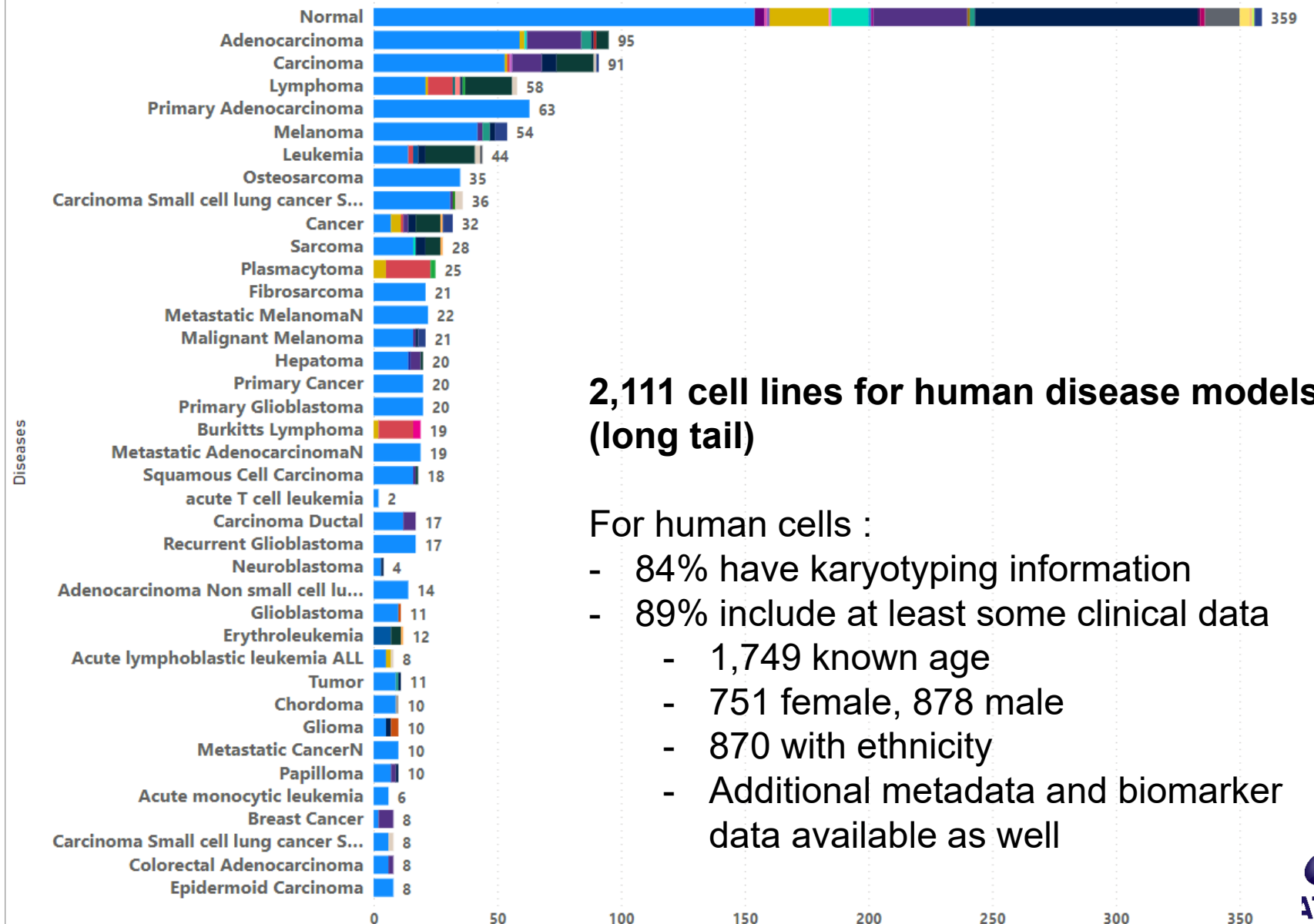
100+
Tissue types

400+
Diseases types



ATCC Cell Biology Collection (by disease type)

Cell line models for over 400 disease types



2,111 cell lines for human disease models (long tail)

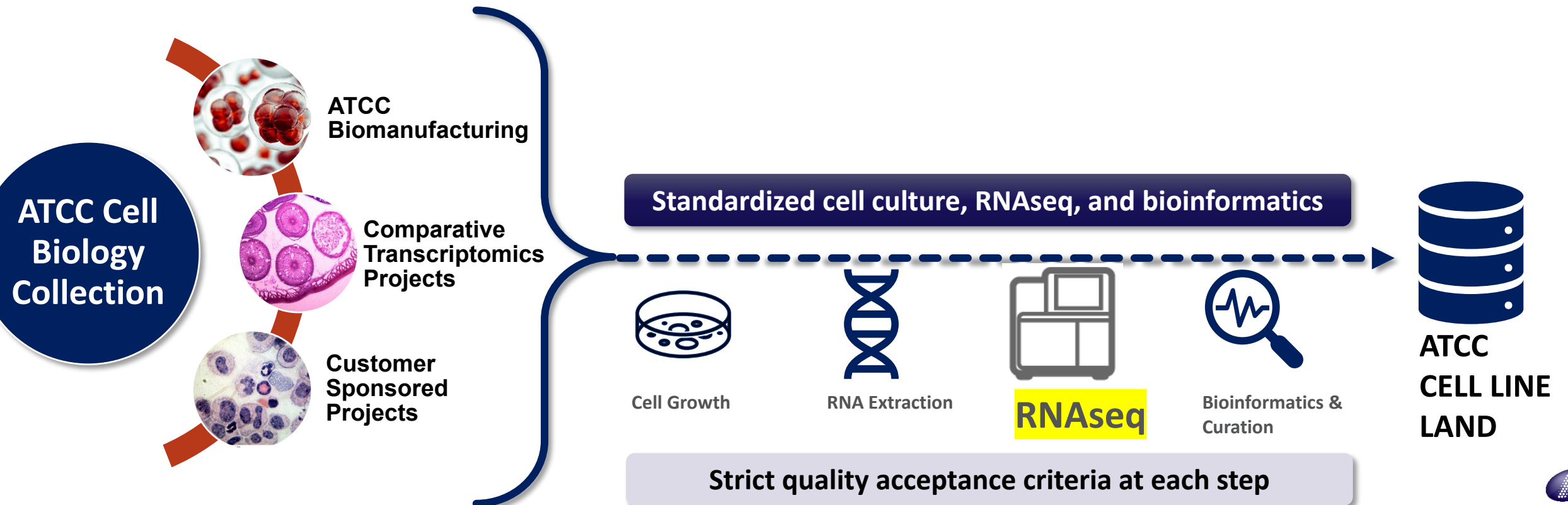
For human cells :

- 84% have karyotyping information
- 89% include at least some clinical data
 - 1,749 known age
 - 751 female, 878 male
 - 870 with ethnicity
- Additional metadata and biomarker data available as well

ATCC Cell Line Land

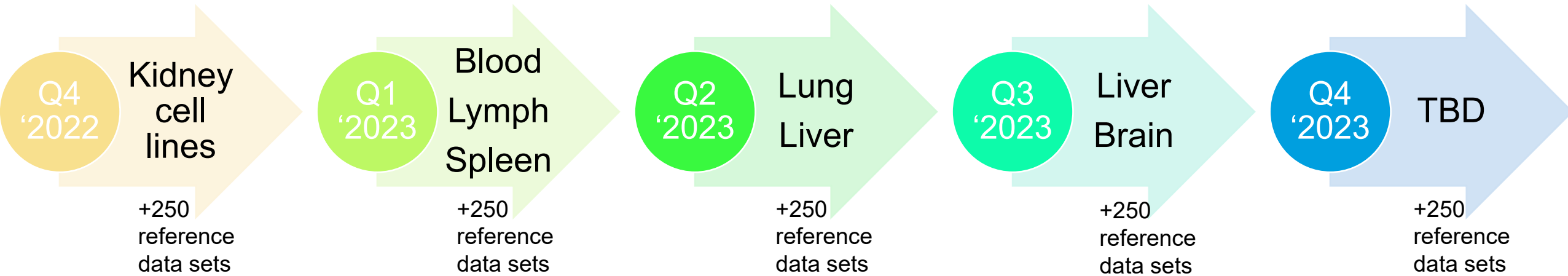
KEY FEATURES

1. Repository of ***authenticated 'omics data traceable to physical materials***
2. Data production, curation, and analysis uniformly standardized
3. Enables the highest level of **scientific reproducibility**
4. End-to-end **data provenance**



ATCC Cell Line Land

A partnership with QIAGEN Digital Insights



- Current road-map for data production is subject to change
- Based on customer feedback
- **1,000+ traceable, authenticated RNAseq datasets per year**

ATCC Cell Line Land

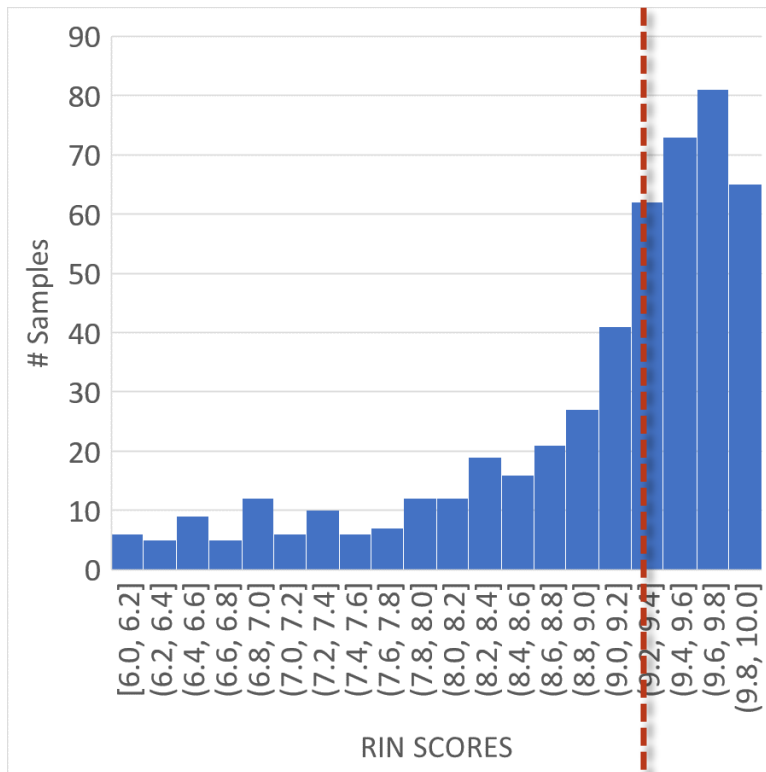
A partnership with QIAGEN Digital Insights



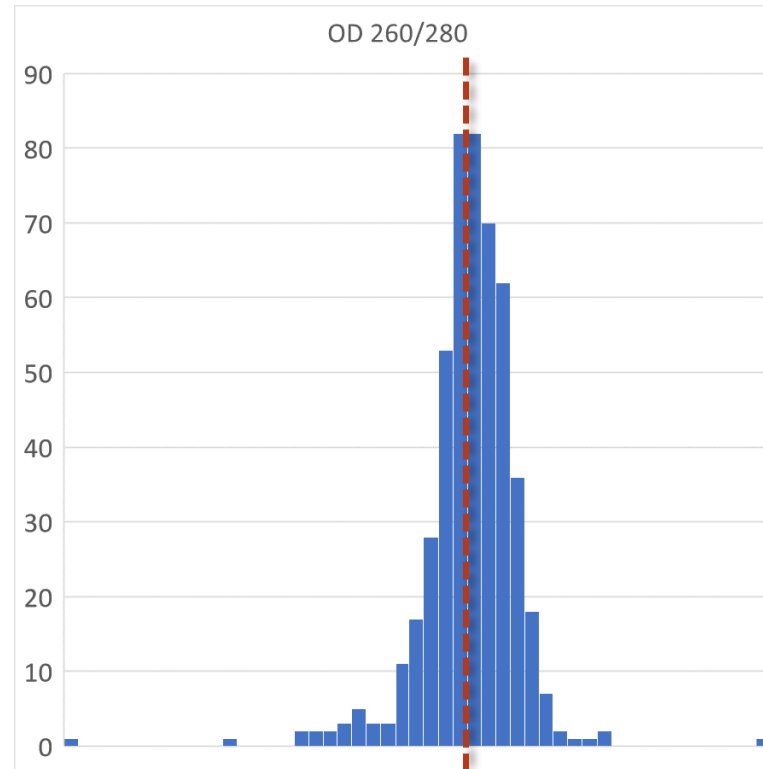
- Quality-controlled data from ATCC cell lines
- Over 1000 new datasets added each year, released quarterly
- Careful metadata curation with controlled vocabulary
- Reprocessed and normalized RNAseq expression
- Metadata include standard culture conditions, extraction protocols, sample preparation, and library preparation
- Data grows based on what you, as a researcher, need most:
 - Our team takes your requests to prioritize the cell lines you want added to our ATCC Cell Line Land collection, as well as the type of experimental data you want curated

ATCC Cell Line Land

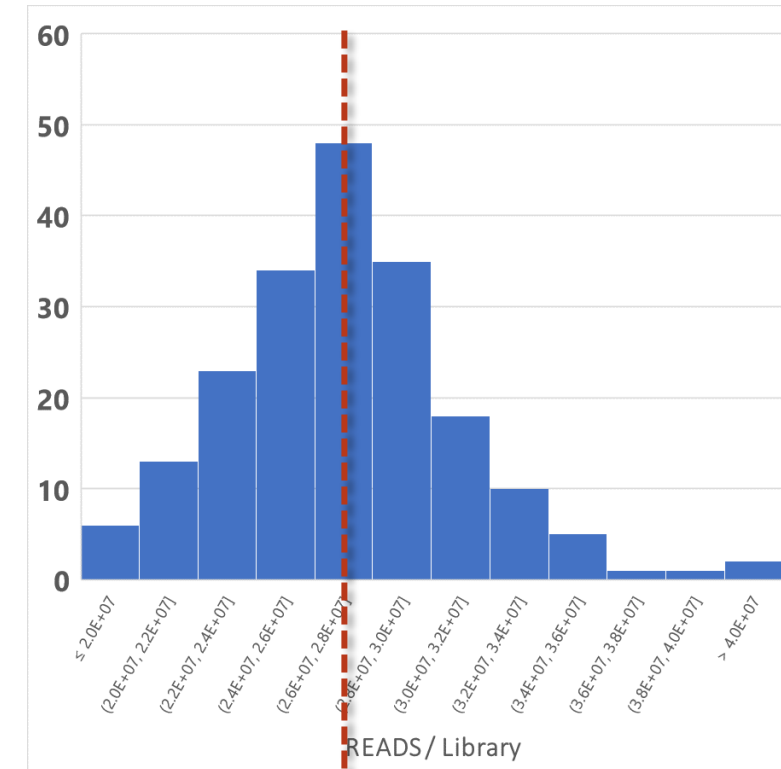
A partnership with QIAGEN Digital Insights



**9.3 median
RIN score**



**2.071 median
OD260/280**



**26.9M reads per
library (median)**

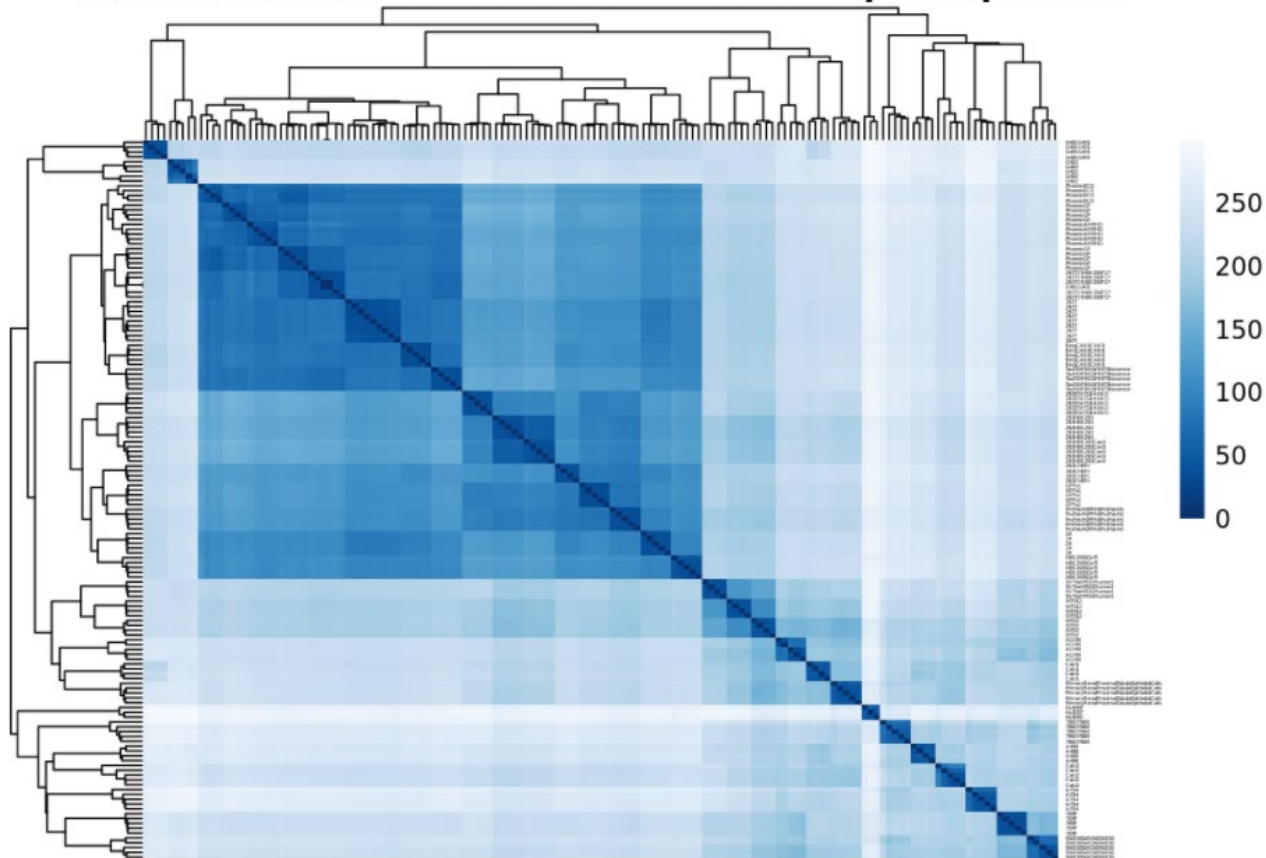


ATCC®

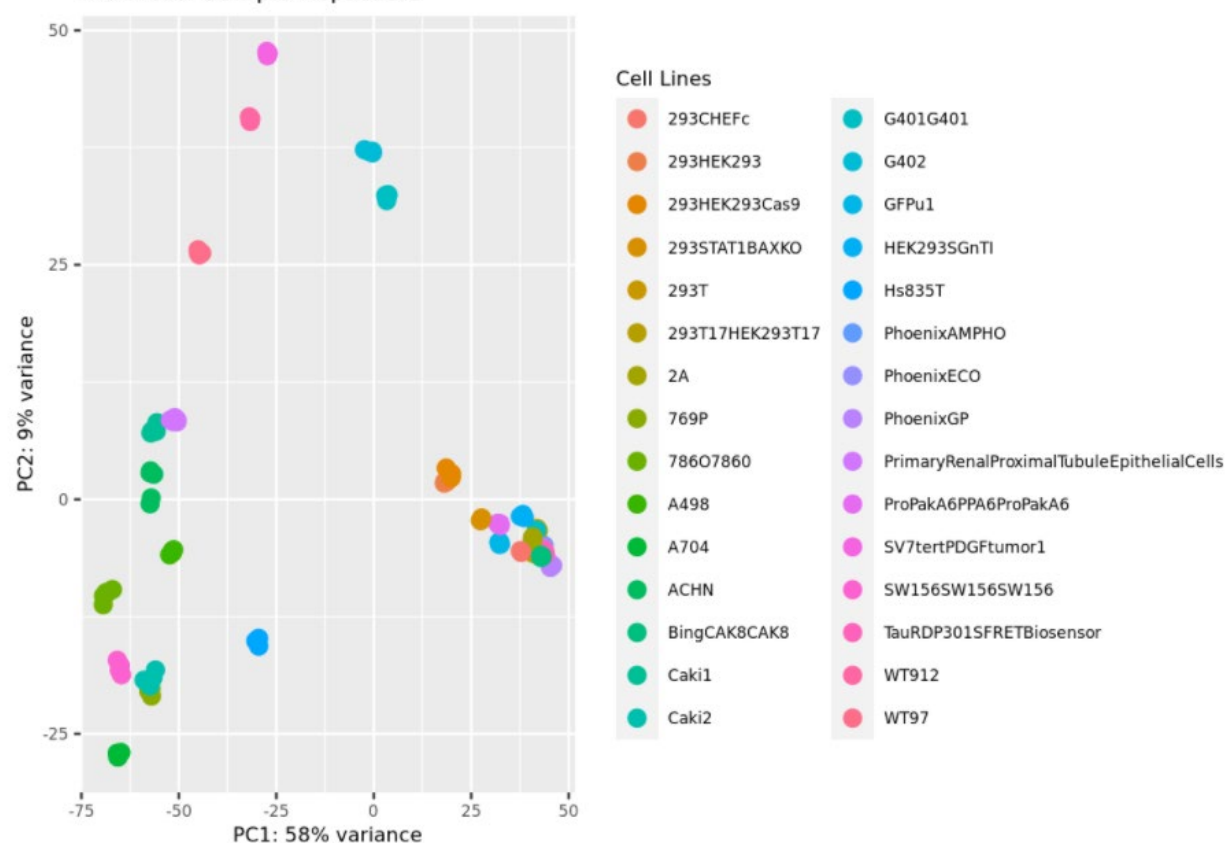
ATCC Cell Line Land – Example (kidney cell lines)

A partnership with QIAGEN Digital Insights

Normalized Read Counts of Individual Sample Replicates



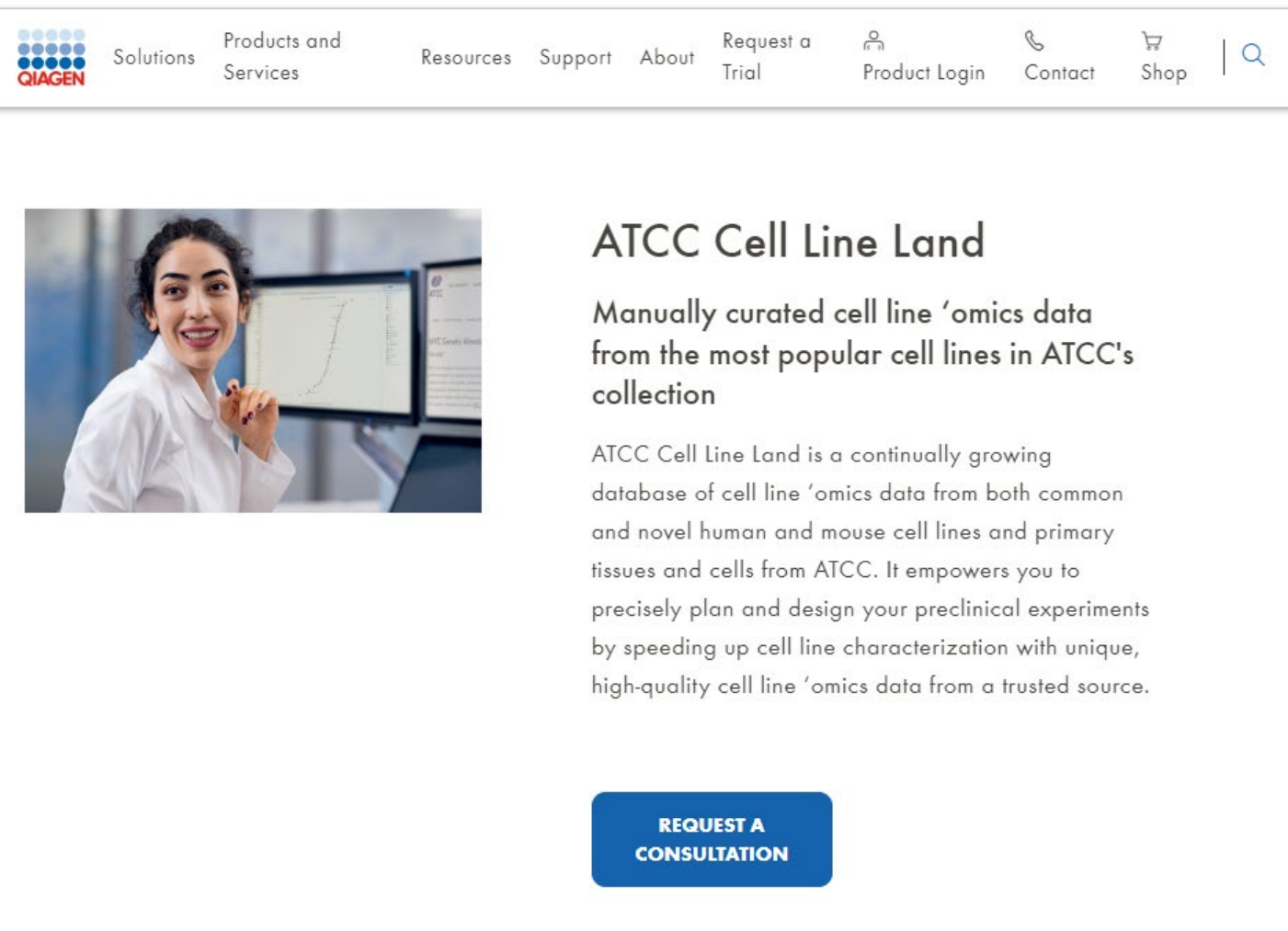
Individual Sample Replicates



Full data for over 60 kidney cell lines will be presented at the American Society of Cell Biology (December 2022)

ATCC Cell Line Land – Available through QIAGEN

A partnership with QIAGEN Digital Insights



ATCC Cell Line Land

Manually curated cell line 'omics data from the most popular cell lines in ATCC's collection

ATCC Cell Line Land is a continually growing database of cell line 'omics data from both common and novel human and mouse cell lines and primary tissues and cells from ATCC. It empowers you to precisely plan and design your preclinical experiments by speeding up cell line characterization with unique, high-quality cell line 'omics data from a trusted source.


REQUEST A CONSULTATION

Currently includes
Authenticated RNAseq Data for over 200 ATCC cell lines.

<https://digitalinsights.qiagen.com/atcc-cell-line-land/>



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QIAGEN Digital Insights

One Codex



Thank you!

ATCC Genome Portal

<https://genomes.atcc.org>

ATCC Cell Line Land

<https://digitalinsights.qiagen.com/atcc-cell-line-land/>